Identification of a novel anti-sigmaE factor in Neisseria meningitidis
Hopman, C.T.P.; Speijer, D.; van den Ende, A.; Pannekoek, Y.

Published in:
BMC Microbiology

DOI:
10.1186/1471-2180-10-164

Citation for published version (APA):

General rights
It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations
If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: http://uba.uva.nl/en/contact, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.
Identification of a novel anti-σE factor in *Neisseria meningitidis*

Carla Th P Hopman¹, Dave Speijer², Arie van der Ende¹ and Yvonne Pannekoek*¹

**Abstract**

**Background:** Fine tuning expression of genes is a prerequisite for the strictly human pathogen *Neisseria meningitidis* to survive hostile growth conditions and establish disease. Many bacterial species respond to stress by using alternative σ factors which, in complex with RNA polymerase holoenzyme, recognize specific promoter determinants. σE, encoded by *rpoE* (NMB2144) in meningococci, is known to be essential in mounting responses to environmental challenges in many pathogens. Here we identified genes belonging to the σE regulon of meningococci.

**Results:** We show that meningococcal σE is part of the polycistronic operon NMB2140-NMB2145 and autoregulated. In addition we demonstrate that σE controls expression of methionine sulfoxide reductase (MsrA/MsrB). Moreover, we provide evidence that the activity of σE is under control of NMB2145, directly downstream of *rpoE*. The protein encoded by NMB2145 is structurally related to anti-sigma domain (ASD) proteins and characterized by a zinc containing anti-σ factor (ZAS) motif, a hallmark of a specific class of Zn²⁺-binding ASD proteins acting as anti-σ factors. We demonstrate that Cys residues in ZAS, as well as the Cys residue on position 4, are essential for anti-σE activity of NMB2145, as found for a minority of members of the ZAS family that are predicted to act in the cytoplasm and responding to oxidative stimuli. However, exposure of cells to oxidative stimuli did not result in altered expression of σE.

**Conclusions:** Together, our results demonstrate that meningococci express a functional transcriptionally autoregulated σE factor, the activity of which is controlled by a novel meningococcal anti-σ factor belonging to the ZAS family.

**Background**

RNA polymerase holoenzyme, consisting of a 5-subunit core RNA polymerase (α₂ββ′ω) and a dissociable subunit, sigma (σ), initiates bacterial transcription. The σ factor contains many of the promoter recognition determinants and several σ factors each recognizing their specific class of promoter sequences have been described [1-5]. In general, in exponentially growing bacteria transcription is initiated by RNA polymerase carrying the housekeeping σ, known as σ70 [6]. Alternative σ factors mediate transcription of regulons activated under specific environmental conditions [7,8]. The activity of many alternative σs is inhibited by a specific anti-σ factor. In a wide variety of bacterial species the σ factor σ55, also known as extracytoplasmic factor or ECF, belonging to the group IV σs, is essential in mounting responses to environmental challenges such as oxidative stress, heat shock, and misfolding of membrane proteins [9,10]. In addition, σE is of importance for virulence of bacterial pathogens [11-22]. The regulon size of σE varies widely among bacterial species studied, ranging from 89 unique σE controlled transcription units in *E. coli* and related bacteria [23] to a relatively small regulon of 5 genes in *Neisseria gonorrhoeae* [24]. In most examples, the gene encoding σE (rpoE) is located in an autoregulated operon that also contains, directly downstream of rpoE, the gene encoding its cognate anti-σE factor [25-28]. Extensive sequence analysis showed that about one third (1265/3600) of known and predicted anti-group IV σ factors, encoded in a gene cluster with a group IV σ (with only one exception), contain a conserved structural N-terminal fold, recently described as the anti-sigma domain (ASD) [26]. Typically, the ASD is in the N-terminus, oriented towards the cytoplasm, preceding a C-terminal transmembrane domain.

* Correspondence: y.pannekoek@amc.uva.nl

¹ Academic Medical Center, Center for Infection and Immunity Amsterdam (CINIMA), Department of Medical Microbiology, Amsterdam, the Netherlands

Full list of author information is available at the end of the article

© 2010 Hopman et al; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
segment. However, 20% of the 1265 ASD containing proteins are not predicted to contain a transmembrane spanning C-terminal domain [26]. Among these, 95% (227/248) are characterized by the presence of an invariant Hisx3Cysx2Cys sequence motif important for anti-sigma activity, co-ordinating Zn2+, described as the zinc containing anti-σ factor (ZAS) group IV anti-σs proteins [29]. ASD proteins and ASD proteins containing the ZAS motif are predicted to bind specifically to σs and inhibit their activities [25-28].

The strictly human pathogen Neisseria meningitidis colonizes the nasopharynx of approximately 10 to 30% of the population. In rare instances colonization results in invasive disease leading to life-threatening septicemia and meningitis [30]. Meningococci possess a variety of genes involved in adaptation to specific changes in the environment encountered in the host [31-36]. In addition to nutrient limitation, meningococci are also exposed to massive amounts of reactive oxygen species produced by host defenses [37,38]. Fine tuning expression of genes required to survive hostile growth conditions is a prerequisite for the meningococcus to establish disease.

