



## UvA-DARE (Digital Academic Repository)

### Predictability of bacterial activity and denitrification in aquatic sediments with continuous measurements of redox potential.

Hunting, E.R.; van der Geest, H.G.

**DOI**

[10.1007/BF03326241](https://doi.org/10.1007/BF03326241)

**Publication date**

2011

**Document Version**

Final published version

**Published in**

International Journal of Environmental Science and Technology

[Link to publication](#)

**Citation for published version (APA):**

Hunting, E. R., & van der Geest, H. G. (2011). Predictability of bacterial activity and denitrification in aquatic sediments with continuous measurements of redox potential. *International Journal of Environmental Science and Technology*, 8(3), 553-560. <https://doi.org/10.1007/BF03326241>

**General rights**

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

**Disclaimer/Complaints regulations**

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.

*UvA-DARE is a service provided by the library of the University of Amsterdam (<https://dare.uva.nl>)*

## Predictability of bacterial activity and denitrification in aquatic sediments with continuous measurements of redox potential

\*E. R. Hunting; H. G. van der Geest

*Institute for Biodiversity and Ecosystem Dynamics, Aquatic Ecology and Ecotoxicology, University of Amsterdam, Amsterdam, the Netherlands*

Received 25 August 2010; revised 20 October 2010; accepted 27 December 2010; available online 1 June 2011

**ABSTRACT:** Redox potential has been adopted as a qualitative parameter for interpreting solubility changes of nutrients and contaminants and the biological activity within wetland systems for several decades. The majority of studies considering the redox geochemistry in sediments used measurements of bulked material and single point measurement of biogeochemical parameters for interpretation, yet it remains questionable whether this information is reliable for environments that are very dynamic, such as wetlands. In this study it is evaluated whether variations in redox potential reflect dynamics of denitrification and overall bacterial respiration using continuous measurements of redox potential in time-series experiments in laboratory microcosms, in which the biogeochemical variation was enhanced by bioturbation. The results presented here suggest that measurements of redox potential have predictive potential in approximating rates of denitrification and overall bacterial respiration in aquatic sediments. The data clearly suggest that, while sediment bulk measurements and measurements of single profiles of redox potential, denitrification and bacterial activity often fail to provide ecological relevant information in dynamic systems, measurements of spatial and temporal redox potential profiles provide a useful parameter that reflects biogeochemical processes and functioning of sediments.

**Keywords:** *Biogeochemistry; Bioturbation; Ecosystem functioning; Tubifex; Wetland dynamics*

### INTRODUCTION

The redox potential (Eh) provides an electrochemical measure of a system's electric potential standardized as relative to a hydrogen electrode. Eh electrodes are alluring due to the rapidity and inexpensiveness of measurements and they are adopted as a qualitative parameter for interpreting solubility changes of nutrients and contaminants and the biological activity within wetland systems for already several decades (Jackson, *et al.*, 1993; Teasdale *et al.*, 1998; Biati and Karbassi, 2010; Biati *et al.*, 2010; Mohiuddin *et al.*, 2010; Sekabira *et al.*, 2010).

However, Eh is formally defined by the Nernst equation and thereby carries an erroneous assumption that redox reactions in a system are reversible, complete and in equilibrium, while many reactions in nature are neither complete or reversible. The incomplete and irreversible nature of many natural redox reactions in wetland soils and sediments may result in the

occurrence of multiple redox 'equilibria' and the measurement of a mixed potential. Different reactions are therefore expected to contribute differently to the signal, i.e. irreversible reactions fail to give a potential signal, while reactions that are kinetically slow may provide weak and unstable potentials, whereas other reactions may participate effectively in the measured signals (Grenthe *et al.*, 1992).

The interpretability of Eh relies on rigorous theoretical considerations and only a limited number of studies examined actual processes underlying Eh. Although the complexity of natural wetland systems generally flocculates predictability and interpretability, several studies were able to relate Eh to concentrations of terminal electron acceptors involved in electron transfer activity, in which Eh correlated well to concentrations of the redox species Mn (II) and Fe (II) and dominant redox processes (Brünger, 1982; Gao *et al.*, 2002; Mansfeldt, 2004; Abdel-Ghani *et al.*, 2009). However, the majority of studies considering the

✉ \*Corresponding Author Email: [e.r.hunting@uva.nl](mailto:e.r.hunting@uva.nl)  
Tel./ Fax: +31205257659

redox geochemistry in soils and sediments used measurements of bulked material and single point measurement without paying attention to temporal and spatial variability. Although this may provide information on underlying mechanisms in stable laboratory microcosms that are in quasi steady state conditions, it remains questionable whether this information is reliable for environments in which components that govern biogeochemical processes are highly variable, such as wetlands.

