

Interplay between Folding and Assembly of Fibril-Forming Polypeptides

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Polypeptides can self-assemble into hierarchically organized fibrils consisting of a stack of individually folded polypeptides driven together by hydrophobic interaction. Using a coarse-grained model, we systematically studied this self-assembly as a function of temperature and hydrophobicity of the residues on the outside of the building block. We find the self-assembly can occur via two different pathways—a random aggregation-folding route and a templated-folding process—thus indicating a strong coupling between folding and assembly. The simulation results can explain experimental evidence that assembly through stacking of folded building blocks is rarely observed, at the experimental concentrations. The model thus provides a generic picture of hierarchical fibril formation.

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While, perhaps surprisingly, most proteins and even short peptides share the general ability to form fibrils under appropriate conditions [1–3], the interest in filamentous proteins originates to a large extent from their association with neurodegenerative disorders such as Alzheimer's and Parkinson's disease [4]. However, protein fibers also have promising applications in biomaterials [5,6]. For instance, silk-collagen-like triblock copolymers self-assemble into micrometer long fibrils, which form dilute gels with surprisingly high modulus, serving as promising candidates for novel materials such as artificial tissues [7]. Similar to amyloid fibrils, these silk-collagen fibrils consist of stacked cross- β structures made out of the silklike block, sterically stabilized by the collagenlike hydrophilic flanks. Contrary to amyloid fibrils, the silklike building blocks are so-called β rolls, consisting of two interconnected parallel β sheets with a hydrophobic outside surface, which promotes stacking into long fibers [8]. The mechanism of such hierarchical protein fibril formation is still poorly understood. Yet, such understanding is crucial for controlling fibril nucleation and growth. One clue comes from the fact that experimental fibril elongation speeds are more than 4 orders of magnitude smaller than theoretically predicted by diffusion [9]. This suggests that the fibril growth (and nucleation) encounters pronounced free energy barriers, possibly due to significant conformational changes, e.g., folding, of each polypeptide. Hierarchical fibril formation thus involves a combination of both folding and assembly processes. One can therefore imagine (at least) three possible scenarios: (i) an aggregation-folding process, in which from disordered aggregates of unfolded polypeptides a fibril structure nucleates and grows, (ii) a templated-folding process, in which existing fibrils sequester unfolded polypeptides and induce them to fold, or (iii) a folding-docking process, in which polypeptides first fold

individually before stacking together into fibrils. Computer simulation can complement experiments and yield mechanistic insight, but due to computational challenges, the folding and assembly processes of fibril-forming polypeptides are usually studied separately [8,10–16]. Thus, the interplay between folding and assembly in the self-assembly of polypeptides is still an open question. Here, we investigate such interplay for a system closely related to the silk-collagen-like triblock copolymer, which is, due to its hierarchical structure, an excellent model system.

While in principle atomistic models can provide molecular insight in the folding and assembly process, such all-atom simulations are prohibitively expensive. Instead we employ a coarse-grained polypeptide model in which each residue occupies a single site on a cubic lattice [17], and all other sites are considered as solvent. For each residue a unit vector indicates the direction of the side chain. This highly efficient model captures two essential features for a correct description of folding: the formation of hydrogen bonds and the directionality of side chains. The total potential energy of the system is given by $E = E_{aa} + E_{solvent} + E_{hb} + E_{steric}$, where E_{aa} and $E_{solvent}$ are a knowledge based interaction potential between amino acids, and between amino acids and solvent, respectively (see Table I). This potential has been shown to prevent artificial aggregation of proteins in their native state. E_{hb} is the potential energy of formed hydrogen bonds, and E_{steric} represents the steric hindrance between consecutive residues in a polypeptide chain [18]. Two amino acids in contact interact only when their side chains are either in parallel or pointing towards each other. Similarly, interaction between a residue and solvent only exists when the side chain points to a solvent site. When a residue is not part of a turn in the backbone, it can adopt a strand state, depending on the side chains. Two residues in contact and

both in strand state can form a hydrogen bond with an energy $\epsilon_{\text{hb}} = -0.5$ (all interaction potentials in this work are in reduced units), when their side chains are aligned. An energy penalty $\epsilon_s = 0.55$ prevents side chains of consecutive residues from pointing in the same direction, mimicking steric hindrance and restrictions in bond rotation. Due to its small size alanine becomes considerably more hydrophobic in a β -strand environment [19]. This effect is not captured in the original parametrization of the potential. To compensate for this, and to investigate the influence of the hydrophobicity of the building block's surface, the interaction between alanine (A) and water (w) is varied between 0.01 and 0.6 (see Table I). The model is simulated by using the lattice Monte Carlo (MC) method with a classic move set [17].

