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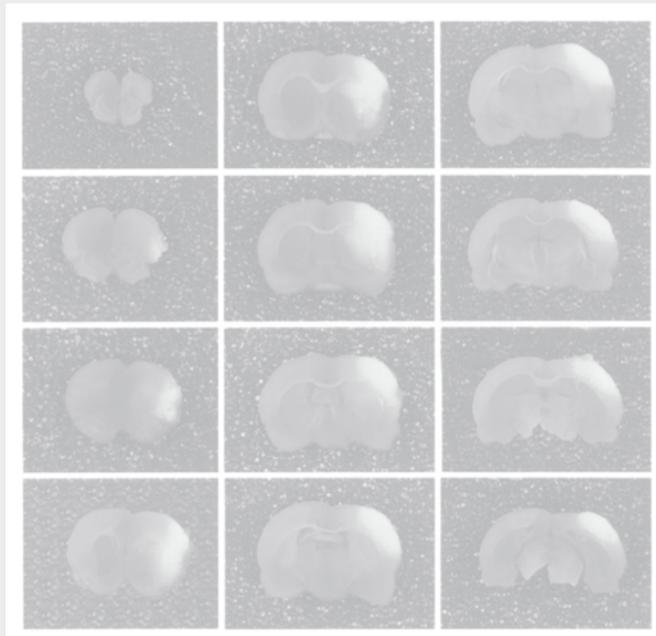
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Chapter 3

A Comparative Study of the Effects of Two Nitric Oxide Synthase Inhibitors and Two Nitric Oxide Donors on Temporary Focal Ischemia in the Wistar Rat



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ABSTRACT

A critical review of the literature indicates that the effects of nitric oxide synthase (NOS) inhibitors on focal cerebral ischemia are contradictory. In this experiment the authors methodically examined the dose-dependent effects of two NOS inhibitors and two NO donors on cortical infarction volume in an animal model of temporary focal cerebral ischemia simulating potential ischemia during neurovascular interventions. Ninety-two Wistar rats underwent 3 hours of combined left middle cerebral artery and bilateral common carotid artery occlusion after having been anesthetized with 1% halothane. A nonselective NOS inhibitor, N^G-nitro-L-arginine-methyl-ester (L-NAME), and two NO donors, 3-morpholinosydnonimine hydrochloride and NOC-18, DETA/NO, (Z)-1-[2(2-aminoethyl)-N-(2-ammonioethyl)amino]diazene-1,2-diolate, were administered intravenously 30 minutes before ischemia was induced. A selective neuronal NOS inhibitor, 7-nitroindazole (7-NI), was administered intraperitoneally in dimethyl sulfoxide (DMSO) 60 minutes before ischemia was induced. Two ischemic control groups, to which either saline or DMSO was administered, were also included in this study. Seventy-two hours after flow restoration, the animals were perfused with tetrazolium chloride for histological evaluation. Cortical infarction volume was significantly reduced by 71% in the group treated with 1 mg/kg L-NAME when compared with the saline-treated ischemic control group ($27.1 \pm 37 \text{ mm}^3$ compared with $92.5 \pm 26 \text{ mm}^3$, $p < 0.05$). The NOS inhibitor 7-NI significantly reduced cortical infarction volume by 70% and by 92% at doses of 10 and 100 mg/kg: $35.2 \pm 32 \text{ mm}^3$ ($p < 0.05$) and $9 \pm 13 \text{ mm}^3$ ($p < 0.005$), respectively, when compared with the DMSO-treated ischemic control group ($119 \pm 43 \text{ mm}^3$). There was no significant difference between the saline-treated and DMSO-treated ischemic control groups. Treatment with NO donors did not significantly alter cortical infarction volume. These results support an important role for NO in ischemic neurotoxicity and indicate that neuronal NOS inhibition may be valuable in reducing cortical injury in patients suffering temporary focal cerebral ischemia during neurovascular procedures.

INTRODUCTION

During cerebral ischemia, excitatory amino acid release activates the N-methyl-D-aspartate (NMDA) receptor, which mediates a transmembrane influx of calcium with subsequent deleterious effects^{7,43,44}. This rise in intracellular calcium activates calcium-dependent enzymes including neuronal (n) and endothelial (e) nitric oxide synthase (NOS)⁴⁵. Inducible NOS, which is calcium independent, requires protein synthesis to be expressed and, thereby, slowly produces large amounts of nitric oxide (NO) over a long period of time during the postischemic phase¹⁰. Excessive concentration of NO may mediate the majority of NMDA toxicity⁴⁵. The half-life of NO in an oxygen-containing environment is short, in the order of seconds²⁹, which complicates direct measurements of NO levels in ischemic tissue. Accordingly, information on the mechanisms and actions of NO has been largely derived from studies in which NOS inhibitors have been used *in vivo*. Published reports have shown contradictory results, with both cerebroprotective^{2,50} and detrimental³⁰ effects of NOS inhibition. Combining these results has led to a proposed dose-dependent dual role for NO^{1,10}. Selective inhibitors of nNOS have demonstrated more consistent protective results²⁴, suggesting that this dualism is related to the different isoforms of NOS. However, even the selectivity and mechanisms of action of NOS antagonists have been questioned^{40,51}. With the availability of genetically altered animals, it has been possible to by-pass possible nonspecific side effects of pharmaceutical intervention. nNOS "knockout" animals developed smaller infarctions after middle cerebral artery (MCA) occlusion when compared with the wild type^{13,20}. The successful use of NO donors in acute myocardial ischemia has raised questions about its similar efficacy in focal cerebral ischemia. In a rat model of focal cerebral ischemia, NO donors were protective if hypotension was avoided⁵³. Alternately, in cortical cultures, NO donors were neurotoxic¹². A comparison of the effects of different NO modulators *in vivo* between different investigations is complicated by differences in methodology, including animal model, anesthetic agent, occlusion technique, duration of ischemia, and drug dosing. The objective of this study was to examine rigorously the dose-dependent effects of two NOS inhibitors and two NO donors on cortical infarction volume in a model of temporary focal cerebral ischemia that simulates possible ischemia during neurovascular procedures.

