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Published in:
PLoS ONE

DOI:
10.1371/journal.pone.0052118

Link to publication

Citation for published version (APA):
https://doi.org/10.1371/journal.pone.0052118
Phox2b Influences the Development of a Caudal Dopaminergic Subset

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Abstract

The developing mesencephalic dopaminergic (mdDA) neuronal field can be subdivided into several molecularly distinct domains that arise due to spatiotemporally distinct origins of the neurons and distinct transcriptional pathways controlling these neuronal subsets. Two large anatomically and functionally different subdomains are formed that eventually give rise to the SNc and VTA, but more subsets exist which require detailed characterization in order to better understand the development of the functionally different mdDA subsets, and subset-specific vulnerability. In this study, we aimed to characterize the role of transcription factor Phox2b in the development of mdDA neurons. We provide evidence that Phox2b is co-expressed with TH in a dorsal-caudal subset of neurons in the mdDA neuronal field during embryonic development. Moreover, Phox2b transcripts were identified in FAC-sorted Pitx3 positive neurons. Subsequent analysis of Phox2b mutant embryos revealed that in the absence of Phox2b, a decrease of TH expression occurred specifically in the midbrain neuronal subset that normally co-expresses Phox2b with TH. Our data suggest that Phox2b is, next to the known role in the development of the oculomotor complex, involved in the development of a specific caudal mdDA neuronal subset.

Introduction

In Parkinson’s disease (PD) the onset is highlighted by a specific degeneration of meso-diencephalic dopaminergic (mdDA) neurons of the Substantia nigra pars compacta (SNc). To understand this neuron specific vulnerability, a thorough understanding of the development of mdDA neurons is essential. Based on a study in Lmx1b null mutants, it was suggested that Lmx1b is required for the generation of properly differentiated mdDA neurons. This study demonstrated a remarkable loss of developing mdDA neurons co-expressing Pitx3 and Th, in E12.5 Lmx1b−/− tissue [1]. Furthermore, Lmx1b plays an important role in the correct specification of the mid-hindbrain boundary (MHB), where it regulates expression of Fgfl, Wnt1, and several isthmus-related transcription factors, and it is required for the inductive activity of the isthmic organizer (io) itself [2,3]. Notably, the clear reduction of mdDA neurons in Lmx1b−/− embryos is likely due to an early loss of a large part of the midbrain, due to affected MHB patterning [2].

Previously, we identified Lmx1b as upstream activator of Phox2a (unpublished data). These data were recently confirmed as they showed the dependence of Phox2a expression in oculomotor neurons on the activity of Lmx1b [4]. Phox2a is expressed in the midbrain oculomotor complex (OMC), that partially overlaps with the mdDA neuronal field during development. Over the years, Phox2a has been identified as an important regulator of midbrain motorneuron development, in mice and humans, and in Phox2a−/− mice, midbrain oculomotor neurons are absent [5–7]. A recent study in chick suggested that exogenous Phox2a can induce a complete OMC molecular program, and can act as a primary developmental determinant for the oculomotor complex [8]. The functional parologue of Phox2a, Phox2b, is expressed in the hindbrain where it plays an essential role in the specification of cranial motor neurons [6,9–11]. Importantly, Phox2b is expressed in the OMC as well, and recently some degree of cooperation between Phox2a and Phox2b was discovered in motorneuron development [12]. In addition, molecular evidence was provided that Phox2b can regulate the expression of Phox2a by specifically interacting with the 5′-regulatory region of Phox2a [13].

The collective data on regulation of Phox2a by Lmx1b and Phox2b, aimed our interest towards a putative role of Phox2b in caudal subset specification of mdDA neurons. In this study, we show that Phox2b is co-expressed with TH and importantly, with Pitx3 in mdDA neurons during development. Interestingly, subsequent analysis of Phox2b null mutants revealed decreased expression of TH in the exact subset that normally expresses Phox2b, indicating that Phox2b plays a role in the development of this specific caudal subset of mdDA neurons.