All four publicly available, completely sequenced genomes of N. meningitidis contain a gene (NMA0233, NMB2144, NMC2123 and NMCC:2103) encoding a protein with homology to σ E, the σ factor involved in stress responses [39-42]. In this study we explored the σ E regulon of N. meningitidis. In addition, we provide evidence that the expression of σ E (encoded by NMB2144) in meningococci is auto-regulated and that its activity is under control of a protein encoded directly downstream of rpoE. This protein, encoded by NMB2145, is structurally related to ASD proteins and contains the ZAS motif (His30x3Cys34x2Cys37). We demonstrate that the Cys residues in the ZAS motif, as well as a Cys on position 4, are important (Cys4 and C37) or essential (Cys34) for anti-σ activity of NMB2145.

**Results**

The gene cluster containing rpoE is transcribed as a polycistronic operon and transcriptionally regulated by σE

In many bacterial species, rpoE is part of an autoregulated polycistronic operon also encoding its cognate anti-sigma factor [25-28]. In meningococci, NMB2144 is annotated as rpoE, encoding a protein with a molecular weight of approximately 23 kDa, 98% identical to the σE orthologue of N. gonorrhoeae [24] and 28% identical to σE of E. coli. Meningococcal rpoE is part of a ~3 kb cluster of genes NMB2140 through NMB2145 (Fig.1a) having a genomic arrangement similar to that found in N. gonorrhoeae [24]. All genes, except NMB2144, are annotated as hypothetical proteins. The minimal spacing found in the cluster suggests co-transcription of its genes.

To investigate whether the genes found in the gene cluster NMB2140 through NMB2145 are co-transcribed and under transcriptional control of σE, a meningococcal strain in which expression of rpoE can be controlled, was generated by transformation of H44/76 with the shuttle vector pEN11 carrying rpoE under control of an IPTG-inducible promoter, creating H44/76 + pNMB2144. Transcript levels of the gene cluster were analysed by RT-PCR using RNA isolated from these cells grown in the absence and presence of IPTG and primer pairs as depicted in Fig. 1a. With either wt cells (not shown) or H44/76 + pNMB2144 cells grown in the absence of IPTG, hardly any detectable RT-PCR products of co-transcripts were found (see Fig. 1B, lane 1 to 3). Only the small 735 nt product (NMB2144-NMB2145, see Fig. 1B, lane 3) could be seen (the band in lane 2 is an unrelated product as shown by sequence analysis). In contrast, only when H44/76 + pNMB2144 cells were grown in the presence of IPTG, specific RT-PCR products, with sizes correspond-
ing to calculated sizes (2412 nt (Fig. 1B, lane 4) and 2123 nt (Fig. 1B, lane 5) containing the predicted sequences of NMB2140-NMB2144, were detected, while the 735 nt product was strongly induced (Fig. 1B, lane 6). These observations indicate that the gene cluster containing rpoE is transcribed as a polycistronic operon and transcriptionally regulated by σE<sup>e</sup>. The fact that complete transcript levels of the rpoE operon were only found upon overexpression of rpoE suggests that in H44/76 wt cells, under the growth conditions tested, the levels of (active) σE allow only barely detectable transcription.

**Identification of proteins under control of σ<sup>e</sup>**

To further explore the meningococcal σE regulon, protein patterns of the H44/76 wt strain, ΔrpoE and H44/76<sup>pNMB2144</sup> were compared by SDS-PAGE. No apparent protein expression level differences between H44/76 wt and ΔrpoE were observed in the proteomes of the cells (not shown). The addition of IPTG to the culture medium of cells transformed with pNMB2144 only gave minor changes in protein expression in the cytoplasm (Fig. 2a).

In contrast, in the crude membrane fraction, a dramatic increase in the expression of a ~60 kDa protein was observed (Fig. 2a). The increase in expression of this protein was IPTG dependent as the protein was hardly detectable in crude membranes prepared from the same cells not exposed to IPTG (Fig. 2a). Peptide mass fingerprinting, using MALDI-TOF MS to analyze the tryptic fragments generated by in gel digestion, identified this protein as the methionine sulfoxide reductase MsrA/MsrB, encoded by NMB0044 (Mowse score: 191 with 14 matching peptides).

**MsrA/MsrB is transcriptionally controlled by σ<sup>e</sup>**

To ascertain that msrA/msrB is under direct control of σ<sup>e</sup>, transcript levels of msrA/msrB in diverse meningococcal genetic backgrounds were analyzed by RT-PCR using RNA isolated from cells grown in the absence and presence of IPTG and primers targeting msrA/msrB. When H44/76 wt or H44/76<sup>+pNMB2144</sup> cells were grown in the absence of IPTG, no detectable RT-PCR products were observed. In contrast, when H44/76<sup>+pNMB2144</sup> cells were grown in the presence of IPTG, an RT-PCR product with a size indicative of transcription of msrA/msrB was found (Fig. 2b). The identity of the transcript was confirmed by sequencing of the RT-PCR product. These results strongly suggest that msrA/msrB is transcriptionally controlled by σE<sup>e</sup>.

**NMB2145 inhibits transcription of the rpoE operon**

One possible explanation for low σ<sup>e</sup> activity in H44/76 wt cells under the growth conditions tested is that σ<sup>e</sup> is kept in an inactive state through an interaction with an anti-σ factor, thereby preventing σ<sup>e</sup> binding to core RNA polymerase, one of the ways to inhibit σ activity found in σ-regulator circuits in other bacteria [43-47]. Interestingly, it was recently reported that NMB2145 contains the ZAS motif Hisx3Cysx2Cys [48], characteristic for a subset of group IV σ anti-σ factors, usually encoded directly downstream of rpoE and cotranscribed [26].