MATERIALS AND METHODS

Following review and approval of the protocol by our institutional Animal Care and Use Committee, 92 adult male Wistar rats were administered halothane in a mixture of oxygen and air through a face mask at 1.5% during the surgical procedure and 1% during the occlusion period. Subcutaneous glycopyrrolate was administered preoperatively

at 4 mg/kg to reduce respiratory secretions. Core body temperature was continuously monitored at the beginning of the surgical preparation and throughout the experiment by using a rectal probe. This rectal probe was connected to an infrared heating lamp that maintained both body and head temperature at $37 \pm 0.5^\circ\text{C}$ throughout the experiment. Polyethylene catheters (PE- 50) were inserted into the right femoral artery and vein to monitor mean arterial blood pressure (MABP) and arterial blood sampling (pH, PaCO_2 , PaO_2 , and serum glucose). These physiological parameters were measured at the beginning of the surgical preparation, 15 minutes before ischemia was induced, and at the completion of the ischemia experiment.

Model of Focal Cerebral Ischemia

The original technique, as described by Tamura, et al.⁴⁷, was modified for our experiments to increase the severity and reliability of the ischemic model⁸. A ventral midline incision was made for exposure of both common carotid arteries (CCAs). The contralateral (right) CCA was permanently ligated using a No. 3.0 silk suture. The ipsilateral (left) CCA was temporarily occluded (for 3 hours) using a Mayfield microaneurysm clip. A skin incision was made between the left outer canthus and the tragus. The temporal muscle was deflected anteriorly and a portion of the left zygomatic arch was removed. Care was taken to avoid damaging the facial nerve. After anterior and downward retraction of the musculature, the mandibular nerve was identified and followed back to the foramen ovale. Using a high-speed air drill, a 3- to 4-mm craniectomy was made just anterior and superior to the foramen ovale. The dura was opened with a sharp needle and the MCA was freed from arachnoid. The portion of the left MCA that crosses the olfactory tract was temporarily occluded (for 3 hours) using a No. 3 Sundt arteriovenous malformation microclip.

Experimental Groups

The animals were divided into 16 groups as follows. The nonspecific NOS inhibitor, N^G -nitro-L-arginine-methyl-ester (L-NAME), was administered intravenously 30 minutes before MCA occlusion at doses of 0.1 (six rats), 1 (five rats), 10 (five rats), and 30 (six rats) mg/kg. A selective nNOS inhibitor, 7-nitroindazole (7-NI), was administered intraperitoneally 60 minutes before MCA occlusion at doses of 0.1 (five rats), 1 (six rats), 10 (six rats), and 100 (six rats) mg/kg. Both NO donors, 3-morpholinosydnonimine (SIN-1) and NOC-18, DETA/NO, (Z)-1-[2(2-aminoethyl)-N-(2-ammonioethyl) amino]diazene-1-ium-1,2-diolate (DETA NONOate), were administered intravenously 30 minutes before MCA occlusion. The SIN-1 was given at doses of 0.1 (five rats), 1 (six rats), and 10 (six rats) mg/kg, and the DETA NONOate at doses of 0.1, 1, and 10 mg/kg (six rats in each group). The L-NAME, DETA NONOate, and SIN-1 were dissolved in 0.35 ml of 0.9% NaCl. Because of its poor solubility in aqueous solution, the 7-NI was dissolved in 0.35 ml dimethyl sulfoxide

(DMSO). Two groups of ischemic control animals (each group consisting of six animals) were administered 0.35 ml of either 0.9% NaCl (intravenously) or DMSO (intraperitoneally) at 30 or 60 minutes before MCA occlusion, respectively. Each solution was prepared directly before administration. Changes in the animals' cardiovascular status (heart rate and arterial blood pressure) were measured and recorded.

Histological Study

Three days (72 hours) after removal of the left MCA and CCA clips, anesthesia was again induced in the animals by using pentobarbital and the rats were intracardially perfused with a warm (37°C) 2% 2,3,5,-triphenyltetrazolium chloride solution. The rat brains were quickly removed, immersed in the 37°C 2,3,5,-triphenyltetrazolium chloride solution for 15 minutes to enhance staining, and placed in 10% buffered formaldehyde for 5 days. Twelve serial sections from each brain were cut at 1-mm intervals from the frontal pole and photographed. Photographic slides were analyzed using a computer-assisted image analyzer. Total cortical infarction volume was calculated by integrating the infarcted areas of all twelve sections (area of infarction in square millimeters x thickness of section).

Statistical Analysis

Statistical analysis was performed using analysis of variance with Scheffé's post hoc test for multiple comparisons. Differences were considered significant if the probability value was less than 0.05. The data are depicted as the mean \pm standard deviation.