Continuous monitoring of redox conditions with the use of permanently installed redox electrodes has demonstrated that in situ redox conditions may show large spatial and temporal fluctuations in a variety of habitats, where Eh may fluctuate from -400 to 100 mV over several days and may vary up to 200 mV in daily cycles (Vorenhout *et al.*, 2004; 2011). This variability is easily overlooked in measurements of bulked soils and sediments and single profiles and it remains illusive whether parameters related to redox potential reflect similar spatiotemporal variability and whether this variability provides ecological relevant information on geochemical and bacterial processes in natural, dynamic environments.

This study sets out to determine whether continuous measurements of Eh provide a key correlate to geochemical and bacterial processes in dynamic aquatic sediments, in which denitrification is under focus, since denitrification is a very active process and also a dominant, very important sink for mineralized nitrogen in Lake ecosystems and constructed wetlands (Saunders and Kalf, 2001). To this purpose, continuous redox potentials, rates of denitrification and bacterial metabolic activity were measured in time-series in microcosms containing sediment collected from Lake Drontermeer, the Netherlands, in 2007, in which spatial and temporal variability was induced by the introduction of the aquatic oligochaete *Tubifex* spp.

## MATERIALS AND METHODS

### *Microcosm*

Organic rich sediment was collected from Lake Drontermeer, the Netherlands (52°58'N, 5°50'E) for natural bacterial communities and organic matter. The collected sediment was frozen in order to kill infauna and thawed to allow for bacterial re-growth. This approach generally does not have major impacts on bacterial community structures (Morly *et al.*, 2005; Männistö *et al.*, 2009). Natural sediments were

subsequently mixed with clean river sand (ignited quartz; grain size 0.5 – 1.0 mm; ratio 1:5), homogenized and oxygenated prior to experimentation.

Microcosm consisted of a circular polyvinyl chloride core (110 mm diameter, 250 mm height) with 7 gastight connectors at different depths for pore water probes modified after Vink (2002) and bacterial probes as described below. The pore water probes consisted of a semi permeable polyethersulfone polymer of which its mesh size (~0.1 µm) allows sampling of dissolved compounds in pore water but is impermeable to sediment particles and bacteria (Fig. 1). The probes were flushed with nitrogen gas before use to prevent oxidation processes during sampling. Bacterial probes consisted of a Nuclepore polycarbonate film mounted on polymer HDPE tubes by polymerization of vinyl chloride dissolved in 1,1,2-trichloroethane (Fig. 1). The membrane acts as semi-permeable membrane (mesh size 5 µm) to sample porewater containing bacterial cells while retaining particles and floccules. Sterile 3 mL syringes were connected to the probes for sample extraction. Cores were incubated at 20 °C under continuous light conditions (40 µmol photons/m<sup>2</sup>/s).

Overlying water was kept at 30 mm above sediment water interface with continuous inflow of oxygen saturated water (Dutch Standard Water, DSW, a standardized synthetic analogue of common Dutch surface waters, containing 200 mg CaCl<sub>2</sub>·2H<sub>2</sub>O, 180 mg MgSO<sub>4</sub>·7H<sub>2</sub>O, 100 mg NaHCO<sub>3</sub>, and 20 mg KHCO<sub>3</sub> per liter demineralized water: pH 8.1; hardness 210 mg/L CaCO<sub>3</sub>; alkalinity 1.2 meq/L; flow rate 20 mL/h). Overlying water was oxygenated, in which oxygen levels were generally stable at 70 % saturation over the course of the experiment.

### *Experimental procedure*

Time series experiments were performed to link short term variability in Eh to different biogeochemical processes. The experiment consisted of two parts. First, the replicate (n=4) cores were allowed to stratify for 48 h, after which *Tubifex* spp. (approximately 6500 individuals/m<sup>2</sup>) were added to rework the sediment for an additional 48 h. Measurements of parameters in cores commenced immediately after homogenization and continued towards the end of the experimental runs at fixed time-intervals over a period of 4 d. Eh was measured every 15 min and oxygen profiles of the cores were measured twice per day. Pore water was