Amino acid sequence comparison of orthologues of NMB2145 in genomes of three other meningococcal strains, two gonococcal strains and six commensal neisserial species (N. cinerea, N. flavescence, N. lactamica, N. mucosa, N. sicca and N. subflava) revealed that the region containing the ZAS motif, as well as the region around Cys4, are highly conserved in these neisserial orthologues of NMB2145. This in contrast with other much less well conserved parts, highlighting the importance of the conserved regions (Fig. 3). The relative positions of the Cys residue and the ZAS motif in NMB2145 (Cys4, His30, Cys34 and Cys37) correspond exactly with those of the Cys residue and the ZAS motif in RsrA (Cys11, His37, Cys41 and Cys44), the anti-σ factor of Streptomyces coelicolor, of which the Cys residues, but not His37, are essential for anti-σ activity of the protein [29] (Fig. 3). These observations suggest that NMB2145 codes for the meningococcal anti-σ<sup>e</sup> factor.

To test this hypothesis we first investigated the effect of deletion or overexpression of NMB2145 on transcript levels of the rpoE operon. To this end, a NMB2145 deletion mutant (ΔNMB2145) was constructed and comple-
upon induction of overexpression of \textit{NMB2145} in the absence of IPTG was no longer detectable. This effect was dependent on the presence or absence of protein \textit{\Delta NMB2145} mutant by induction of expression of \textit{msrA}/\textit{msrB} using pEN11 carrying \textit{NMB2145} under control of an IPTG-inducible promoter (generating \textit{\Delta NMB2145} + p\textit{NMB2145}). Transcript levels of the \textit{rpoE} operon were assessed by semi-quantitative RT-PCR using primers annealing to \textit{NMB2140} and \textit{NMB2143}, respectively.

As noticed before, RT-PCR products derived from the transcript encoding MsrA/MsrB (Fig.2b) and products indicative of co-transcription of \textit{NMB2140-NMB2145} (Fig.1b and Fig. 4) were found only upon overexpression of \textit{rpoE} in trans. However, deletion of \textit{NMB2145} resulted in the direct detection of the \textit{NMB2140-2143} without the need for overexpression of \textit{rpoE} in the \textit{H44/76} wt background. As expected, upon complementation of the \textit{\Delta NMB2145} mutant by induction of expression of \textit{NMB2145} in trans, the \textit{NMB2140-2143} RT-PCR product was no longer detectable. This effect was then upon induction of overexpression of \textit{NMB2145} in \textit{\Delta NMB2145} as it was not observed in the absence of IPTG (Fig.4).

As one would predict, MsrA/MsrB protein was detected in \textit{\Delta NMB2145} and could not be detected anymore upon complementation of \textit{\Delta NMB2145} by \textit{NMB2145} when IPTG was added to the culture medium (Fig. 5a). Also, \textit{NMB0044} (\textit{msrA}/\textit{msrB}) RT-PCR product was indeed detected in \textit{\Delta NMB2145} cells but hardly in

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{figure3.png}
\caption{Sequence alignment of orthologues of \textit{NMB2145} as found in other neisserial species and the ZAS containing anti-\sigma factors RsrA and ChrR of \textit{R. sphaeroides}. Numbers at the end of each sequence represent the total length of the protein and bracketed numbers show the number of residues not shown in the alignment of RsrA and ChrR with \textit{NMB2145}. The ZAS motif (His\textsubscript{x3}Cys\textsubscript{x2}Cys) is indicated in yellow, the additional zinc ligand \cite{48} is indicated in red. Conserved residues of neisserial \textit{NMB2145} orthologues are indicated in green. Protein IDs or genomic coordinates (in case of missing protein annotation) are indicated on the right. Details regarding strains of which sequences were obtained are listed in the Materials and Methods section.}
\end{figure}

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{figure4.png}
\caption{\textit{NMB2145} represses transcription of the \textit{rpoE} operon

Products obtained by RT-PCR were separated on agarose gel. RT-PCR analysis of transcription of the \textit{rpoE} operon in the \textit{wt} strain (\textit{H44/76}), \textit{H44/76} transformed with \textit{pNMB2144} before (-) and after (+) induction of expression of \textit{rpoE}, \textit{H44/76} transformed with \textit{pNMB2145} before (-) and after (+) induction of \textit{NMB2145} in the \textit{\Delta NMB2145} background (upper panel). RT-PCR was carried out using primer pair 2140-01/2143-02 (cf Fig 1a) and RT-PCR on \textit{rmpM} (lower panel) was used as input control of total RNA.

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{figure5.png}
\caption{\textit{MsrA}/\textit{msrB} is expressed upon deletion of and transcriptionally repressed by \textit{NMB2145}. Protein analysis of crude membranes by SDS-PAGE (A) and corresponding transcriptional analysis of \textit{msrA}/\textit{msrB} by RT-PCR (B) of the \textit{NMB2145} knockout (\textit{\Delta NMB2145}) and \textit{\Delta NMB2145} transformed with \textit{pNMB2145} before (-) and after induction (+). Molecular weight markers (kDa) are indicated on the right. Arrow indicates MsrA/MsrB.}
ΔNMB2145 cells when complemented by NMB2145 (Fig. 5b).