Lattice models in general are not expected to fold natural sequences. We therefore first need to design a sequence to fold into the desired β -roll structure. Proper folding of a β roll on a cubic lattice demands a palindromic sequence and a slightly different topology compared to its off-lattice counterpart: whereas the experimental β roll consists of two interconnected parallel β sheets, the lattice β roll is comprised of antiparallel sheets. Restricting the silk part to 80 residues to make the calculation tractable, the design procedure results in the optimal sequence $(E(AI)_3RE(IA)_3R)_5$. This sequence has a repetitive nature comparable to the experimental silk block $(E(AG)_3)_{24}$ [9], but with isoleucine replacing glycine. This is not unrealistic, as glycines in β sheets are more hydrophobic than the average glycine [19]. The extra (arginine) residue has been introduced to fit the β -roll structure on the lattice. In experiments, random aggregation of fibrils is prevented by the presence of hydrophilic flanks [9]. A similar effect has been shown by lattice model simulations [20]. To mimic those flanks we attached ten glutamate residues to both peptide termini. As shown in Fig. 1, the designed polypeptide indeed folds into a β -roll structure with a hydrophobic outside surface covered with alanine side chains.

Whereas polypeptide folding is driven by several types of interactions, including hydrophobicity and hydrogen

TABLE I. Interaction matrix (in reduced units) for the residues in the used sequence [17]. Amino acids are denoted by their one-letter code (I = isoleucine, A = alanine, R = arginine, E = glutamate) and w denotes the solvent (water). The hydrophobicity of alanine is varied via the A - w interaction ($\epsilon_{A,w}$) as indicated.

	I	A	R	E	w
I	-0.79	-0.40	0.5	0.69	0.7
A		-0.34	0.49	0.77	0.01; 0.1; 0.2; 0.4; 0.6
R			0.43	-0.6	-0.57
E				1.02	-0.78
w					0.0

bond formation, the assembly is purely hydrophobicity driven [19]. To investigate the interplay between folding and assembly, it is necessary to be able to influence both independently. Control parameters in the folding and assembly process are the temperature and the concentration, but these will probably influence both aspects simultaneously. Therefore we also change the hydrophobic strength of the outside surface, by varying the alanine-solvent interaction.

We performed replica exchange Monte Carlo (REMC) simulations of the optimized sequence on a 200^3 lattice with periodic boundary conditions. Each REMC simulation consisted of 64 replicas with a (reduced) temperature distribution around the transition temperature ($0.35 \leq T^* \leq 0.45$), optimized by a feedback-optimization algorithm [21]. A replica exchange was attempted each 1000 MC cycles. After equilibrating for 1×10^{10} MC cycles per replica, we performed 5×10^{10} MC cycles for production. Employing the virtual-move parallel tempering method [22] ensured optimal use of the simulation data.

For a single polypeptide chain, corresponding to infinite dilution, we obtained the folding temperature from the heat capacity peak. Figure 1 shows that this temperature is weakly dependent on the alanine hydrophobicity; at high $\epsilon_{A,w}$ the folded chain is slightly more stable, possibly because alanine side chains are more shielded from the water. In addition, with increasing hydrophobicity we observe more collapsed structures in the unfolded state.