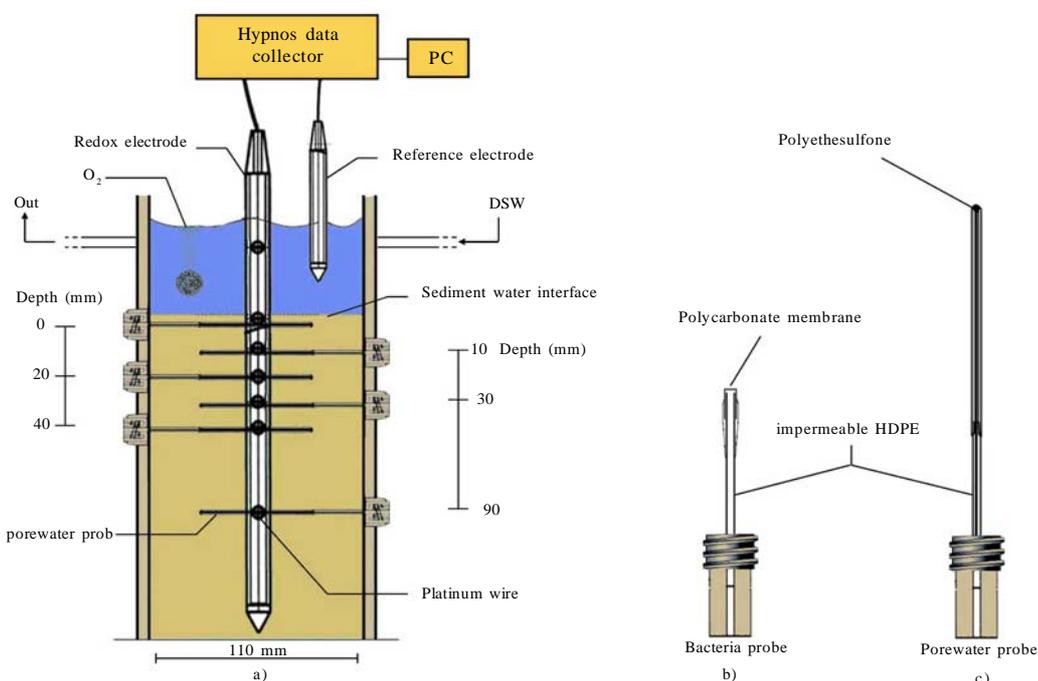


Fig. 1: Conceptual impression of the experimental set up. All experiments were performed in a) a circular polyvinyl chloride core that allows for continuous monitoring of undisturbed sediment. The core has 7 gastight connectors for b) polycarbonate microbial probes described in this study and c) porewater probes modified after by Vink (2002). Eh was measured at various depths using permanently installed platinum electrodes and a calomel reference electrode connected to the datalogger Hypnos 2.0 (Vorenhout *et al.*, 2004). See text for further details

simultaneously sampled alongside the platinum electrodes 1-2 times per day and immediately analyzed for rates of denitrification and overall bacterial respiration. Measurements of biogeochemical processes in this study are restricted to denitrification since denitrification is a very active process, and a dominant, very important sink for mineralized nitrogen in Lake ecosystems (Saunders and Kalff, 2001; Nouri *et al.*, 2010).

#### Analytical techniques

Dissolved oxygen concentrations were measured with an oxygen microsensor (Unisense OX25) connected to a picoammeter (Unisense, PA2000). Eh was measured at various depths in the sediment (0 mm; 10 mm; 20 mm; 30 mm; 40 mm; 90 mm) using permanently installed platinum electrodes and a calomel reference electrode connected to Hypnos 2.0 datalogger (Fig. 1) (Accuracy of measurements  $\pm 10$  mV). Construction of electrodes and datalogger is described in detail

elsewhere (Vorenhout *et al.*, 2004). Eh was corrected for standard hydrogen electrode by adding 245 mV. Concentrations of nitrate in porewater (200  $\mu$ L porewater sample) were colorimetrically determined (Shimadzu, 1601-UV) using Griess reagents (Griess, 1864). Decreases in concentrations of nitrate ( $\text{NO}_3^-$ ) over a defined time interval provide rough estimations of rates of denitrification (Anderson, 1977). It should be noted, however, this approach is used to address the predictive potential of Eh and accurate estimations of denitrifications rates were beyond the scope of this study, provided that the approach is very likely to overestimate the actual process (Seitzinger, 1988). Overall respiratory capacity of the bacterial community was approximated by dehydrogenase enzyme activity via the reduction of 2-(p-iodophenyl)-3-(p-nitrophenyl)-5-phenyl tetrazolium chloride (INT) to formazan (INTF). INT successfully competes with  $\text{NADH}^+$  and  $\text{NADPH}^+$  and INT reduction proved an