Together, these experiments demonstrate that NMB2145 inhibits transcription of the rpoE regulon. Conceivably, NMB2145 binds to σE, thereby inactivating it, resulting in decreased transcription by means of auto-regulation of the rpoE operon and, as a consequence of that, decreased transcription of msrA/msrB.

The residues Cys4, Cys34 and Cys37 of NMB2145 are essential for optimal anti-σE activity

To investigate whether the Cys residues of the ZAS motif and the conserved Cys at position 4 of NMB2145, in analogy to corresponding Cys residues in RsrA of S. coelicolor [29], are also essential for anti-σE activity of NMB2145, we generated single Ala substitutions at each of the Cys residues and also of the single His residue of the ZAS motif (His30xCys34xCys37) and at position 4 of NMB2145. The ability of these mutant NMB2145 proteins to inhibit σE activity in meningococci was investigated by SDS-PAGE assessment of crude membranes, using MrsA/MrsB as reporter protein. All substitutions except His30Ala resulted in expression of MrsA/MrsB (MALDI-TOF confirmed). The substitution Cys34Ala resulted in MsrA/MsrB levels comparable to those found in crude membranes prepared from ΔNMB2145 cells while the substitutions Cys4Ala and Cys37Ala resulted in more modest, but clearly detectable levels of MsrA/MsrB (Fig. 6). Collectively, these experiments demonstrate that the Cys residues of the ZAS motif, as well as Cys4 of NMB2145 are important for functionality of NMB2145 as an anti-σE factor.

Involvement of σE in the response to hydrogen peroxide, diamide and singlet oxygen

The Cys4 and Cys37 in NMB2145, essential in anti-σE activity, correspond exactly with Cys11 and Cys44 residues of RsrA of S. coelicolor involved in disulphide bond formation. In addition, residue His30 in the ZAS motif of NMB2145 is not required for anti-σE activity consistent with anti-σ properties of RsrA [29] and ChrR, the ZAS containing anti-σE factor of Rhodobacter sphaeroides [26,49,50]. In S. coelicolor, exposure to superoxide, hydrogen peroxide or the thiol specific oxidant diamide causes dissociation of the σE-RsrA complex [46,51,52]. In contrast, ChrR anti-σE activity is not affected by these reactive oxygen species, but responds to singlet oxygen (1O2) [53].

To investigate stimuli activating the σE response in meningococci, cells were exposed to hydrogen peroxide, diamide or singlet oxygen and transcript levels of rpoE and msrA/msrB were analysed by RT-PCR before and after exposure to these stress agents. No differences in transcript levels of either rpoE or msrA/msrB were detected, suggesting that in meningococci σE is not involved in the response to such stimuli. In addition, no detectable differences in transcript levels of rpoE and msrA/msrB were observed after exposure of cells to SDS-EDTA, a stimulant known to induce membrane stress and activate RpoE in other bacterial species (not shown).

In silico genome wide search for additional genes under control of σE using a deduced neisserial σE-promoter consensus sequence

Each σ factor recognizes specific promoter sequences, characterized by relatively highly conserved -35 and -10 upstream DNA sequences. Using the promoter sequences of genes under the control of σE, a consensus sequence can be deduced. In several bacterial species, this motif has been successfully used for in silico genome searches to identify genes putatively controlled by σE. The σE dependent transcription of these genes can subsequently be confirmed by in vitro experiments [23,54-56].

To further explore the meningococcal σE regulon, we used a similar strategy. However, in the meningococcus we were able to demonstrate transcriptional control by σE for only one operon (the rpoE operon itself) and one gene (msrA/msrB), so far. Therefore, we extended the number of genes from which a σE promoter consensus sequence could be deduced with orthologues of NMB2140 and NMB0044 found in the sequences of 3 other meningococcal genomes, 2 gonococcal genomes and the genomes of 6 commensal neisserial species. In total, putative promoter sequences of 24 genes were used to generate a consensus promoter sequence by Weblogo [57]. Thus, the conserved putative -35 (GTAGBWT'T) and -10 (CGTCTAAH) motifs could be identified (Fig. 7). These motifs are separated by spacer of 12-13 nt (not shown). In addition, an AT rich sequence was observed 30 nt upstream of the -35 motif, corresponding to a consensus

![Figure 6 Residues Cys4, Cys34 and Cys37 of NMB2145 are essential for optimal anti-σE activity of NMB2145. SDS-PAGE assessment of MsrA/MsrB protein levels in crude membranes extracted from ΔNMB2145 cells in which mutant NMB2145 proteins pNMB2145(His30Ala), pNMB2145(Cys4Ala), pNMB2145(Cys34Ala) and pNMB2145(Cys37Ala) are expressed. Crude membranes were extracted before (-) and after (+) induction. Molecular weight markers (kDa) are indicated on the right. Arrow indicates MsrA/MsrB.](http://www.biomedcentral.com/1471-2180/10/164)
sequence designated the UP element [58-60]. Six nucleotides downstream of the -10 motif a highly conserved adenosine is found. This nucleotide and its position correspond exactly with the transcriptional start as experimentally identified for msrA/msrB in gonococci [24].