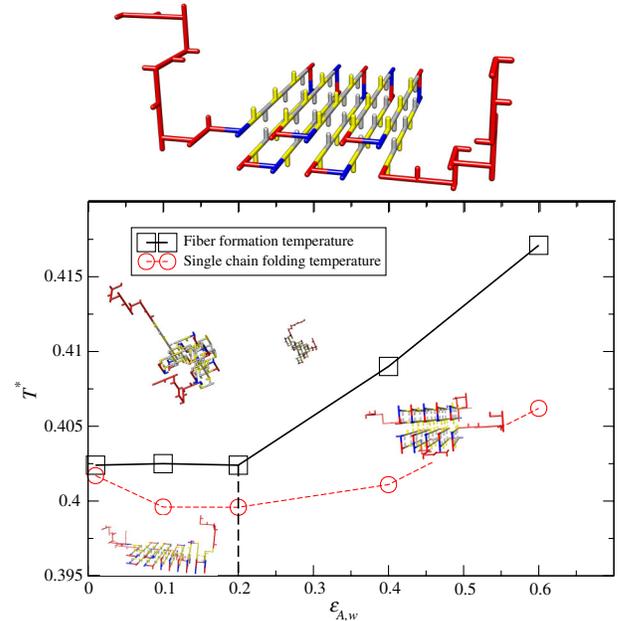


FIG. 1 (color online). Top: folded β -roll structure of a single polypeptide [alanine (A) = yellow, isoleucine (I) = white, glutamate (E) = red, arginine (R) = blue]. Bottom: equilibrium phase diagram in the T^* - $\epsilon_{A,w}$ plane, with transition temperatures for two polypeptides (squares) and the folding temperature of single polypeptide (circles). Typical snapshots of the isolated unfolded, aligned, and stacked phases are included.

Next, we performed REMC simulations of two polypeptides. Assuming a lattice space of 0.38 nm [17], this corresponds to a concentration of 7.57 μM . At high temperature the polypeptides are unfolded and mostly separated (see the top left of Fig. 1). Upon decreasing temperature, a sharp peak in the heat capacity signals a transition to a self-assembled folded state, with a structure that depends on the alanine hydrophobicity $\epsilon_{A,w}$. For $\epsilon_{A,w} < 0.2$, the two folded β rolls align side by side. As in this regime the transition temperature remains almost constant, the formation of the aligned state is not driven by the hydrophobicity of the β -roll surface, but instead by hydrogen bonds between the sides of two β rolls. For $\epsilon_{A,w} \geq 0.2$, the two folded β rolls are stacked on top of each other. In this regime the transition temperature increases monotonically with $\epsilon_{A,w}$ as the stacked state is stabilized by the hydrophobicity of the outside surface of the folded β roll.

To shed light on the self-assembly mechanism of fibril-forming polypeptides, we plot in the left-hand side of Fig. 2, for two temperatures, the free energy landscapes associated with the stacked state formation at $\epsilon_{A,w} = 0.6$ as a function of the total number of hydrogen bonds H_{tot} , and

the silk-block center-to-center distance R_{cc} . For both temperatures the global free energy minimum at ($R_{\text{cc}} = 2$, $H_{\text{tot}} \approx 45$) corresponds to the stacked state, and a local minimum at ($R_{\text{cc}} \approx 5$, $H_{\text{tot}} \approx 35$) represents the metastable aligned state. However, the mechanistic folding pathways differ significantly for the two temperatures. At $T^* = 0.4109$, above the single polypeptide folding temperature, there exists [besides the minimum at ($R_{\text{cc}} \approx 100$, $H_{\text{tot}} \approx 0$) corresponding to isolated random coils] a second local minimum at ($R_{\text{cc}} \approx 2$, $H_{\text{tot}} \approx 0$), associated with a disordered aggregate of two polypeptides. The presence of this aggregated state causes the folding mechanism to pass through a saddle point around ($R_{\text{cc}} \approx 8$, $H_{\text{tot}} \approx 1$). This implies that above the single polypeptide folding temperature the self-assembly follows the pathway unfolded state \rightarrow random aggregation \rightarrow stacked state.