### Predictability of continuous redox potential measurements

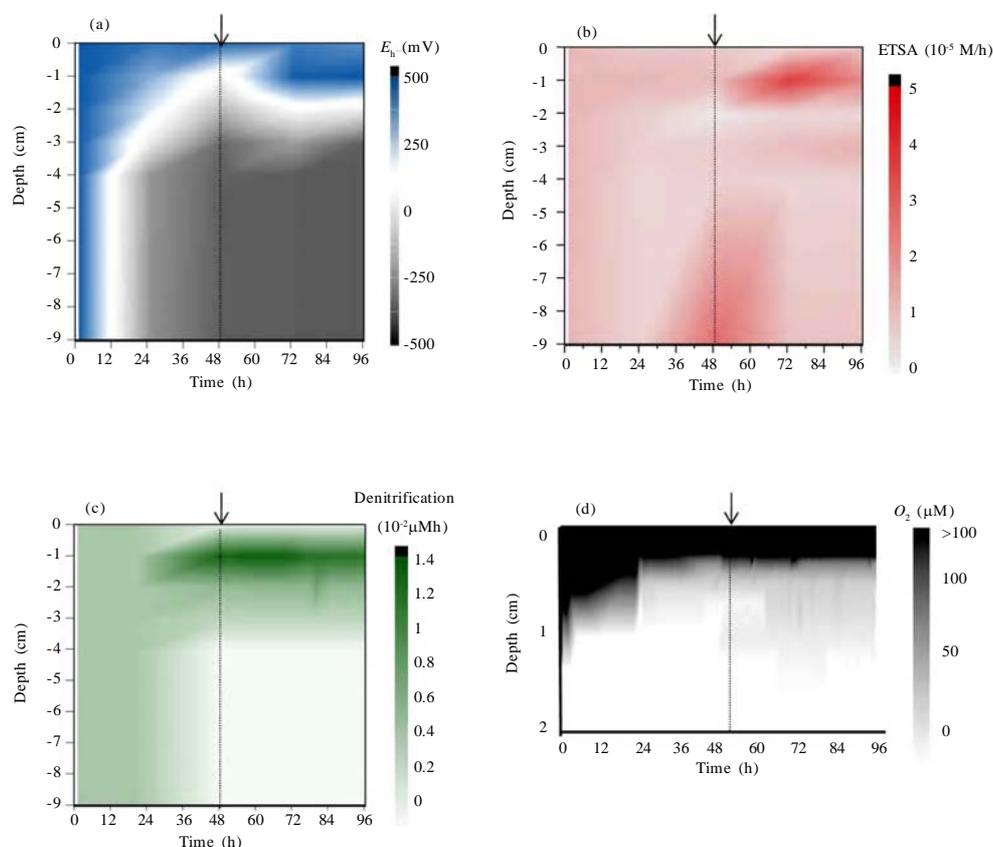


Fig. 2: Spatio-temporal measurements of redox geochemical characteristics of the experimental microcosm. Redox geochemical parameters include a) Eh; b) bacterial activity, ETSA; c) denitrification rates; and d) oxygen concentrations,  $O_2$ . Contour plots were obtained by linear interpolation (Deltagraph 5.0) from the averages of 4 replicates and indicate changes of parameters in depth over a period of 96 h. Arrow indicates addition of *Tubifex* spp.

accurate assay of Electron transfer system activities (ETSA) under both anaerobic and aerobic conditions (Smith and Mcfeters, 1997; Bhupathiraju *et al.*, 1999; Rozzi and Remigi, 2004; Hunting *et al.*, 2010). Bacterial cells were simultaneously sampled alongside the platinum electrodes (400  $\mu$ L porewater sample). The samples were vortexed and incubated at 20  $^{\circ}$ C for 30 min. Bacterial cell integrity was subsequently disrupted by 5 min sonication at room temperature (Branson, 1510). An aqueous INT solution (200  $\mu$ L;  $6.2 \cdot 10^{-2}$  M) was added and the samples were mixed and incubated for 1 hour at 20  $^{\circ}$ C under dark room conditions. Enzymatic activity was stopped by adding 500  $\mu$ L mL of acetone and measured spectrophotometrically at 490 nm (Shimadzu, 1601-UV). An INT formazan standard was used for calibration, in which two protons equal the transfer of a single electron and samples were

corrected for abiotic reduction of INT by subtracting formalin treated replicates.

## RESULTS AND DISCUSSION

Homogenized sediments generally reached quasi steady state redox condition after 36 to 48 h (Fig. 2). Redox geochemical characteristics are provided in figure 2a-d. Contourplots represent the average of four replicates and show the changes in the parameters in depth over a period of 96 h. Eh ranged -300 to 500 mV at all depth measured in the core (Fig. 2a). Oxygen generally penetrated to 10 mm depth. Oxygen concentrations were highest at the sediment surface (237  $\mu$ M) and rapidly decreased after a few millimeters (Fig. 2d). Rates of denitrification ranged 0 –  $1.35 \cdot 10^{-2}$   $\mu$ M/h and were highest at 1 cm depth at steady state conditions (Fig. 2c). Overall bacterial activity (ETSA)

ranged from 0 to  $5 \cdot 10^{-5}$  M/h. Bacterial activity was generally most pronounced at 1 cm depth and often decreased with increasing depth in deeper layers of the sediment, yet was occasionally enhanced at 4 cm and 9 cm depth in some of the replicates (data of replicates not shown), which is visible as slightly increased colour intensities at these depths in the contourplots of the averaged bacterial activity (Fig. 2b). Bacterial cells were simultaneously sampled alongside the platinum electrodes (400  $\mu$ L porewater sample). As a consequence, the activity of the bacterial fraction sorbed to sediment particles is overlooked. However, it is consistently reported that the sorbed fraction fails to contribute significantly to measurements of bacterial activity (Bright and Fletcher, 1983; Bonin *et al.*, 2001; Yousefi Kebria *et al.*, 2009; Zhang *et al.*, 2011).