Materials and Methods

Ethics statement

Mice were bred in our laboratory under standard conditions and all procedures were fully approved by the Dutch Ethical Committee (DEC) for animal experimentation of the University Medical Center Utrecht in the Netherlands (DEC-UMC-U) and international guidelines.
Animals

Experiments were carried out in C57Bl/6J wild-type mice (Charles River). Pregnant mice were decapitated or euthanized by CO2 asphyxiation and embryos were collected at E12.5, E13.5 and E14.5 (the morning of detection of a copulatory plug was considered E0.5). Pups were euthanized by CO2 asphyxiation and brains were isolated at postnatal (P) day 0, P7 and P14. Phox2b-LacZ mutant mice [14] were maintained under the same conditions. Embryos were collected at E12.5, E14.5, E16.5 and E18.5. Since homozygous mutants rarely survive after E13.5, due to a noradrenalin deficit [10], we treated drinking water of pregnant Phox2b-LacZ/+ mice by supplementing with 100 ug/mL of L-phenylephrine (Merck), 100 ug/mL isoproterenol (Sigma) and 2 mg/mL ascorbic acid (Sigma), from E8.5 onwards. Animals were genotyped by means of PCR, using a forward primer located in intron 1 of the Phox2b coding sequence (5'GTTCTGGTTCAGTGGCCCTTC) and a reverse primer in the LacZ inserted sequence (5'-AGGGTGGCAAACGTGGTGG) resulting in a product of 260 bp in mutants, or no product, in wild-type animals. To discriminate between heterozygous and homozygous LacZ mutants, the same forward primer was used, with a reverse primer in the wild-type sequence directly after the LacZ insertion (5'-CCAAGGCTATCCAGGATCTTAG). Pitx3-GFP mice were described previously [15]. RNA from Pitx3-GFP/+ embryos was used for fluorescence-activated cell sorting (FACS).

In situ hybridization

Embryos were collected in ice-cold buffer and immediately frozen on dry ice. Sagittal sections of E12.5, E13.5 and E14.5 wild-type mouse brains, from lateral to medial to lateral, ISH staining was shown for Phox2b, and for LacZ as a reference. Dashed lines represent the mid-hindbrain boundary. (a–l) LacZ and Phox2b mRNA expression at E12.5. LacZ is broadly expressed throughout the midbrain, in P1, P2 and P3, and in the hindbrain. Except for the most medial part, Phox2b is expressed in the posterior midbrain and anterior hindbrain. (m–x) LacZ and Phox2b mRNA expression at E13.5, and (y–jj) at E14.5. (kk) Schematic overview of a sagittal mouse brain at E14.5, depicting several neuronal fields in the midbrain area (red box). C, caudal; R, rostral; FB, forebrain; MB, midbrain; HB, hindbrain; RN, red nucleus; OM, oculomotor complex.

doi:10.1371/journal.pone.0052118.g001

Figure 1. Expression of Phox2b and LacZ in the midbrain of wild-type mouse embryos. Sagittal sections of E12.5, E13.5 and E14.5 wild-type mouse brains, from lateral to medial to lateral. ISH staining is shown for Phox2b, and for LacZ as a reference. Dashed lines represent the mid-hindbrain boundary. (a–l) LacZ and Phox2b mRNA expression at E12.5. LacZ is broadly expressed throughout the midbrain, in P1, P2 and P3, and in the hindbrain. Except for the most medial part, Phox2b is expressed in the posterior midbrain and anterior hindbrain. (m–x) LacZ and Phox2b mRNA expression at E13.5, and (y–jj) at E14.5. (kk) Schematic overview of a sagittal mouse brain at E14.5, depicting several neuronal fields in the midbrain area (red box). C, caudal; R, rostral; FB, forebrain; MB, midbrain; HB, hindbrain; RN, red nucleus; OM, oculomotor complex.

Combined ISH-immunohistochemistry

ISH on fresh frozen sections was performed as described [16,17]. After termination of the alkaline phosphatase coloring reaction of the ISH, slides were washed in 1x TBS, incubated in 0.3% H2O2 in 1x TBS for 30 min, washed again, blocked with 4% FCS in 1x TBS, washed again and incubated overnight at 4°C with rabbit anti-TH (Pel-Freez, Arkansas, 1:1000) in 1x TBS, incubated for 1h with avidin-biotin-peroxidase reagent mix (ABC Elite kit, Vector Laboratories) in 1x TBS. After this, slides were washed again, and stained with 3,3'-diamino-benzidine (DAB) until a maximum of 10 minutes. Color reaction was stopped by washing with water, slides were dehydrated with ethanol and mounted with Entellan (Merck).