At $T^* = 0.3907$, below the single polypeptide folding temperature, the free energy landscape changes significantly and features a local minimum around ($R_{\text{cc}} \approx 100$, $H_{\text{tot}} \approx 30$), which corresponds to an intermediate state consisting of a separated folded and unfolded polypeptide. The channel connecting this state to the stacked state

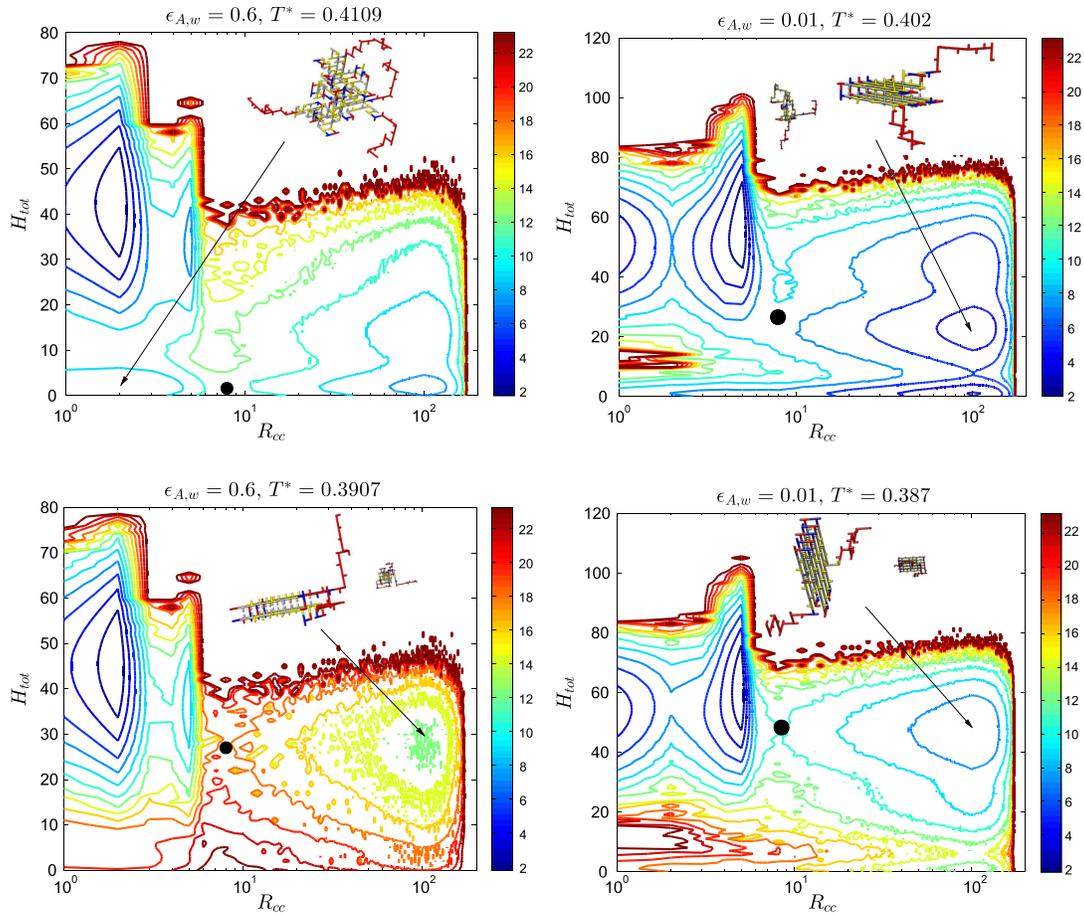


FIG. 2 (color online). Free energy landscape as a function of the silk-block center-to-center distance R_{cc} , and the total number of hydrogen bonds H_{tot} . Left column: $\epsilon_{A,w} = 0.6$, $T^* = 0.4109$ (top) and 0.3907 (bottom). Right column: $\epsilon_{A,w} = 0.01$ at the temperature $T^* = 0.402$ (top) and 0.387 (bottom). Solid circles denote saddle points.

travels through a saddle point around ($R_{cc} \approx 10$, $H_{tot} \approx 30$), suggesting that the assembly is a self-templated process [23], in which a polypeptide folds on top of an already folded polypeptide. At very low temperatures the two polypeptides may fold independently before stacking together. However, we failed to observe such a folding-docking process down to $T^* = 0.35$, more than 15% below the transition temperature. Apparently, this is a very unlikely process, which explains why in experiments individual β rolls are rarely observed, whereas they readily form on the growing end of an existing filament [9].