Addition of *Tubifex* spp. at  $t = 48$  h resulted in a delayed increased penetration of dissolved oxygen (Fig. 2d) and increased rates of denitrification and bacterial activity in deeper areas of the sediment (Fig. 2b and c). It is well established that *Tubifex* spp. increase heterogeneity of the sediment through selective feeding and burrowing activity (Wavre and Brinkhurst, 1971; Van de Bund *et al.*, 1994; Rodriguez

*et al.*, 2001; Kaonga *et al.*, 2010). Mechanical reworking of sediments principally influences biogeochemical processes, as burrowing activity enhances irrigation and allocates terminal electron acceptors to deeper layers of the sediments (Robbins *et al.*, 1979; Chakrabarty and Das, 2006; Nwuche and Ugoji, 2010). This enhances the redox potential, which, in turn, create redox conditions at which nitrate is more stable and dissimilatory nitrate reduction is allowed. An increased heterogeneity after addition of the aquatic oligochaete *Tubifex* spp. was visible in the spatio-temporal measurements of Eh, denitrification and bacterial respiration (Fig. 2b and c), coinciding with an increased oxygen penetration depth (Fig. 2d), suggesting that biogeochemical characteristics in studied microcosms evolved in a comparable fashion and were tightly coupled. Continuous measurements of Eh thus reflected the spatial and temporal heterogeneity of biogeochemical processes in studied microcosms.

Performing linear regression on measured variables reveals a positive relation between redox potential and the overall bacterial respiration (Fig. 3) and rates of denitrification and redox potential (Fig. 4). Although some studies were able to correlate Eh with concentrations of nitrate in water bodies and soil

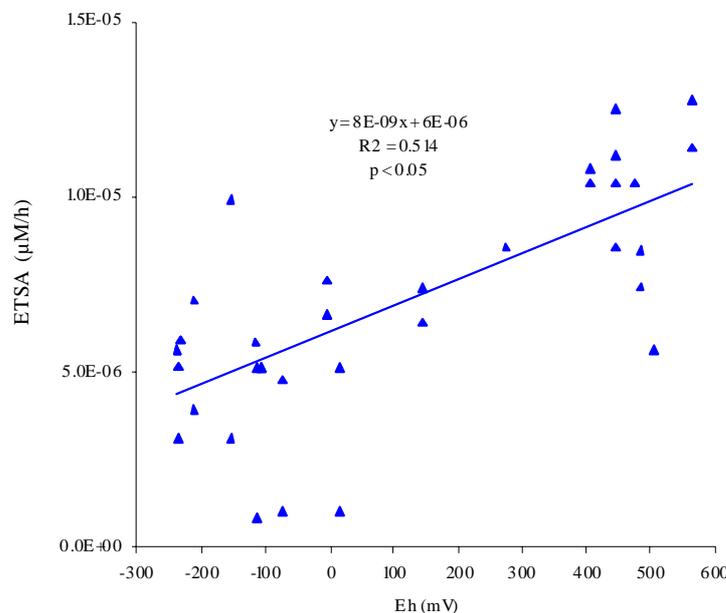


Fig. 3: Linear relation between redox potential and overall bacterial respiration (ETSA). Data reveal a positive relation between Eh and bacterial activity. Linear fit:  $y = 8E-09x + 6E-06$ . Statistics:  $R^2 = 0.514$ ;  $p < 0.05$

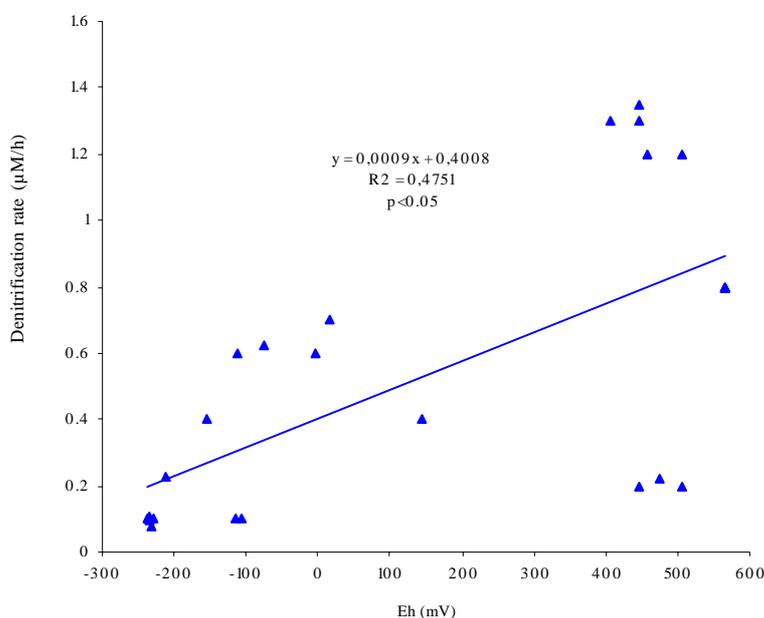


Fig. 4: Linear relation between Eh and rates of denitrification. Data reveal a positive relation between Eh and rates of denitrification. Linear fit:  $y = 0.0009x + 0.6139$ . Statistics:  $R^2 = 0.4751$ ;  $p < 0.05$ .