Immunohistochemistry

Embryos were directly after isolation incubated in 4% para-formaldehyde (PFA) in 1x PBS at 4°C for at least 3 h or overnight, followed by cryoprotection in 30% sucrose solution in 1x PBS. After this, embryos were frozen on dry ice. For immunohistochemistry (IHC), sections were washed twice for 5 min in 1x TBS, blocked in 4% fetal calf serum (FCS) in 1x TBS for 30 min, and
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One-step RT PCR
Total RNA was purified from MN9D cells [19] or Pitx3-GFP/+ FACS-sorted neurons, using Trizol according to manufacturer’s protocol (Invitrogen). Gene expression levels were determined using a One-step RT-PCR kit (Qiagen). We used 1 ng total RNA from MN9D per 20 μL PCR reaction. Samples were separated on 1.5–2.0% agarose gels, and gels were scanned using a FLA-5000 imaging system (Fuji). Primers used: Phox2a forward 5'-GCTTTCTTAGGAACAGGGATC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 226 bp); Slc18a2 forward primer 5'-AGCTGAATAGCTCCAATCCAAG (product size 259 bp). Primers used: Phox2b forward primer 5'-GGCTCTTCCCCTCTAGTGTC, reverse primer 5'-GCGCTCTTCCCCCTCTAGTGTC (product size 228 bp); Vmat2 forward primer 5'-CAAGAGTTGGAGAGGGTC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 226 bp); Slc18a2 forward primer 5'-GCTATGCCTTCCTGCTGATC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 228 bp); Lmx1b forward primer 5'-GCTTTCTTAGGAACAGGGATC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 228 bp); Vmat2 forward primer 5'-CAAGAGTTGGAGAGGGTC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 226 bp); Slc18a2 forward primer 5'-GCTATGCCTTCCTGCTGATC, reverse primer 5'-CTTTGCTCTCGTCGTCC (product size 228 bp). 

Results

Phox2b is expressed in developing mdDA neurons
Since Phox2a and Phox2b are paralogues and it is suggested that Phox2a and Lmx1b can regulate Phox2a [4,12,13], we were interested whether Phox2b is expressed in a similar, specific pattern in the dorsal-caudal mdDA neuronal field as we previously found for Phox2a (unpublished data). Therefore, we analyzed the expression pattern of Phox2b by means of in situ hybridization (ISH) on E12.5, E13.5 and E14.5 sagittal sections of wild-type (C57Bl/6J) embryos, and compared this to Lmx1b expression (Fig. 1). At E12.5, Phox2b was clearly expressed in the posterior midbrain, and anterior hindbrain (Fig. 1G–L), except for the most medial sections, where no expression was detected (Fig. 1i). In the lateral areas, where Phox2b was highly expressed, this expression overlapped with the caudal domain of Lmx1b. At E13.5 and E14.5, Phox2b was also expressed in the most medial sections (Fig. 1S–X, EE–JJ). When comparing with Lmx1b expression, in both developmental stages a clear overlap of both Phox2b and Lmx1b expression fields was observed. In order to assess the possibility that Phox2b is involved in developing mdDA neurons, we compared Phox2b expression to Th. At E12.5, Phox2b only partially overlapped with Th (Fig. 2A–D), whereas more laterally, Phox2b was expressed more caudally and outside the Th domain (Fig. 2B,C,e,F). At E13.5, major overlap of Phox2b with the dorsal-caudal Th domain was observed, a pattern that was confirmed at E14.5 (Fig. 2g–r). In addition, by using Phox2b ISH analysis

were washed again. When incubating with sheep anti-TH, blocking was performed in 5% normal donkey serum in 1x TBS. Slides were incubated in primary antibody in THZT (50 mM Tris-HCl pH 7.6, 0.5 M NaCl, 0.5% Triton) at 4°C overnight, washed 3x with 1x TBS for 5 min and incubated for 1 h with secondary antibody in THZT at room temperature. Slides were washed three times in 1x PBS for 10 min and mounted using FluorSave (Calbiochem, Darmstadt). Antibodies used: rabbit anti-TH (Pel-Freez, Arkansas, 1:1000), sheep anti-TH (Millipore, 1:500), mouse anti-b-GAL (Promega, 1:300), rabbit anti-b-GAL (Cappel, 1:1000). Secondary antibodies: goat anti-rabbit Alexa-Fluor-488, donkey anti-sheep Alexa-Fluor-488, goat anti-mouse Alexa-Fluor-555, goat anti-rabbit Alexa-Fluor-555, all 1:1000 (Invitrogen).