For an alanine hydrophobicity of $\epsilon_{A,w} = 0.01$, typical free energy landscapes in the right-hand side of Fig. 2 show that below the transition temperature the global minimum is located at ($R_{cc} \approx 5$, $H_{tot} \approx 60$), corresponding to the aligned state (see Fig. 1). For $T^* = 0.402$, just below the transition temperature, the assembly already occurs via the self-templated process, as indicated by the saddle point at ($R_{cc} \approx 8$, $H_{tot} \approx 25$) and the metastable intermediate state located at ($R_{cc} \approx 100$, $H_{tot} \approx 25$). The transition temperature is very close to the single polypeptide folding temperature (see Fig. 1), revealing that the binding between the folded polypeptides is weak. At $T^* = 0.387$ one finds individual folded polypeptides in the solution, as indicated by a local minimum at ($R_{cc} \approx 100$, $H_{tot} \approx 50$). The saddle point at ($R_{cc} \approx 10$, $H_{tot} \approx 50$) suggests that the assembly then occurs via the folding-docking process. The local minimum at ($R_{cc} \approx 1$, $H_{tot} \approx 50$) corresponds to a metastable structure consisting of two interlocked folded β sheets, which is probably an artifact of the model. In contrast to the strong hydrophobicity case, there is no third pathway via a random aggregate. We note that the high hydrophobicity case is probably a more realistic approximation to the experiments of Ref. [9].

To investigate the effect of polypeptide concentration, we performed REMC simulations at concentrations ranging from 1 to 60 μM , by changing the lattice size. Figure 3 shows the heat capacity for $\epsilon_{A,w} = 0.6$ as a function of temperature. The decreasing height of the heat capacity peak with decreasing concentration indicates the self-assembly process becomes less cooperative at lower concentration. Still, the self-assembly transition temperature is always above the single polypeptide folding temperature, and the mechanism does not change. The inset in Fig. 3 shows the linear relationship between the reciprocal transition temperature and the logarithm of polypeptide concentration. We explain this behavior as follows. If assembly of two peptides yields an energy gain $\Delta E < 0$, then at the transition temperature T_c the free energy difference between the isolated and assembled state is $\Delta F = \Delta E - T_c \Delta S = 0$, where ΔS is the entropy lost in the assembly. Assuming ideal gas behavior of isolated peptides $\Delta S = k_B \ln C_p - \Delta S_c$, where C_p is the polypeptide concentration. The first term is the translational entropy lost, while the second term is the conformational entropy

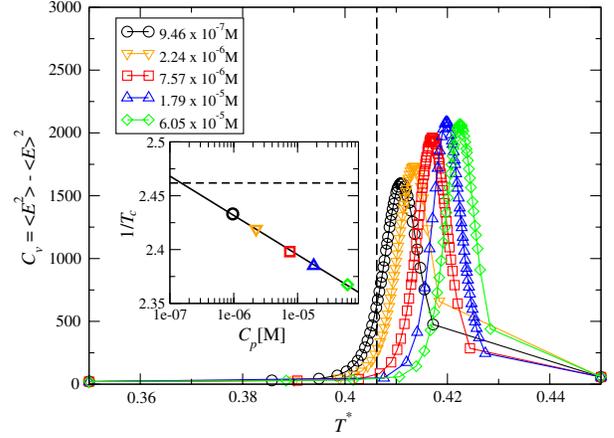


FIG. 3 (color online). Main: heat capacity versus temperature for two polypeptides at various concentrations with $\epsilon_{A,w} = 0.6$. Inset: the reciprocal transition temperature T_c versus the logarithm of peptide concentration. In both plots the dashed line indicates the single polypeptide folding temperature.

lost ($\Delta S_c > 0$). It follows that $1/T_c = (k_B/\Delta E) \times \ln C_p - \Delta S_c/\Delta E = a_1 \ln C_p + a_2$, with $a_1 < 0$ and $a_2 > 0$, as we find in the simulations. We note that at very low concentration ($C_p < 0.1 \mu\text{M}$) the assembly will not influence the folding temperature anymore, as indicated by the dashed line in the inset. We speculate that below this concentration a folding-docking mechanism may indeed be present. For very high concentration $C_p \sim 1/8R_g^3$, with R_g the radius of gyration, the unfolded chains are touching, and T_c will also become independent of concentration.