Sources of Supplies and Equipment

The L-NAME was purchased from Sigma Chemical Co. (St. Louis, MO) and the 7-NI, SIN-1, and DETA NONOate from Alexis Biochemicals Corp. (San Diego, CA). The JAVA image analyzer was obtained from SPSS Inc. (Chicago, IL) and the Sundt No. 3 arteriovenous malformation microclip from Johnson & Johnson Professional, Inc. (Raynham, MA).

RESULTS

Physiological Measurements

Physiological parameters measured just before completion of the ischemia experiment for each of the 16 groups are listed in Table 1. These physiological measurements were similar to those obtained at the beginning of the experiment and just prior to induction of ischemia. The MABP decreased momentarily after the intravenous bolus administration of SIN-1 but normalized before left MCA and bilateral CCA occlusion. A slight, non-significant reduction in MABP was noted in the DETA NONOate-treated groups. Mean arterial blood pressure was significantly higher in the L-NAME groups when compared with

the saline-treated ischemic control group. In the group treated with the highest dose of 7-NI (100 mg/kg administered intraperitoneally), bradycardia occurred with heart rates decreasing to 150 to 200/minute (normal approximately 300/minute).

Table 1 Physiological parameters in rats subjected to temporary focal cerebral ischemia.

Groups (no. of rats)	Weight (g)	pH	PaCO ₂ (mm Hg)	PaO ₂ (mm Hg)	Glucose (mg/dl)	MABP (mm Hg)
L-NAME						
0.1 mg/kg (6)	413 ± 42	7.35 ± 0.04	48.1 ± 2.3	251 ± 46	194 ± 28	104 ± 8†
1 mg/kg (5)	362 ± 56	7.36 ± 0.02	51.3 ± 4.5	218 ± 54	175 ± 33	103 ± 6†
10 mg/kg (5)	390 ± 60	7.35 ± 0.03	47.0 ± 3.4	192 ± 54	174 ± 24	104 ± 8†
30 mg/kg (6)	419 ± 8	7.43 ± 0.05	38.4 ± 8.6	241 ± 18	179 ± 34	104 ± 7†
7-NI						
0.1 mg/kg (5)	349 ± 8	7.40 ± 0.02	40.7 ± 4.0	196 ± 18	159 ± 11	94 ± 6
1 mg/kg (6)	338 ± 21	7.42 ± 0.05	37.8 ± 6.2	222 ± 35	168 ± 59	94 ± 5
10 mg/kg (6)	368 ± 31	7.42 ± 0.05	44.9 ± 9.6	275 ± 54	173 ± 16	95 ± 5
100 mg/kg (6)	352 ± 12	7.43 ± 0.06	38.2 ± 6.4	247 ± 45	219 ± 64	95 ± 6
SIN-1						
0.1 mg/kg (5)	371 ± 11	7.40 ± 0.08	47.1 ± 10	265 ± 84	174 ± 22	103 ± 6
1 mg/kg (6)	348 ± 21	7.41 ± 0.08	41.5 ± 9.2	243 ± 59	161 ± 36	93 ± 9
10 mg/kg (6)	379 ± 48	7.45 ± 0.09	42.1 ± 11	224 ± 25	172 ± 22	94 ± 6
DETA NONOate						
0.1 mg/kg (6)	360 ± 24	7.47 ± 0.04	38.6 ± 5.4	239 ± 31	186 ± 32	93 ± 6
1 mg/kg (6)	334 ± 42	7.46 ± 0.06	38.1 ± 6.5	218 ± 45	166 ± 30	96 ± 7
10 mg/kg (6)	419 ± 18	7.41 ± 0.04	42.9 ± 4.3	246 ± 64	170 ± 22	90 ± 8
ischemic control						
0.9% NaCl (6)	372 ± 45	7.44 ± 0.07	38.9 ± 6.8	240 ± 60	192 ± 36	91 ± 6
DMSO (6)	342 ± 16	7.46 ± 0.04	36.6 ± 4.9	224 ± 24	158 ± 19	90 ± 6

Head and body temperature was maintained at $37 \pm 0.5^\circ\text{C}$ throughout the experiment. The parameters listed in this table are those obtained at the third measurement just before completion of the ischemia experiment. Physiological parameters obtained in the first two measurement periods at the beginning of the experimental preparation and just prior to ischemia were similar to these values and are not listed. Values are expressed as the mean \pm standard deviation. † $p < 0.05$ when compared with the ischemic control group (0.9% NaCl).

Ischemic Control Groups

Three hours of left MCA and bilateral CCA occlusion resulted in mean cortical infarction volumes of $92.5 \pm 26 \text{ mm}^3$ and $119 \pm 43 \text{ mm}^3$ in the saline- and DMSO-treated ischemic control groups, respectively (Fig. 1). There was no significant difference (Student's unpaired t-test) between these two groups. Because the infarcted area was not directly located at the craniotomy site, it was easily distinguishable from possible direct damage caused by the surgical procedure. To assess the presence and extent of edema, the ischemic hemisphere volume was compared with the hemisphere volume on the contralateral side. In the saline treated group, the mean hemisphere volume was $648.1 \pm 33 \text{ mm}^3$ for the ischemic (left) side and $621.2 \pm 29 \text{ mm}^3$ for the nonischemic (right) side. The right/left ratio (the right hemispheric volume divided by the left hemispheric volume) was 0.96 for this group. The range of the right/ left ratio was 0.96 to 1.01 in all groups studied, with no significant differences between groups.