(Brünger, 1982; Gao *et al.*, 2002; Mansfeldt, 2004), the majority of studies have difficulties obtaining relations of Eh with bacteria-mediated geochemical processes. In the set up, only ~50 % of the variations in the measured rates in bacterial respiration and rates of denitrification are explained by redox potential. The lack of accuracy is most probably the result of the spatial resolution of measurements (10 mm increments), which unlikely captures fluctuations that may occur at millimeter and micrometer scales relevant to the microenvironment of the platinum electrodes, and this proved a critical issue in many experimental approaches in studying underlying mechanisms of Eh (Kappler *et al.*, 2005). It is anticipated that correlations will improve if analytical sensitivity is enhanced.

Since environmental redox reactions fail to behave in a pure Nernstian fashion, it is impossible to derive absolute quantitative information on redox processes from Eh measurements. For this reason it has been proposed that Eh measurements may at best be used qualitatively to describe sediments and soils as either oxidizing or reducing (Gao *et al.*, 2002), or semi-quantitatively by considering critical Eh ranges sensu Mansfeldt (2004), in which highly oxidizing conditions

provide little information due to the lack of electro-active redox systems, i.e. the low electron exchange density of oxygen at the platinum surface, while the lower levels of Eh can be used as an indication as to whether specific species are present or absent and which reactions dominate. Despite both theoretical and practical concerns regarding the interpretability of Eh, the redox potentials measured with permanently installed electrodes are positively related to rates of denitrification and bacterial respiration in the microcosms. Concordantly, continuous measurements of Eh were recently positively related to Cu-availability in artificial streams (Van der Geest and Leon Paumen, 2008). Although these results are correlative and the parameters were not exhaustive, the emerging patterns clearly reveal a predictive potential of Eh and encourage a closer look into the correlative nature of Eh with other biogeochemical processes. Evidently, since Eh subsumes a variety of biogeochemical reactions, which may be typical to specific systems (e.g. sulphate reduction is more common in marine sediments compared to freshwater sediments), prior knowledge on dominant processes in the system of interest is required to obtain

predictability in Eh. If this information is available, Eh may provide a low cost and rapid measure that reflects biogeochemical processes and functioning of sediments.

## CONCLUSION

This study aimed to evaluate the quantitative interpretability of Eh in relation to underlying processes within spatially and temporally dynamic aquatic sediments. The results presented here suggest that measurements of Eh have predictive potential in approximating rates of denitrification and overall bacterial respiration in aquatic sediments.

The presented data clearly suggests that, while sediment bulk measurements and measurements of single profiles of Eh and bacterial activity likely fail to provide ecological relevant information, the use of continuous measurements of Eh provide a useful parameter that reflects biogeochemical processes and dynamics of sediments.

## ACKNOWLEDGEMENTS

The authors express their gratitude to Michel Vorenhout and Girly Kelly for assistance and useful discussions.