In conclusion, we used a coarse-grained lattice model to systematically study the interplay between folding and assembly in the formation of hierarchically organized polypeptide fibrils. Despite the simplicity of the model, the results not only show a remarkably rich phase behavior, but also, and most importantly, reveal a generic mechanism for protein fibril formation, depending on the relative strength of the intermolecular driving force for assembly (the hydrophobicity of the folded protein surface) with respect to the strength of the intramolecular driving force for protein folding (the hydrogen bonding). Coupling between these driving forces leads to an effect similar to allostery where a conformation change goes along with binding, and produces a templated-folding growth process at temperatures below the melting point. The model explains why experiments rarely detect the presence of individual β rolls, but instead provide evidence for a templating mechanism for nucleation and growth [9].

Our results show that the formation and growth of fibril structures can be tuned by varying the temperature and/or the hydrophobicity of specific residues, and are thus not only of fundamental interest, but may also provide guidance to fabricating novel biofibrils with high quality in experiments.

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- [1] C. M. Dobson, *Trends Biochem. Sci.* **24**, 329 (1999).
- [2] F. Chiti and C. M. Dobson, *Annu. Rev. Biochem.* **75**, 333 (2006).
- [3] C. Dobson, *Nature (London)* **426**, 884 (2003).
- [4] C. Haass and D. J. Selkoe, *Nat. Rev. Mol. Cell Biol.* **8**, 101 (2007).
- [5] T. Scheibel, R. Parthasarathy, G. Sawicki, X. M. Lin, H. Jaeger, and S. L. Lindquist, *Proc. Natl. Acad. Sci. U.S.A.* **100**, 4527 (2003).
- [6] K. J. Channon, G. L. Devlin, and C. E. MacPhee, *J. Am. Chem. Soc.* **131**, 12 520 (2009).
- [7] A. A. Martens, G. Portale, M. W. T. Werten, R. J. de Vries, G. Eggingk, M. A. Cohen Stuart, and F. A. de Wolf, *Macromolecules* **42**, 1002 (2009).
- [8] M. Schor, A. A. Martens, F. A. de Wolf, M. A. Cohen-Stuart, and P. G. Bolhuis, *Soft Matter* **5**, 2658 (2009).
- [9] H. B. Lennart, X. J. Beaudoux, J. M. Kleijn, F. A. de Wolf, and M. A. Cohen Stuart, *ACS Nano* **6**, 133 (2012).
- [10] A. Irbäck, S. Æ. Jónsson, N. Linnemann, B. Linse, and S. Wallin, *Phys. Rev. Lett.* **110**, 058101 (2013).
- [11] S. Auer and D. Kashchiev, *Phys. Rev. Lett.* **104**, 168105 (2010).
- [12] J. E. Straub and D. Thirumalai, *Annu. Rev. Phys. Chem.* **62**, 437 (2011).
- [13] M. Schor and P. G. Bolhuis, *Phys. Chem. Chem. Phys.* **13**, 10 457 (2011).
- [14] H. D. Nguyen and C. K. Hall, *J. Am. Chem. Soc.* **128**, 1890 (2006).
- [15] V. A. Wagoner, M. Cheon, I. Chang, and C. K. Hall, *J. Mol. Biol.* **416**, 598 (2012).
- [16] E. M. Phelps and C. K. Hall, *Proteins: Struct., Funct., Bioinf.* **80**, 1582 (2012).
- [17] S. Abeln and D. Frenkel, *Biophys. J.* **100**, 693 (2011).
- [18] S. Abeln and D. Frenkel (to be published).
- [19] See Supplemental Material at <http://link.aps.org/supplemental/10.1103/PhysRevLett.111.058101> for the description of hydrophobicity of glycine and alanine in beta strands, and the energetic contribution to the polypeptide folding.
- [20] S. Abeln and D. Frenkel, *PLoS Comput. Biol.* **4**, e1000241 (2008).
- [21] H. G. Katzgraber, S. Trebst, D. A. Huse, and M. Troyer, *J. Stat. Mech.: Theory Exp.* 2006, P03018 (2006).
- [22] I. Coluzza and D. Frenkel, *ChemPhysChem* **6**, 1779 (2005).
- [23] S. Auer, C. M. Dobson, M. Vendruscolo, and A. Maritan, *Phys. Rev. Lett.* **101**, 258101 (2008).