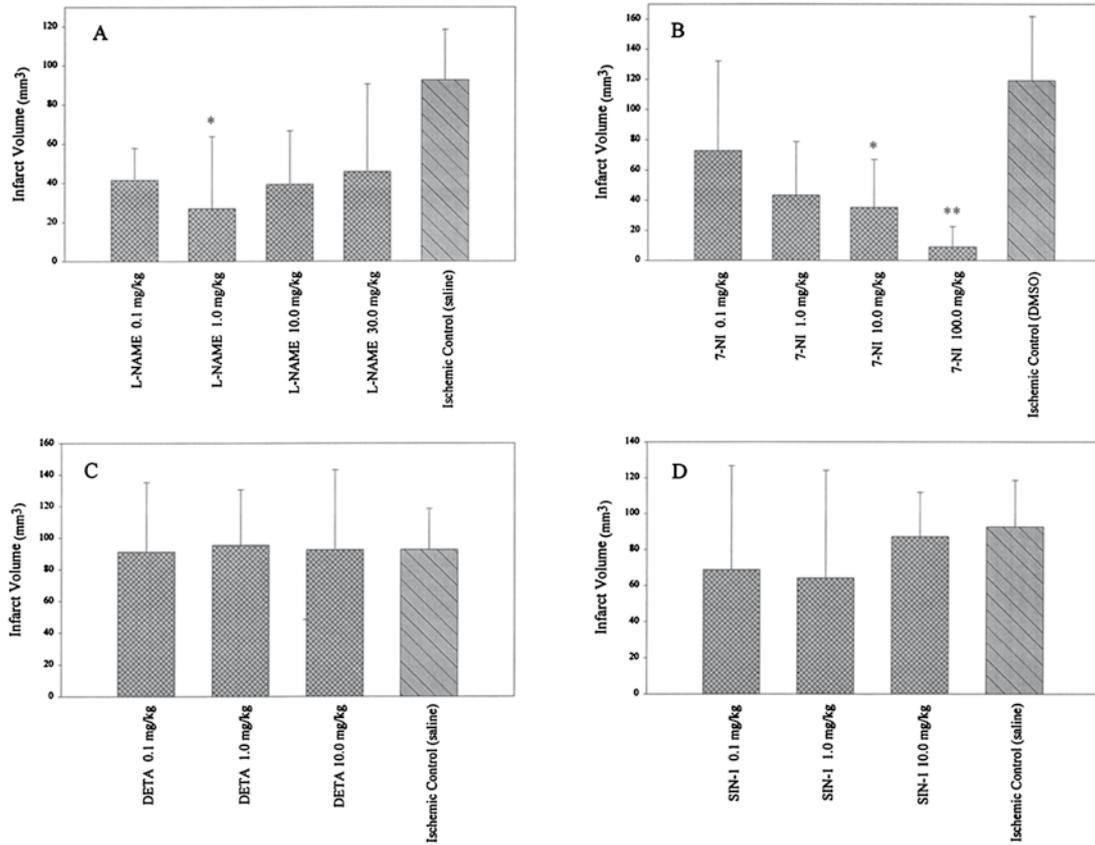


Figure 1. Bar graphs showing infarction volumes in the four drug groups (cross-hatched bars) and their respective saline- or DMSO-treated ischemic control group (slashed bars).

A: Groups treated with L-NAME, a nonselective NOS inhibitor, at four different doses. Note a subtle bell-shaped dose response with a significant decrease in infarction volume observed only at the 1-mg/kg dose. B: Groups treated with 7-NI, a selective NOS inhibitor, at four different doses. Note the uniphase dose response, with the two highest doses resulting in significant reduction in infarction volume, most notably at the 100-mg/kg dose. C: Groups treated with DETA NONOate, a NO donor, at three different doses. There was no significant alteration in infarction volume associated with any dose. D: Groups treated with SIN-1, an NO donor, at three different doses. There was no significant alteration in infarction volume in this group associated with any dose. Values are expressed as the mean \pm standard deviation for each group. * $p < 0.05$; ** $p < 0.005$.

Drug Treatment Groups

The L-NAME-Treated Groups

The mean cortical infarction volume was reduced by 50 to 71% in all four L-NAME-treated groups studied when compared with the saline-treated ischemic control group (Fig. 1A). Only in the 1 mg/kg group was this reduction significant ($27.1 \pm 37 \text{ mm}^3$ compared with $92.5 \pm 26 \text{ mm}^3$; $p < 0.05$).

The 7-NI-Treated Groups

The mean cortical infarction volume was significantly reduced in the 10 mg/kg-treated group ($35.2 \pm 32 \text{ mm}^3$, $p < 0.05$) and the 100 mg/kg-treated group ($9 \pm 13 \text{ mm}^3$; $p < 0.005$) when compared with the DMSO-treated ischemic control group ($119 \pm 43 \text{ mm}^3$ Fig. 1B). The high-dose of 7-NI (100 mg/kg) tended to potentiate anesthesia and delay recovery.

The DETA NONOate-Treated Groups

No significant difference was found between the mean cortical infarction volumes in the DETA NONOate-treated groups and the saline-treated ischemic control group (Fig. 1C).

The SIN-1-Treated Groups

The mean infarction volumes in the SIN-1-treated groups were not significantly different from those in the saline-treated ischemic control group (Fig. 1D).