## REFERENCES

- Abdel Ghani, N. T.; Hegazy, A. K.; El-Chaghaby, G. A., (2009). *Typha domingensis* leaf powder for decontamination of aluminium, iron, zinc and lead: Biosorption kinetics and equilibrium modeling. *Int. J. Environ. Sci. Tech.*, 6 (2), 243-248 (6 pages).
- Andersen, J. M., (1977). Rates of denitrification of undisturbed sediment from six lakes as a function of nitrate concentration, oxygen and temperature. *Arch. Hydrobiol.*, 80, 147- 159 (13 pages).
- Bhupathiraju, V. K.; Hernandez, M. T.; Landfear, D.; Alvarez-Cohen, L., (1999). Application of a tetrazolium dye as an indicator of viability in anaerobic bacteria. *J. Microbiol. Method.*, 37 (3), 231-243 (13 pages).
- Biati, A.; Karbassi, A. R.; Hassani, A. H.; Monavari, S. M.; Moattar, F., (2010). Role of metal species in flocculation rate during estuarine mixing. *Int. J. Environ. Sci. Tech.*, 7 (2), 327-336 (10 pages).
- Biati, A.; Karbassi A. R., (2010). Comparison of controlling mechanisms of flocculation processes in estuaries. *Int. J. Environ. Sci. Tech.*, 7 (4), 731-736 (6 pages).
- Bonin, P.; Rontani, J. F.; Bordenave, L., (2001). Metabolic differences between attached and free-living marine bacteria: inadequacy of liquid cultures for describing in situ bacterial activity. *FEMS Microbiol. Let.*, 194 (1), 111-119 (8 pages).
- Bright, J. J.; Fletcher, M., (1983). Amino acid assimilation and electron transfer system activity in attached and free-living marine bacteria. *Appl. Environ. Microbiol.*, 45 (3), 818-825 (8 pages).
- Brinkhurst, R. O.; Chua, K. E.; Kaushik, N. K., (1972). Interspecific interactions and selective feeding by tubificid oligochaetes. *Limno. Oceanograph.*, 17 (1), 122-133 (12 pages).
- Brünger, R., (1982). The redox potential as an indicator of microbial metabolic processes in north German waters. *Radiat. Environ. Biophysics*, 21 (2), 141-154 (14 pages).
- Chakrabarty, D.; Das, S. K., (2006). Alteration of macroinvertebrate community in tropical aquatic systems in relation to sediment redox potential and overlying water quality. *Int. J. Environ. Sci. Tech.*, 2 (4), 327-334 (8 pages).
- Gao, S.; Tanji, K. K.; Scardaci, S. C.; Chow, A. T., (2002). Comparison of redox indicators in a paddy soil during rice-growing season. *Soil Sci. Soc. Am. J.*, 66 (3), 805-817 (13 pages).
- Grenthe, I.; Stumm, W.; Laaksuharju, M.; Nilsson, A. C.; Wikberg, P., (1992). Redox potentials and redox reactions in deep groundwater systems. *Chem. Geol.*, 98, 131-150 (20 pages).
- Griess, P., (1864). On a new series of bodies in which nitrogen is substituted for hydrogen. *Phil. Trans. R. Soc.*, 154, 667-731 (65 pages).
- Hunting, E. R.; de Goeij, J. M.; Asselman, M.; van Soest, R. W. M.; van der Geest, H. G., (2010). Degradation of mangrove-derived organic matter in mangrove associated sponges. *Bull. Marine Sci.*, 86 (4), 871-877 (7 pages).
- Jackson, L. J.; Kalf, J.; Rasmussen, J. B., (1993). Sediment pH and redox potential affect the bioavailability of Al, Cu, Fe, Mn, and Zn to rooted aquatic macrophytes. *Can. J. Fish. Aqua. Sci.*, 1, 143-148 (6 pages).
- Kappler, A.; Emerson, D. E.; Edwards, D.; Amend, J. P.; Gralnick, J. A.; Grathwohl, P.; Hoehler, T.; Straub, K. L., (2005). Microbial activity in biogeochemical gradients - new aspects of research. *Geobiol.*, 3 (3), 229-233 (5 pages).
- Kaonga, C. C.; Kumwenda, J.; Mapoma, H. T., (2010). Accumulation of lead, cadmium, manganese, copper and zinc by sludge worms; *Tubifex tubifex* in sewage sludge. *Int. J. Environ. Sci. Tech.*, 7 (1), 119-126 (7 pages).
- Männistö M. K.; Tirola, M.; Häggblom, M. M., (2009). Effect of Freeze-Thaw cycles on bacterial communities of Arctic Tundra. *Soil Microb. Ecol.*, 58 (3), 621-631 (11 pages).
- Mansfeldt, T., (2004). Redox potential of bulk soil and soil solution concentration of nitrate, manganese, iron, and sulfate in two Gleysols. *J. Plant Nutr. Soil Sci.*, 167 (1), 7-16 (10 pages).
- Mohiuddin, K. M.; Zakir, H. M.; Otomo, K.; Sharmin, S.; Shikazono, N., (2010). Geochemical distribution of trace metal pollutants in water and sediments of downstream of an urban river. *Int. J. Environ. Sci. Tech.*, 7 (1), 17-28 (12 pages).
- Morley, C. R.; Trofymow, J. A.; Coleman, D. C.; Cambardella, C., (2005). Effects of freeze-thaw stress on bacterial populations in soil microcosms. *Microb. Ecol.*, 9 (4), 329-340 (12 pages).
- Nouri, J.; Mirbagheri, S. A.; Farrokhan, F.; Jaafarzadeh, N.; A. Alesheikh, A., (2010). Water quality variability and