Next, we used the -35 and -10 motif, allowing a spacer length of 10 to 16 nt to explore the genome of N. meningitidis MC58 to identify genes containing the rpoE promoter motif. Besides NMB2140 and NMB0044, no other genes were identified.

Discussion

According to the annotation of the genome of N. meningitidis four genes are supposed to encode σ factors: rpoD (σ70), rpoH (σ32), rpoN (σ24) and rpoE (σE) [24,39-42]. To our knowledge, so far no information is available regarding the functionality of alternative σ factors in the meningococcus. Here, we describe the first detailed investigation of the functionality of σE of N. meningitidis. In addition, we provide strong evidence that NMB2145, encodes a novel anti-σ factor structurally related to ASD proteins and containing the ZAS motif, making NMB2145 the first anti-σ-factor described for any neisserial species.

Experimental evidence for transcriptional control by σE could be provided for only 7 genes, the 6 gene containing σE operon and msrA/msrB. In line with this, genome wide in silico searches for genes with a σE promoter motif also did not result in additional genes putatively controlled by σE. This suggests a surprisingly small σE regulon in meningococci, as well as in gonococci [24] as compared to that of other bacterial species as σE regulons can comprise up to 89 transcription units (in E. coli K-12 and related bacteria) [23].

Although the consensus σE promoter recognition motifs of the -35 region of meningococci (GTAAGGTT) and E. coli (GGAACTT) are quite different, the last 5 residues of the -10 motifs of meningococci (TCTAA) and E. coli (TCAAA) differ in only one nucleotide [23]. In addition, other similarities between the structural elements of these promoter regions were observed, such as the AT-rich sequence ~30 nt upstream of the -35 motif. This sequence, designated the UP-element, is a binding site for the C-terminal domain of the α-subunit of RNA polymerase [58-60] and has recently been shown to increase transcription of σE dependent promoters [61].

Recently, the first comprehensive analysis of conservation and variation of the σE regulon in E. coli and related organisms was reported [23]. The products of the core genes of the conserved σE regulon coordinate assembly and maintenance of lipopolysaccharide (LPS) and outer membrane proteins (OMPs) of Gram-negative bacteria, in response to cell envelope stress. The majority of the variable regulon members are functionally involved in pathogenesis [23]. Of interest, it was also recently demonstrated that σE promoters in E. coli and its close relatives exhibit a large dynamic range, with a few strong and many weak promoters [61]. The three strongest promoters all carry out regulatory roles in the σE response, the strongest transcribing σE itself and its negative regulators, and the next two strongest transcribing small RNAs (sRNAs) involved in downregulation of porin expression [61]. We did not observe expression differences of major outer membrane proteins upon overexpression of σE, making the involvement of σE in meningococci cell envelope stress uncertain. This was confirmed by our observation that membrane stress did not alter σE activity. However, as mentioned before, the majority of σE dependent proteins are expressed at low levels [61], which might be below the detection limit of the assay used in this study. As it is much easier to detect small changes in the transcriptome comparing ΔrpoE (σE knock out) or H44/76 + pNMB2144 (σE overexpression) with H44/76 (wt strain), we are planning those experiments. The recent identification in N. meningitidis of an sRNA controlling a gene and functional Hfq facilitating the interaction between sRNA and target mRNA, suggests the existence of a ribo-regulated network in this pathogen [62-65]. In many other species links between the σE regulon and the ribo-regulated network exist [66-71], but in meningococci this is as yet unexplored.

The genetic organization of the rpoE operon (NGO1948 through NGO1943) of N. gonorrhoeae is identical to that of meningococci (NMB2140-NMB2145), and four genes, NGO1946, NGO1947, NGO1948 belonging to the rpoE operon, and NGO2059, encoding MsrA/MrsB, were also upregulated, along with σE (NGO1944) itself, in a gonococcal strain overexpressing rpoE [24]. We demonstrated cotranscription of all genes in the meningococcal rpoE operon. The function of proteins encoded by NMB2140-NMB2143 is currently unknown. NMB2140 might encode a protein with possible trans membrane domains and contains motifs found in the
MsrA/MsrB is upregulated together with the genes of Caulobacter crescentus. Interestingly, in N. meningitidis, msrA/msrB, which we demonstrated to be transcriptionally controlled by σE, was abundantly expressed. Irrefutable evidence for a functional interaction of NMB2145 with σE was obtained by the substitution of Cys residues with Ala at positions in NMB2145 that correspond to Cys residues in RsrA. We found that Cys34 of NMB2145 is essential and, albeit to a lesser extent, Cys4 and Cys37 are also required for optimal anti-σE activity of NMB2145. We therefore suggest annotating NMB2145 as MseR, Meningococcal σe Regulator.

RsrA is a metalloprotein, containing near-stoichiometric amounts of Zn^{2+}. Oxidation induces a disulphide bond between two of the Zn^{2+} ligands (Cys11 and Cys44) resulting in loss of Zn^{2+} and dissociation of the σE-RsrA complex, thereby allowing σE transcription. Thioredoxin is able to reduce oxidized RsrA, and the induction of expression of thioredoxin itself is σE dependent, suggesting that σE, RsrA and the thioredoxin system in S. coelicolor comprise a novel feedback homeostasis loop, sensing and responding to changes in the intracellular thiol-disulphide redox balance. The Cys4 and Cys37 in NMB2145, of importance in anti-σE activity, correspond exactly with Cys11 and Cys44 residues of RsrA involved in disulphide bond formation, suggesting that MseR also contains Zn2+. Therefore, it was tempting to speculate that a similar thiol-disulphide redox balance also exists in meningococci. However, in N. meningitidis thioredoxin appears not to be upregulated upon exposure to hydrogen peroxide and we showed that transcription levels of MsrA/MsrB are not affected after exposure of meningococci to hydrogen peroxide, diamide or singlet oxygen. Whether NMB2145 is also a Zn^{2+} containing protein, deserves further study.