Fluorescence-activated cell sorting (FACS)
The micro-dissected mdDA region of several Pitx3-GFP/+ embryos was dissociated using a Papain dissociation system (Worthington) and cells were sorted on a Cytopeia Influx Cell sorter. Sort gates were set on forward scatter versus side scatter (live cell gate), on forward scatter versus pulse width (elimination of clumps) and on forward scatter versus fluorescence channel 1 (520/38 filter; GFP fluorescence). Cells were sorted (98% purity) using a 100 um nozzle at a pressure of 15 PSI with an average speed of 7000 cells/second and collected in Trizol reagent (Invitrogen). [15].

One-step RT PCR
Total RNA was purified from MN9D cells [19] or Pitx3-GFP/+ FACS-sorted neurons, using Trizol according to manufacturer’s protocol (Invitrogen). Gene expression levels were determined using a One-step RT-PCR kit (Qiagen). We used 1 ng total RNA from MN9D per 20 μL PCR reaction. Samples were separated on 1.5–2.0% agarose gels, and gels were scanned using a FLA-5000 imaging system (Fuji). Primers used: Phox2a Forward primer 5’-GCTTTCTTAGGAACAGGGATC, Phox2a Reverse primer 5’-GGCTCTTCCCCTCTAGTGTC, Phox2b Reverse primer 5’-GCAAGAGTTGGAGAGGGTC, Phox2b Forward primer 5’-CAAGAGTTGGAGAGGGTC, Phox2b Forward primer 5’-GCTATGCCTTCCTGCTGATC, reverse primer 5’-CTTTGCTCTCGTCGTCC (product size 228 bp); Slc18a2 Forward primer 5’-GCTTTCTTAGGAACAGGGATC, reverse primer 5’-GGCTCTTCCCCTCTAGTGTC (product size 228 bp); Vmat2 reverse primer 5’-AGCTGAATAGCTCCAATCCAAG (product size 259 bp). Primers used: Phox2b forward primer 5’-GCTTTCTTAGGAACAGGGATC, reverse primer 5’-GGCTCTTCCCCTCTAGTGTC (product size 228 bp); Vmat2 forward primer 5’-GCTATGCCTTCCTGCTGATC, reverse primer 5’-CTTTGCTCTCGTCGTCC (product size 226 bp); Slc18a2 Forward primer 5’-GCTTTCTTAGGAACAGGGATC, reverse primer 5’-GGCTCTTCCCCTCTAGTGTC (product size 228 bp); Vmat2 reverse primer 5’-AGCTGAATAGCTCCAATCCAAG (product size 259 bp).
domains. However, close to the expression domains of Lmx1b has been analyzed before [26,27], and we confirmed in adjacent sections, no expression in the (dorsal) raphe nucleus (DR), the parabrachial and posterior midbrain, nucleus of the trigeminal nerve (V), which is in the same domain as the PB and PSV but represents a different set of neurons. In the anterior hindbrain and posterior midbrain, Phox2b is expressed in neurons of the trochlear and oculomotor nuclei (OM). Phox2b is not expressed in neurons of the VTA or SNc. (c–e) Th is expressed in the periaqueductal gray (F), the retrorubral field (I) and the VTA and SNc. Some Th positive cells are located near the OM nuclei, but not in the same domain (L.U. C, caudal; R, rostral).