DISCUSSION

Nitric oxide has been recognized as an important mediator of NMDA and hypoxic neurotoxicity¹³. Malinski, et al.³⁵, conducted the first direct NO measurement study in which a NO-sensitive microsensor was used, and their results confirmed increased levels of NO in focal cerebral ischemia and provided valuable information about the time course of NO production. In that study, a rapid increase in the NO signal was found in the parietal cortex after the onset of focal cerebral ischemia; this increase reached a semiplateau at 6 minutes and the signal decreased to below detectable levels by 60 minutes³⁵. Using a nitrite assay technique as an indirect method of measuring NO levels, Kader, et al.²⁷, found a similar pattern with a rise in nitrite levels maximizing at 5 to 10 minutes and normalizing within 60 minutes after onset of ischemia. Although of great importance, absolute values of NO production are very dependent on the local microenvironment and measurement methodology³⁵.

Nitric Oxide Synthase Inhibitors

Nitric oxide synthase inhibitors have been widely used to determine the overall effect of NO on ischemic damage. Selective inhibitors and genetically altered animals have made it possible to estimate contributions of the different isoforms of NOS to neurotoxicity and neuroprotection. Results with nonselective inhibitors such as L-NAME have shown a wide spectrum of results varying from cerebroprotection² to augmentation of ischemic damage¹⁹. Selective nNOS inhibitors and nNOS knockout animal studies have shown consistent protective results²⁴. Inhibition of NO's physiological activity may account for some

side effects. Because the head and core body temperature was maintained throughout the experiment at 36.5 to 37.5°C, the effects of the NOS antagonists on infarction size were not related to hypothermia. Vasoconstriction resulting in a rise in MABP was found in studies in which nonspecific NOS inhibitors were used, such as L-NAME³³ administered intravenously at high dosages, whereas reductions in cerebral blood flow (CBF) were observed in both nNOS-specific (7-NI) and non-specific (for example, L-NAME) inhibitors^{28,33}. Bradycardia and sedation have been reported with the use of L-NAME at higher doses³³. The 7-NI has potent antinociceptive properties³⁸ and induces central nervous system depression similar to high doses of narcotic/sedative/hypnotic agents^{15,40}. It may also inhibit eNOS and, therefore, may not be a completely selective nNOS inhibitor⁵¹. In our present study, bradycardia and sedation were seen in the group treated with an intraperitoneal injection of 100 mg/ kg of 7-NI. In the L-NAME groups studied, the MABP increased after intravenous administration of L-NAME. Brain NOS activity has been reported to diminish for at least 6 hours⁴⁸ and up to 96 hours after a single dose of L-NAME is given intravenously²⁶. The inhibitor L-NAME has been shown to be effective in reducing infarction size mainly at lower dosages and in accentuating ischemia at high dosages²⁵. This suggests that partial inhibition of nNOS from lower dosages is sufficient to acquire an optimum neuroprotective effect, but that higher doses can result in inhibition of eNOS, thereby exacerbating cerebral ischemia via vasoconstriction. Intraperitoneal 7-NI administration in rats has resulted in maximum brain nNOS inhibition at 30 minutes and in complete recovery at 24 hours³². Significant reductions in mean cortical infarction volumes were seen in two of the 7-NI groups (10 and 100 mg/ kg administered intraperitoneally in DMSO). A DMSO treated ischemic control group was added to the 7-NI-treated groups because, as an HO• scavenger, DMSO can reduce oxidant injury³. The mean cortical infarction volume in this DMSO-treated group was $119 \pm 43 \text{ mm}^3$, which was not significantly different (Student's unpaired t-test) from that found in the saline-treated ischemic control group ($92.5 \pm 26 \text{ mm}^3$). Overall, the maximum protective effect was seen in the 100-mg/kg 7-NI group, in which a mean cortical infarction volume reduction of 92% was seen. The enhancement and prolongation of the anesthetic effect observed in this group could have contributed to its protective effects.

Nitric Oxide Donors

The anti-schemic effects of nitrovasodilators in coronary artery disease have been recognized for more than 100 years⁶ and are still used to treat acute coronary syndromes. In focal cerebral ischemia NO donors appeared to reduce ischemic brain damage if hypotension was pharmacologically avoided by coadministration of an ionotrope²⁵. Cyclic guanosine monophosphate-mediated vasodilation and reduction in platelet aggregation³⁷, as well as direct downregulation of the NMDA receptor by reacting with the redox modulatory site³¹, have been proposed to contribute to the cerebroprotective effect of NO.