MsrA/MrsB, encoding methionine sulfoxide reductase, an enzyme repairing proteins exposed to reactive oxygen species, is a major target of σE, and abundantly expressed when active σE levels are high. Expression of MsrA/MrsB is also controlled by σE in N. gonorrhoeae and Caulobacter crescentus. Interestingly, in N. gonorrhoeae MsrA/MrsB is upregulated together with the genes NGO1947 and NGO1948 in response to hydrogen peroxide. However, none of the meningococcal orthologues, nor σE activity, as shown in our study, appear to respond to hydrogen peroxide, strongly indicating the existence of different modes of regulation of σE between gonococci and meningococci. In addition we did not find detectable differences in transcription levels of MsrA/MsrB after exposure to SDS-EDTA, a stimulant known to activate RpoE in other bacterial species. Thus, in vivo stimuli activating the σE response in N. meningitidis are most likely different from those of gonococci and remain to be further explored.

**Conclusions**

The results show the existence of a σE regulon in meningococci. The product of NMB2145 (MseR) functions as an anti-σE factor with properties different from membrane spanning anti-σE factors responding to signals in the periplasm. Our data strongly indicate that MseR, the meningococcal anti-σE factor, closely mimics structural properties of members of the ZAS family that are acting on novel stimuli encountered in the cytoplasm. Stimuli of MseR differ from those of the ZAS family anti-sigma factors suggesting that MseR is a novel anti-σ factor. This could indicate a potentially important, specific role for σE in the pathogenesis of meningococcal disease.

**Methods**

**Bacterial strains and culture conditions**

*N. meningitidis* strain H44/76, B: P1.7,16: F3-3: ST-32 (cc32), is closely related to the sequenced serogroup B strain MC58, belonging to the same clonal complex. Meningococci were grown on GC plates (Difco) supplemented with 1% (vol/vol) Vitox (Oxoid) at 37°C in a humidified atmosphere of 5% CO2. Broth cultures were incubated in tryptic soy broth (BD) on a gyratory shaker (180 rpm) at 37°C. When appropriate, plates or broths were supplemented with erythromycin (Erm) (5 μg/ml) and/or chloramphenicol (Cm) (5 μg/ml). Growth was monitored by measuring optical density of cultures at 600 nm (OD_{600}) at regular time intervals. To investigate the effect of various stress agents on RpoE activity, cells were grown to mid log phase (OD_{600} = 0.6-0.7) and treated for different time periods (30 min-1 h) with hydrogen peroxide (5 mM), diamide (100 mM), 0.01% SDS-0.1 mM EDTA or methylene blue (1 μM) in the presence of white light (source of singlet oxygen). Sequences from the following strains (with Genbank ID) were downloaded for comparative alignments: *N.meningitidis_MC58* (AE002098); *N.meningitidis_FAM18* (AM421808); *N.meningitidis_053442* (CP000381); *N.meningitidis_Z2491* (AF157959); *N gonorrhoeae_FA1090* (AE004969); *N gonorrhoeae_NCCP11945* (CP001050).
N. cinerea_ATCC_14685 (ACDY00000000); N. flavescens_NRL30031/H210 (ACEN00000000); N. lactamica_ATCC_23970 (ACE00000000); N. subflava_NJ9703 (ACE00000000); N. sicca_ATCC_29256 (ACK00000000); N. mucosa_ATCC_25996 (ACDX00000000); Streptomyces coelicolor_A3(2) (AL645882); Rhodobacter sphaeroides_ATCC_17025 (CP000661).

Construction of ΔrpoE and ΔNMB2145 mutants of N. meningitidis

N. meningitidis H44/76 knock-out mutants of rpoE (NMB2144) and NMB2145 were constructed using the PCR-ligation-PCR method [79,80]. All primers used in this study are listed in Table 1. PCR products were generated with primer pairs CTsE-1/CTsE-2 and CTsE-3/CTsE4 for creating ΔrpoE and primer pairs CT-2145-1/CT2145-2 and CT-2145-3/CT-2145-4 for creating ΔNMB2145, ligated and the ligation products were reamplified with primer pairs CTsE-1/CTsE-4 (for ΔrpoE) and CT-2145-1/CT-2145-4 (for ΔNMB2145). The resulting PCR products were cloned into pCR2.1 (Invitrogen). The EcoRI digested Erm resistance cassette from pAErmC' was introduced into the created unique MfeI restriction site yielding plasmids pCR2.1-NMB2144 and pCR2.1-NMB2145. The ΔrpoE and ΔNMB2145 strains were generated by natural transformation of strain H44/76 with pCR2.1-NMB2144 and pCR2.1-NMB2145 respectively, and selection for Erm resistance. Replacement of NMB2144 and NMB2145 by the Erm cassette was confirmed by PCR with primer pair CTsE-5/CTsE-6 (for ΔrpoE) and primer pair 2144-01/2144-02 for ΔNMB2145. The orientation of the Erm cassette was determined by PCR using primer pair JP19/JP20 and mutant strains in which the transcriptional direction of the Erm cassette was in accordance with the transcriptional direction of the deleted genes were selected.