doi:10.1371/journal.pone.0052118.g003

Figure 3. Phox2b transcript is absent in mdDA neurons at P14. ISH analysis of Lmx1b, Th and Phox2b in postnatal day 14 (P14) coronal mid- and hindbrain tissue. (a–p) Lmx1b expression in the hindbrain (pons) in the dorsal raphe nucleus (DR), the parabrachial nucleus (PB) and the principal sensory trigeminal nucleus (PSV). In the anterior hindbrain, Lmx1b is expressed in the superior central raphe nucleus (CS), and more rostrally, in the posterior midbrain, in mdDA neurons of the ventral tegmental area (VTA). Most rostrally, in the anterior midbrain, expression is observed in mdDA neurons of the substantia nigra pars compacta (SNc) and in the supramammillary nucleus (SuM). (b–q) In the pons, Phox2b is expressed in the motor nucleus of the trigeminal nerve (V), which is in the same domain as the PB and PSV but represents a different set of neurons. In the anterior hindbrain and posterior midbrain, Phox2b is expressed in neurons of the trochlear and oculomotor nuclei (OM). Phox2b is not expressed in neurons of the VTA or SNc. (c–e) Th is expressed in the periaqueductal gray (F), the retrorubral field (I) and the VTA and SNc. Some Th positive cells are located near the OM nuclei, but not in the same domain (L.U. C, caudal; R, rostral).
Figure 4. Phox2b is present in TH positive and Pitx3 FAC-sorted neurons. (a–c) Sagittal sections of E14.5 wild-type mouse brain, from medial to lateral. TH protein staining is shown as a marker for the mdDA neuronal field (brown staining). Dashed lines represent the mid-hindbrain boundary. Phox2b mRNA (purple staining) is selectively expressed in the caudal midbrain and rostral hindbrain. (a'–b') Medially, in the dorsal-caudal mdDA neuronal field, most Phox2b positive cells co-express TH protein (arrowheads). (d–f) E12.5 Phox2b heterozygous LacZ mutant mouse midbrain sections (Phox2b-LacZ/Wt) showing TH and bGAL co-expression in the dorsal-caudal midbrain. Dashed lines represent the mid-hindbrain boundary. (d'–f') Higher magnifications showing co-expression of TH and bGAL in the same cell (arrowhead). (g–l) E14.5 Phox2b-LacZ/Wt midbrain sections showing TH (g–h), and bGAL expression (i–j), and co-expression of both proteins according to overlay images (k–l). (g'–l'). Higher magnifications demonstrate that TH co-localizes with bGAL in the dorsal-caudal midbrain, confirming the in situ hybridization data (a–c). (m) A scatterplot showing the distribution of GFP-positive Pitx3-GFP/+ neurons FAC-sorted from micro-dissected E14.5 mouse midbrains, and compared with wild-type reference tissue. Only GFP-positive (pink cloud) neurons were used for mRNA isolation. (n) One-step RT-PCR for Phox2a, Phox2b, Vmat2, and Tbp as RT-PCR control, on RNA isolated from MN9D cells, and FAC-sorted Pitx3-GFP/+ neurons, confirming the presence of Phox2b, and Phox2a transcripts in mdDA neurons. C, caudal; R, rostral; Wt, wild-type; for an embryonic midbrain reference picture, see figure 1kk.

In addition, we used RNA from MN9D cells as a positive control, (E14.5) (Fig. 4m), and subjected this material to one-step RT PCR. In addition, we used RNA from MN9D cells as a positive control, since Phox2b is expressed in this dopaminergic cell line. We analyzed transcript levels of Phox2b, Phox2a and Vmat2, the latter as a positive control. Tbp was taken along as a loading and PCR reference. We confirmed expression of all transcripts in MN9D cells, where the two Phox2-genes were highly expressed, when compared to Vmat2 (Fig. 4n). Importantly, also in Pitx3-GFP/+ neurons, Phox2b transcript was present, in comparable levels as Vmat2.

To further validate that Phox2b is expressed in mdDA neurons, we used isolated RNA from FAC-sorted Pitx3-GFP/+ neurons (E14.5) (Fig. 4m), and subjected this material to one-step RT PCR. Since a small group located directly posterior of the MHB clearly the latter as a positive control. Tbp was taken along as a loading and PCR reference. We confirmed expression of all transcripts in MN9D cells, where the two Phox2-genes were highly expressed, when compared to Vmat2 (Fig. 4n). Importantly, also in Pitx3-GFP/+ neurons, Phox2b transcript was present, in comparable levels as Vmat2.

In conclusion, by using several approaches, we showed that Phox2b is present in a dorsal-caudal subset of developing mdDA neurons.

Caudal TH expression is affected in Phox2b mutants

The expression of Phox2b in mdDA neurons implicates a role in mdDA neuronal development. To investigate this into more detail, we analyzed TH expression in the developing midbrain of Phox2b null mutant embryos.