- eutrophic state in wet and dry years in wetlands of the semiarid and arid regions. *Environ. Earth. Sci.*, 59 (7), 1397-1407 (**29 pages**).
- Nwuche, C. O.; Ugoji, E. O., (2010). Effect of co-existing plant specie on soil microbial activity under heavy metal stress. *Int. J. Environ. Sci. Tech.*, 7 (4), 697-704 (**8 pages**).
- Robbins, J. A.; McCall, P. L.; Berton Fisher, J.; Krezoski, J. R., (1979). Effect of deposit feeders on migration of <sup>137</sup>Cs in lake sediments. *Earth Planet. Sci. Lett.*, 42 (2), 277-287 (**11 pages**).
- Rodriguez, P.; Martinez-Madrid, M.; Arrate, J. A.; Navarro, E., (2001). Selective feeding by the aquatic oligochaete *Tubifex tubifex* (Tubificidae, Clitellata). *Hydrobiol.*, 463, 133-140 (**8 pages**).
- Rozzi, A.; Remigi, E., (2004). Methods of assessing microbial activity and inhibition under anaerobic conditions: A literature review. *Rev. Environ. Sci. Bio. Tech.*, 3, 93-115 (**23 pages**).
- Saunders, D. L.; Kalf, J., (2001). Denitrification rates in the sediments of Lake Memphremagog, Canada-USA. *Water Res.*, 35 (8), 1897-1904 (**8 pages**).
- Sekabira, K.; Oryem Origa, H.; Basamba, T. A.; Mutumba, G.; Kakudidi, E., (2010). Assessment of heavy metal pollution in the urban stream sediments and its tributaries. *Int. J. Environ. Sci. Tech.*, 7 (3), 435-446 (**12 pages**).
- Smith, J. J.; McFeters, G. A., (1997). Mechanisms of INT (of 2-(4-iodophenyl)-3-(4-nitrophenyl)-5-phenyl tetrazolium chloride), and CTC (5-cyano-2,3-ditolyl tetrazolium chloride) reduction in *Escherichia coli* K-12. *J. Microbiol. Methods*, 29, 161-175 (**15 pages**).
- Seitzinger, S. P., (1988). Denitrification in Freshwater and Coastal Marine Ecosystems: Ecological and Geochemical Significance. *Limnol. Oceanograph.*, 33 (4), 702-724 (**23 pages**).
- Teasdale, P. R.; Minett, A. I.; Dixon, K.; Lewis, T. W.; Batley, G. E., (1998). Practical improvements for redox potential ( $E_h$ ) measurements and the application of a multiple-electrode redox probe (MERP) for characterizing sediment in situ. *Anal. Chimica Acta*, 367 (1-3), 201-213, (**13 pages**).
- Van de Bund, W. J.; Goedkoop, W.; Johnson, R. K., (1994). Effect of deposit-feeder activity on bacterial production and abundance in profundal lake sediment. *J. N. Am. Benhol. Soc.*, 13 (4), 532-539 (**8 pages**).
- Van der Geest, H. G.; Leon Paumen, M., (2008). Dynamics of metal availability and toxicity in historically polluted floodplain sediments. *Sci. Total Environ.*, 406 (3), 419-425 (**7 pages**).
- Vink, J. P. M., (2002). Measurement of heavy metal speciation over redox gradients in natural water-sediment interfaces and implications for uptake by benthic organisms. *Environ. Sci. Tech.*, 23, 5130-5138 (**9 pages**).
- Vorenhout, M.; Van der Geest, H. G.; Van Marum, D.; Wattel, K.; Eijsackers, H. J. P., (2004). Automated and continuous redox potential measurements in soil. *J. Environ. Qual.*, 33, 1562-1567 (**6 pages**).
- Vorenhout, M.; Van der Geest, H. G.; Hunting, E. R., (2011). An improved datalogger and novel probes for continuous redox measurements in wetlands. *Int. J. Environ. Anal. Chem.*, 91 (7-8), 801-810 (**10 pages**).
- Wavre, M.; Brinkhurst, R. O., (1971). Interactions between some tubificid oligochaetes and bacteria found in the sediments of Toronto Harbour, Ontario. *J. Fish. Res. Bd. Can.*, 28, 335-341 (**7 pages**).
- Yousefi Kebria, D.; Khodadadi, A.; Ganjidoust, H.; Badkoubi, A.; Amoozegar, M. A., (2009). Isolation and characterization of a novel native *Bacillus* strain capable of degrading diesel fuel. *Int. J. Environ. Sci. Tech.*, 6 (3), 435-442 (**8 pages**).
- Zhang, S. Y.; Wang, Q. F.; Xie, S. G.; (2011). Microbial community changes in contaminated soils in response to phenanthrene amendment. *Int. J. Environ. Sci. Tech.*, 8 (2), 321-330 (**10 pages**).

**AUTHOR (S) BIOSKETCHES**

**Hunting, E. R.**, University of Amsterdam, Aquatic Ecology and Ecotoxicology, IBED, Sciencepark 904, 1098 XH Amsterdam, the Netherlands. Email: [e.r.hunting@uva.nl](mailto:e.r.hunting@uva.nl)

**van der Geest, H. G.**, University of Amsterdam, Aquatic Ecology and Ecotoxicology, IBED, Sciencepark 904, 1098 XH Amsterdam, the Netherlands. Email: [h.g.vandergeest@uva.nl](mailto:h.g.vandergeest@uva.nl)

**How to cite this article: (Harvard style)**

Hunting, E. R.; van der Geest, H. G., (2011). Predictability of bacterial activity and denitrification in aquatic sediments with continuous measurements of redox potential. *Int. J. Environ. Sci. Tech.*, 8 (3), 553-560.