No protective effect was found *in vitro*, however. The NO donor sodium nitroprusside showed concentration-dependent cell death curves similar to those of NMDA¹². A dual role for NO was proposed in which neuronal NO overproduction played an important role in the development of ischemic damage, whereas endothelial and perivascular NO could protect against ischemia by increasing regional CBF and preventing platelet aggregation^{1,10}. One of the two NO donors used in this study, SIN-1, is routinely used in interventional cardiology as a coronary artery bolus injection¹⁸. In a study by Shukla, et al.⁴⁶, SIN-1 was shown to cross the blood-brain barrier in rats. Nitric oxide formation from SIN-1 occurs spontaneously and does not require the presence of cysteine¹⁷. Two factors were identified as influencing NO release: PO₂ and pH⁵. During cerebral ischemia, oxidative capacity appeared to be high enough to guarantee NO release⁵. With the exception of NO, superoxide was shown to be a product of the SIN-1 oxidation²². Superoxide generation was also described with purified nNOS at suboptimal L-arginine concentrations²¹. By preventing superoxide formation, L-arginine infusion could provide additional protection to the previously discovered beneficial effects on regional CBF³⁹. Together superoxide and NO form peroxynitrite, which is the major contributor to NO and superoxide toxicity. Care should be taken in handling sydnonimines such as SIN-1 because they are highly susceptible to oxygen and light¹⁶. For this reason all our SIN-1 solutions were prepared directly before use and administered as an intravenous bolus. The half-life of NO in air-saturated buffer was calculated to be 6 seconds²⁹, indicating that NO formation is the main determinant of NO levels. Noack and Feelisch⁴¹ studied time-dependent formation of various metabolites of SIN-1 and their velocity of NO liberation, revealing the half-life of SIN-1 to be approximately 150 minutes. The SIN-1 metabolite SIN-1A reached a peak concentration at approximately 75 minutes with a half-life of approximately 300 minutes. The initial NO liberation velocity for SIN-1A was measured to be four times higher than that for SIN-1. Accounting for each relative NO liberating capacity, the overall half-life for NO donation is approximately 230 minutes and the maximum NO liberation is at approximately 50 minutes⁴¹. In our study a temporary reduction in MABP was seen after intravenous administration. Before MCA and bilateral CCA occlusion, the MABP had returned to baseline levels without the use of vasopressor agents. The protective effect of a NO donor may be difficult to determine when a vasopressor is used simultaneously to maintain blood pressure by some investigators^{52,53}. Using a permanent MCA occlusion model with phenylephrine-induced hypertension, Drummond and associates¹⁴ showed a reduction in brain regions in which local CBF is equal to or lower than levels that may result in neuronal death. Using permanent MCA occlusion in spontaneously hypertensive rats, Maiese, et al.³⁴, found no significant difference in infarction volume when the MABP was elevated using phenylephrine. One possible explanation for this discrepancy may be that the protective effect of phenylephrine depends on improvement in collateral flow. Collateral circulation is less developed in spontaneously hypertensive rats⁹, which may

explain the lack of efficacy of phenylephrine. In this present study, SIN-1 without vasopressor did not significantly alter mean cortical infarction volume. The other NO donor used in this study, DETA NONOate, is a zwitterionic polyamine/NO adduct²³ that releases two molecules of NO per molecule of DETA NONOate⁴⁹. The half-life for DETA NONOate was found to be 3400 minutes²³; therefore, it is more stable and longer acting than SIN-1. Although it is longer acting and releases higher concentrations of NO than SIN-1, DETA NONOate also did not significantly affect mean cortical infarction volume in our study. In our model of focal cerebral ischemia in the Wistar rat, NO donors resulted in unchanged mean cortical infarction volumes throughout the concentration range used. However, both a nonspecific (L-NAME) and an nNOS specific (7-NI) inhibitor reduced infarction volume, which suggests that excessive NO production is detrimental. However, the effect of NOS inhibitors on other physiological roles of NO should not be overlooked^{11,25}. Pajewski and associates⁴² studied the effect of L-NAME and 7-NI on anesthesia, discovering that inhibition of the NO pathway decreased levels of consciousness, augmenting sedation, analgesia, and anesthesia. Potencies for 7-NI were found to vary between mice and rats in a study by Moore, et al.³⁸ In that study, the authors showed that there was inhibition of nNOS without alterations in MABP in a dose range of 10 to 80 mg/kg. The lack of effect on MABP would indicate that there would be minimal or no effect on eNOS. Traystman and colleagues⁴⁸ showed that a 20-mg/kg intravenous injection of L-NAME produced the same amount of NOS enzyme inhibition (> 70%) but the half-lives varied widely among cats, dogs, and pigs. These observations suggest that extrapolations between species can lead to inaccurate dosing. The most effective dose for 7-NI found in our study was 100 mg/kg administered intraperitoneally, although a significant reduction in infarction volume was also demonstrated at 10 mg/kg. Dalkara, et al.¹⁰, found 25- and 50-mg/kg doses to be effective in reducing infarction volume by 25 to 27%.

The efficacy of different therapies appears to depend on the severity of the ischemic insult, time of onset, dosing of therapy, and the patient's general vascular condition. Marguill, et al.³⁶, described the time course of glutamate concentrations during temporary MCA and bilateral CCA occlusion. Because the glutamate surge occurred minutes after occlusion and lasted for only approximately 80 minutes, the therapeutic window for intervention was found to be very short³⁶. Direct measurement of NO production showed a similar pattern³⁵. Atherosclerotic lesions tended to modify the response of human (coronary) arteries to vasoactive substances⁴ that curtailed the efficacy of the therapy. Dosing appeared to be a major determinant of the overall effect, especially in the case of L-NAME, a low dose of which (0.1 mg/kg) has been shown to be protective² with a minimum effect on MABP and a high dose (10 mg/kg) was more often detrimental⁵². Before extrapolations can be made to human pathophysiological states, more information should be obtained about the activation of pathophysiological events in humans. The therapeutic window for NOS inhibition appears to be short³⁵, limiting its efficacy in cases

of stroke. In temporary arterial occlusion during vascular surgery, drug therapy could be instituted before occlusion to prevent ischemic brain damage. The results of this experiment indicate that NOS inhibition may be a valuable neuroprotective intervention during neurovascular or endovascular procedures.