Analysis of E12.5 wild-type and Phox2b-LacZ/LacZ (knock-out/knock-in) tissue (Fig. 5a–f) revealed a subtle decrease in TH expression in the most dorsal-caudal TH expression domain of the mdDA neuronal field (Fig. 5c, arrowhead). Within this region, bGAL showed a high expression level (Fig. 5d'–f'). Furthermore, when comparing Phox2b heterozygous with Phox2b-LacZ/LacZ embryonic brains, the restricted loss of dorsal-caudal TH expression was confirmed (Fig. 5, compare g–l with m–r). In the heterozygous dorsal-caudal mdDA neuronal field, a select group of cells was observed, that clearly co-expressed TH and bGAL (Fig. 5g'–l'). In the absence of Phox2b, this subset displayed lower levels of TH, as was shown by decreased co-localization of b-GAL with TH (Fig. 5m'–r'). This loss appeared to be midbrain specific, since a small group located directly posterior of the MHB clearly still co-expressed TH and b-GAL in Phox2b-LacZ/LacZ embryos (Fig. 5n,o,q,r, arrowheads). In conclusion, the loss of Phox2b results in a decreased TH expression in a caudal subset of mdDA neurons specifically (Fig. 5g'–l').

Similar results were observed in the E14.5 Phox2b null mutant. A subtle reduction of TH positive neurons was observed in the Phox2b area in the medial midbrain (Fig. 6a–b',f–g', arrowheads). Interestingly, another mild deficit was observed, in an area that does not express Phox2b at this stage at all. The entire mdDA neuronal field, from lateral to medial, displayed a subtle reduction in TH expression in the rostral (diencephalic) mdDA neuronal field (Fig. 6c–d',h–i', arrowheads).
Taken together, Phox2b is expressed in a subset of neurons that expresses mild levels of TH, in the dorsal-caudal midbrain and in a small group of TH positive cells located near the isthmic organizer. This isthmic group is spared whereas the more anterior located TH positive cells lose TH expression as a consequence of Phox2b ablation, suggesting that Phox2b is involved in the correct specification of this small subset of mdDA neurons.

**Discussion**

Phox2a, and its parologue Phox2b, play important roles in the development of branchiomotor and visceromotor neurons in the ventral hindbrain and are both expressed in noradrenergic centers [6,10,11,20,25,28–30]. In the main noradrenergic center, the locus ceruleus, both genes can fully compensate for each other [25]. In addition, both factors are also specifically expressed in...
midbrain oculomotor neurons and in trochlear neurons, where Phox2a expression precedes that of Phox2b [4,6,8,12]. Since Phox2a displayed an expression pattern that was largely overlapping with the dorsal-caudal mdDA neuronal field, in an earlier study (unpublished data), we suspected a role for Phox2a,
and Phox2b, in the development of these neurons, in addition to the proposed function in oculomotor neuron (OMN) development. Indeed, our current analysis of Phox2b provides insight in a role of this gene in the development of a subset of mdDA neurons.

**Phox2b is temporally expressed in a subset of developing mdDA neurons**

Our detailed analysis of the expression pattern of Phox2b during several developmental stages, confirms that Phox2b is expressed in a specific pattern overlapping with the dorsal-caudal mdDA neuronal field. However, in this midbrain area, motorneurons are formed as well. Moreover, a known marker for motorneurons, Isl1, is expressed in this area in a similar expression pattern as Phox2a and Phox2b [8,31] (online expression databases). The Phox2 genes are involved in OM development, and since a recent paper suggested a role for Phox2a together with Lmx1b in the generation and control of OMNs and red nucleus neurons (RNNs) [4], it is likely that Phox2a and Phox2b expressing cells in the midbrain are (inter-)intermingled with visceral and somatic motorneurons.

In this study, we aimed to investigate in detail whether Phox2b is truly confined to OMNs, or in addition plays a role in mdDA neuronal development. Intriguingly, we clearly showed that many Phox2b expressing cells in the midbrain area, co-express TH protein, which was confirmed in Phox2b-LacZ positive neurons, and by RT-PCR on FAC-sorted Pitx3-positive neurons. In contrast to this, analysis of postnatal mouse brains revealed that Phox2b is not expressed in mature mdDA neurons. We confirmed expression in known sites of Phox2b, in the ventral hindbrain, and in the OM. However, no expression rostral to this nucleus was observed. Since Phox2b is clearly expressed in the developing mdDA neuronal field, the lack of expression in the postnatal mdDA system indicates a role for Phox2b in a small but specific subset of the mdDA domain during development of these neurons.

**References**