Overexpression of rpoE and complementation of ΔNMB2145

To overexpress rpoE and to complement ΔNMB2145, NMB2144 and NMB2145 of strain H44/76 were amplified with primer pair CTsE-7/CTsE-8 (for NMB2144) and primer pair CT-2145-11F/CT-2145-11R (for NMB2145). Forward primers (CTsE-7 and CT-2145-11F) contain an Ndel restriction site and the reversed primers an RcaI restriction site. The resulting PCR products and shuttle vector pEN11-pldA [82] were digested with Ndel and RcaI, ligated into Ndel-RcaI predigested pEN11-pldA and transformed to E. coli TOP10® (Invitrogen). Cm-resistant colonies were checked by colony PCR and sequencing, using pEN11F, pEN11R and ALpEN11F2 primers. Plasmids of clones containing an intact NMB2144 coding region (pNMB2144) or an intact NMB2145 coding region (pNMB2145) were isolated to transform H44/76 and ΔNMB2145 thereby generating H44/76 + pNMB2144 and ΔNMB2145 + pNMB2145, respectively. Expression of recombinant DNA was induced by addition of IPTG to the culture medium to a final concentration of 1 mM.

In vitro mutagenesis of NMB2145

Shuttle vector pNMB2145 was used to generate mutant NMB2145 proteins. Point mutations were generated using Quickchange site-directed mutagenesis (Stratagene). Four mutants of pNMB2145 were generated: Cys4Ala, His30Ala, Cys34Ala and Cys37Ala, using primer pairs YPNMB2145C4AFW-YPNMB2145C4ARP (for mutant pNMB2145(Cys4Ala)), YPNMB2145H30AFW-YPNMB2145H30ARP (for mutant pNMB2145(His30Ala)), YPNMB2145C34AFW-YPNMB2145C34ARP (for mutant pNMB2145(Cys34Ala)) and YPNMB2145C37AFW-YPNMB2145C37ARP (for mutant pNMB2145(Cys37Ala)). Mutations were confirmed by sequence analysis.

RT-PCR

RNA was isolated from meningococci grown to late mid log phase (OD600 1.0 - 1.5) using Rneasy® Midi Kit (Qiagen). RT-PCR was done using SuperScriptIII (Invitrogen). Primer pairs used to investigate whether NMB2140 through NMB2145 is transcribed as a polycistronic operon are depicted in Fig. 1. Primer pair CT-MSR-01/CT-MSR-02 was used to investigate transcription of NMB0044. One single batch of cDNA generated from RNA isolated from H44/76 wt, H44/76 + pNMB2144, ΔNMB2145 and ΔNMB2145 + pNMB2145, grown in the absence and presence of IPTG, was used for transcriptional analyses of the rpoE operon and NMB0044. To investigate the effect of hydrogen peroxide, diamide and singlet oxygen on RpoE activity, RNA was isolated from midlog phase grown cells with and without exposure to the stress stimuli and primer pairs CT-MSR-01/CT-MSR-02 and 2144-01/2144-02 were used to investigate transcription of NMB0044 and NMB2144 respectively. RT-PCR of RmpM (NMB0382) using primerset CT-class4-1/CT-class4-2, was used as loading control. Sequence analysis was carried out to confirm the identity of the generated RT-PCR products.

Cell fractionation

Meningococci were grown in broth until OD600 = 0.6-0.8, harvested by centrifugation (20 min at 5000 × g) and resuspended in 50 mM Tris-HCl (pH 7.8). Meningococcal cells were disrupted by sonication (Branson B15 Sonifier, 50 W, 10 min, 50% duty cycle, 4°C), followed by centrifugation (3000 × g, 4 min, 4°C). The supernatant
Table 1: Oligonucleotides used in this study