REFERENCES

1. Ashwal S, Cole DJ, Osborne TN, et al: Dual effects of L-NAME during transient focal cerebral ischemia in spontaneously hypertensive rats. *Am J Physiol* 267:H276–H284, 1994
2. Ashwal S, Cole DJ, Osborne TN, et al: Low dose L-NAME reduces infarct volume in the rat MCAO/reperfusion model. *J Neurosurg Anesthesiol* 5:241–249, 1993
3. Beckman JS, Beckman TW, Chen J, et al: Apparent hydroxyl radical production by peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc Natl Acad Sci USA* 87:1620–1624, 1990
4. Berkenboom G, Unger P, Fontaine J: Atherosclerosis and responses of human isolated coronary arteries to endothelium-dependent and -independent vasodilators. *J Cardiovasc Pharmacol* 14 (Suppl 11):S35–S39, 1989
5. Bohn H, Schönafinger K: Oxygen and oxidation promote the release of nitric oxide from sydnonimines. *J Cardiovasc Pharmacol* 14 (Suppl 11):S6–S12, 1989
6. Brunton TL: Use of nitrite of amyl in angina pectoris. *Lancet* 2:97–98, 1867
7. Choi DW: Glutamate neurotoxicity and diseases of the nervous system. *Neuron* 1:623–634, 1988
8. Coert B, Anderson RE, Meyer FB: The effects of intermittent versus continuous vessel occlusion in a revalidated model of focal cerebral ischemia in the Wistar rat. *J Cereb Blood Flow Metab* 17 (Suppl 1):S305, 1997 (Abstract)
9. Coyle P: Different susceptibilities to cerebral infarction in spontaneously hypertensive (SHR) and normotensive Sprague–Dawley rats. *Stroke* 17:520–525, 1986
10. Dalkara T, Yoshida T, Irikura K, et al: Dual role of nitric oxide in focal cerebral ischemia. *Neuropharmacology* 33:1447–1452, 1994
11. Dawson VL, Dawson TM: Nitric oxide actions in neurochemistry. *Neurochem Int* 29:97–110, 1996
12. Dawson VL, Dawson TM, Bartley DA, et al: Mechanisms of nitric oxide-mediated neurotoxicity in primary brain cultures. *J Neurosci* 13:2651–2661, 1993
13. Dawson VL, Kizushi VM, Huang PL, et al: Resistance to neurotoxicity in cortical cultures from neuronal nitric oxide synthase-deficient mice. *J Neurosci* 16:2479–2487, 1996
14. Drummond JC, Oh YS, Cole DJ, et al: Phenylephrine-induced hypertension reduces ischemia following middle cerebral artery occlusion in rats. *Stroke* 20:1538–1544, 1989
15. Dzoljic MR, de Vries R, van Leeuwen R: Sleep and nitric oxide: effects of 7-nitro indazole, inhibitor of brain nitric oxide synthase. *Brain Res* 718:145–150, 1996
16. Feelisch M: The biochemical pathways of nitric oxide formation from nitrovasodilators: appropriate choice of exogenous NO donors and aspects of preparation and handling of aqueous NO solutions. *J Cardiovasc Pharmacol* 17 (Suppl 3): S25–S33, 1991
17. Feelisch M, Ostrowski J, Noack E: On the mechanism of NO release from sydnonimines. *J Cardiovasc Pharmacol* 14 (Suppl 11):S13–S22, 1989
18. Foucher-Lavergne A, Kolsky H, Spreux-Varoquaux O, et al: Hemodynamics, tolerability, and pharmacokinetics of linsidomine (SIN-1) infusion during the acute phase of uncomplicated myocardial infarction. *J Cardiovasc Pharmacol* 22: 779–784, 1993
19. Hamada J, Greenberg JH, Croul S, et al: Effects of central inhibition of nitric oxide synthase on focal cerebral ischemia in rats. *J Cereb Blood Flow Metab* 15:779–786, 1995

20. Hara H, Huang PL, Panahian N, et al: Reduced brain edema and infarction volume in mice lacking the neuronal isoform of nitric oxide synthase after transient MCA occlusion. *J Cereb Blood Flow Metab* 16:605–611, 1996
21. Heinzel B, John M, Klatt P, et al: Ca²⁺/calmodulin-dependent formation of hydrogen peroxide by brain nitric oxide synthase. *Biochem J* 281:627–630, 1992
22. Hogg N, Darley-Usmar VM, Wilson MT, et al: Production of hydroxyl radicals from the simultaneous generation of superoxide and nitric oxide. *Biochem J* 281:419–424, 1992
23. Hrabie JA, Klose JR, Wink DA, et al: New nitric oxide-releasing zwitterions derived from polyamines. *J Org Chem* 58:1472–1476, 1993
24. Iadecola C: Bright and dark sides of nitric oxide in ischemic brain injury. *Trends Neurosci* 20:132–139, 1997
25. Iadecola C, Pelligrino DA, Moskowitz MA, et al: Nitric oxide synthase inhibition and cerebrovascular regulation. *J Cereb Blood Flow Metab* 14:175–192, 1994
26. Iadecola C, Xu X, Zhang F, et al: Prolonged inhibition of brain nitric oxide synthase by short-term systemic administration of nitro-L-arginine methyl ester. *Neurochem Res* 19:501–505, 1994
27. Kader A, Frazzini VI, Solomon RA, et al: Nitric oxide production during focal cerebral ischemia in rats. *Stroke* 24: 1709–1716, 1993
28. Kelly PA, Ritchie IM, Arbuthnott GW: Inhibition of neuronal nitric oxide synthase by 7-nitroindazole: effects upon local cerebral blood flow and glucose use in the rat. *J Cereb Blood Flow Metab* 15:766–773, 1995
29. Kelm M, Feelisch M, Spahr R, et al: Quantitative and kinetic characterization of nitric oxide and EDRF released from cultured endothelial cells. *Biochem Biophys Res Commun* 154: 236–244, 1988
30. Kuluz JW, Prado RJ, Dietrich WD, et al: The effect of nitric oxide synthase inhibition on infarct volume after reversible focal cerebral ischemia in conscious rats. *Stroke* 24:2023–2029, 1993
31. Lei SZ, Pan ZH, Aggarwal SK, et al: Effect of nitric oxide production on the redox modulatory site of the NMDA receptor channel complex. *Neuron* 8:1087–1099, 1992
32. MacKenzie GM, Rose S, Bland-Ward PA, et al: Time course of inhibition of brain nitric oxide synthase by 7-nitro indazole. *Neuroreport* 5:1993–1996, 1994
33. Macrae IM, Dawson DA, Norrie JD, et al: Inhibition of nitric oxide synthesis: effects on cerebral blood flow and glucose utilisation in the rat. *J Cereb Blood Flow Metab* 13:985–992, 1993
34. Maiese K, Pek L, Berger SB, et al: Reduction in focal cerebral ischemia by agents acting at imidazole receptors. *J Cereb Blood Flow Metab* 12:53–63, 1992
35. Malinski T, Bailey F, Zhang ZG, et al: Nitric oxide measured by a porphyrinic microsensor in rat brain after transient middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 13: 355–358, 1993
36. Margail I, Parmentier S, Callebort J, et al: Short therapeutic window for MK-801 in transient focal cerebral ischemia in normotensive rats. *J Cereb Blood Flow Metab* 16:107–113, 1996
37. Moncada S, Palmer RMJ, Higgs EA: Nitric oxide: physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 43: 109–142, 1991
38. Moore PK, Babbidge RC, Wallace P, et al: 7-Nitro indazole, an inhibitor of nitric oxide synthase, exhibits anti-nociceptive activity in the mouse without increasing blood pressure. *Br J Pharmacol* 108:296–297, 1993

39. Morikawa E, Moskowitz MA, Huang Z, et al: L-arginine infusion promotes nitric oxide-dependent vasodilation, increases regional cerebral blood flow, and reduces infarction volume in the rat. *Stroke* 25:429–435, 1994
40. Ngai AC, Meno JR, Winn HR: L-NNA suppresses cerebrovascular response and evoked potentials during somatosensory stimulation in rats. *Am J Physiol* 269:H1803–H1810, 1995
41. Noack E, Feelisch M: Molecular aspects underlying the vasodilator action of molsidomine. *J Cardiovasc Pharmacol* 14 (Suppl 11):S1–S5, 1989
42. Pajewski TN, DiFazio CA, Moscicki JC, et al: Nitric oxide synthase inhibitors, 7-nitro indazole and nitroG-L-arginine methyl ester, dose dependently reduce the threshold for isoflurane anesthesia. *Anesthesiology* 85:1111–1119, 1996
43. Rothman SM, Olney JW: Excitotoxicity and the NMDA receptor— still lethal after eight years. *Trends Neurosci* 18:57–58, 1995
44. Rothman SM, Olney JW: Glutamate and the pathophysiology of hypoxic–ischemic brain damage. *Ann Neurol* 19:105–111, 1986
45. Samdani AF, Dawson TM, Dawson VL: Nitric oxide synthase in models of focal ischemia. *Stroke* 28:1283–1288, 1997
46. Shukla A, Dikshit M, Srimal RC: Nitric oxide-dependent bloodbrain barrier permeability alteration in the rat brain. *Experientia* 52:136–140, 1996
47. Tamura A, Graham DI, McCulloch J, et al: Focal cerebral ischaemia in the rat: 1. Description of technique and early neuropathological consequences following middle cerebral artery occlusion. *J Cereb Blood Flow Metab* 1:53–60, 1981
48. Traystman RJ, Moore LE, Helfaer MA, et al: Nitro-L-arginine analogues. Dose- and time-related nitric oxide synthase inhibition in brain. *Stroke* 26:864–869, 1995
49. Villarete LH, Remick DG: Nitric oxide regulation of IL-8 expression in human endothelial cells. *Biochem Biophys Res Commun* 211:671–676, 1995
50. Yoshida T, Limmroth V, Irikura K, et al: The NOS inhibitor, 7-nitroindazole, decreases focal infarct volume but not the response to topical acetylcholine in pial vessels. *J Cereb Blood Flow Metab* 14:924–929, 1994
51. Zagvazdin Y, Sancesario G, Wang YX, et al: Evidence from its cardiovascular effects that 7-nitroindazole may inhibit endothelial nitric oxide synthase in vivo. *Eur J Pharmacol* 303:61–69, 1996
52. Zhang F, Iadecola C: Nitroprusside improves blood flow and reduces brain damage after focal ischemia. *Neuroreport* 4: 559–562, 1993
53. Zhang F, White JG, Iadecola C: Nitric oxide donors increase blood flow and reduce brain damage in focal ischemia: evidence that nitric oxide is beneficial in the early stages of cerebral ischemia. *J Cereb Blood Flow Metab* 14:217–226, 1994