<table>
<thead>
<tr>
<th>Name</th>
<th>Sequence 5'-3'</th>
</tr>
</thead>
<tbody>
<tr>
<td>CTsE-1</td>
<td>AACAGCCTTGTTAAATATGGCG</td>
</tr>
<tr>
<td>CTsE-2</td>
<td>ATCACAATTGATATGGCCGACAGAG</td>
</tr>
<tr>
<td>CTsE-3</td>
<td>CAGGACGGGAAACACC</td>
</tr>
<tr>
<td>CTsE-4</td>
<td>GAAGTGTGATATTAGGATG</td>
</tr>
<tr>
<td>CTsE-5</td>
<td>GCGAGTCCCTCCAATATGGCC</td>
</tr>
<tr>
<td>CTsE-6</td>
<td>CGGAGGGGTGGTGCGC</td>
</tr>
<tr>
<td>CT-2145-1</td>
<td>GCCAGGAAGGCGACGGC</td>
</tr>
<tr>
<td>CT-2145-2</td>
<td>ATCACAATTGGTTTACTTCGGATTTTCTTG</td>
</tr>
<tr>
<td>CT-2145-3</td>
<td>ACGGCAATATGCTGACG</td>
</tr>
<tr>
<td>CT-2145-4</td>
<td>GCTTGAGATTTGTGC</td>
</tr>
<tr>
<td>CT-2145-5</td>
<td>TGGGAGTTCCTCAATATTGCC</td>
</tr>
<tr>
<td>CT-2145-6</td>
<td>ATCATATGGCCGCTACCCGAC</td>
</tr>
<tr>
<td>CTsE-7</td>
<td>TACATGAACGGTTACTTCCGGG</td>
</tr>
<tr>
<td>CT-2145-11F</td>
<td>CGTGCATATGAAATGCCC</td>
</tr>
<tr>
<td>CT-2145-11R</td>
<td>TTTCAATGAGCCGCTAATTAATGGATTTCTG</td>
</tr>
<tr>
<td>2140-01</td>
<td>TGGGAAACACCTGGCA</td>
</tr>
<tr>
<td>2143-02</td>
<td>GACATTCTCTAAGCA</td>
</tr>
<tr>
<td>2142-01</td>
<td>ATGATTCAAACGCCGAG</td>
</tr>
<tr>
<td>2144-01</td>
<td>ACCCGACCTGACCAGG</td>
</tr>
<tr>
<td>2144-02</td>
<td>GGTGGCATATAGGTTGTGTG</td>
</tr>
<tr>
<td>2145-02</td>
<td>CGGGTTGTTTGCAG</td>
</tr>
<tr>
<td>CT-class4-1</td>
<td>CAACAGCCTGAAATTAAGCCG</td>
</tr>
<tr>
<td>CT-class4-2</td>
<td>GTATGATTTGTGTCGCCG</td>
</tr>
<tr>
<td>CT-MSR-01</td>
<td>CTTTGCGCCAAGTTCGGC</td>
</tr>
<tr>
<td>CT-MSR-02</td>
<td>CTTTCACCTTCAACGCC</td>
</tr>
<tr>
<td>pEN11F</td>
<td>TGGGAAATGTGTGACGCGA</td>
</tr>
<tr>
<td>pEN11R</td>
<td>AGCAAAAAAGGAAAGAAA</td>
</tr>
<tr>
<td>ALpEN11F2</td>
<td>GACAATTAATCAGGCTAGT</td>
</tr>
<tr>
<td>JP19</td>
<td>TAAATACAAACGCTACCCG</td>
</tr>
<tr>
<td>JP22</td>
<td>AAATGGTCAATTTCCTGATG</td>
</tr>
<tr>
<td>YPNMB2145-C4A-FW</td>
<td>GCAATGAGAAAAAAAGCCCCCGATATCGCC</td>
</tr>
<tr>
<td>YPNMB2145-C4A-RP</td>
<td>GGGCGGTATCGCGGGGCTTTTTATCGAT</td>
</tr>
<tr>
<td>YPNMB2145-H30A-FW</td>
<td>GATTCCATATACACCCCTGTTCTTCTGCGC</td>
</tr>
<tr>
<td>YPNMB2145-H30A-RP</td>
<td>CGGACAGAACACAGGCGCTTGTGTATGGAATCC</td>
</tr>
<tr>
<td>YPNMB2145-C34A-FW</td>
<td>CACACACCTGCTGCCGCTATATGCTGTG</td>
</tr>
<tr>
<td>YPNMB2145-C34A-RP</td>
<td>CACCCAAATCGGACGACGCGAAGGAGGTGTTGTG</td>
</tr>
<tr>
<td>YPNMB2145-C37A-FW</td>
<td>GCTTTTCTGCGATATCCGCGAATACCAAGAC</td>
</tr>
<tr>
<td>YPNMB2145-C37A-RP</td>
<td>GTCTTTTATATCCCGGGGATACCGAGGACGAC</td>
</tr>
</tbody>
</table>
was centrifuged \((100,000 \times g, 60 \text{ min}, 4^\circ \text{C})\). This way obtained supernatant was considered as the cytoplasmic fraction and pellets, containing crude membranes were resuspended in 2 mM TrisHCl (pH 6.8). Protein concentrations were determined by the method described by Lowry [82,83].

**SDS-PAGE and MALDI-TOF mass spectrometry**

Proteins were resolved by SDS-PAGE [84]. Gels (11%) were stained with PageBlue (Fermentas), washed in MilliQ water and stored in 1% acetic acid at 4°C until bands of interest were excised for further analysis. MALDI-TOF mass spectrometry was carried out as described previously [64].

**Authors’ contributions**

CThPH participated in the design of the study, carried out experiments and analyses of the data and helped to draft the manuscript. DS carried out the MALDI-TOF mass spectrometry and helped to draft the manuscript. AvdE participated in the design of the study, carried out the analyses of the data and helped to draft the manuscript. YP participated in the design of the study, carried out the analyses of the data and drafted the manuscript. All authors read and approved the final manuscript.

**Acknowledgements**

Melanie Nguyen is acknowledged for her technical assistance. This research was partly funded by the Sixth Framework Programme of the European Commission, Proposal/Contract no.: 512061 (Network of Excellence ‘European Virulence & Pathogenesis’ (CINIMA), Department of Medical Microbiology, Amsterdam, the Netherlands and 2Academic Medical Center, Department of Medical Biochemistry, Amsterdam, the Netherlands.

Received: 20 January 2010 Accepted: 4 June 2010

**Published:** 4 June 2010

**References**


