



**UvA-DARE (Digital Academic Repository)**

**Oxic-anoxic regime shifts mediated by feedbacks between biogeochemical processes and microbial community dynamics**

Bush, T. ; Diao, M.; Allen, R.J.; Sinnige, R.; Muijzer, G.; Huisman, J.

*Published in:*  
Nature Communications

*DOI:*  
[10.1038/s41467-017-00912-x](https://doi.org/10.1038/s41467-017-00912-x)

[Link to publication](#)

*Citation for published version (APA):*

Bush, T., Diao, M., Allen, R. J., Sinnige, R., Muijzer, G., & Huisman, J. (2017). Oxic-anoxic regime shifts mediated by feedbacks between biogeochemical processes and microbial community dynamics. *Nature Communications*, 8, [789]. <https://doi.org/10.1038/s41467-017-00912-x>

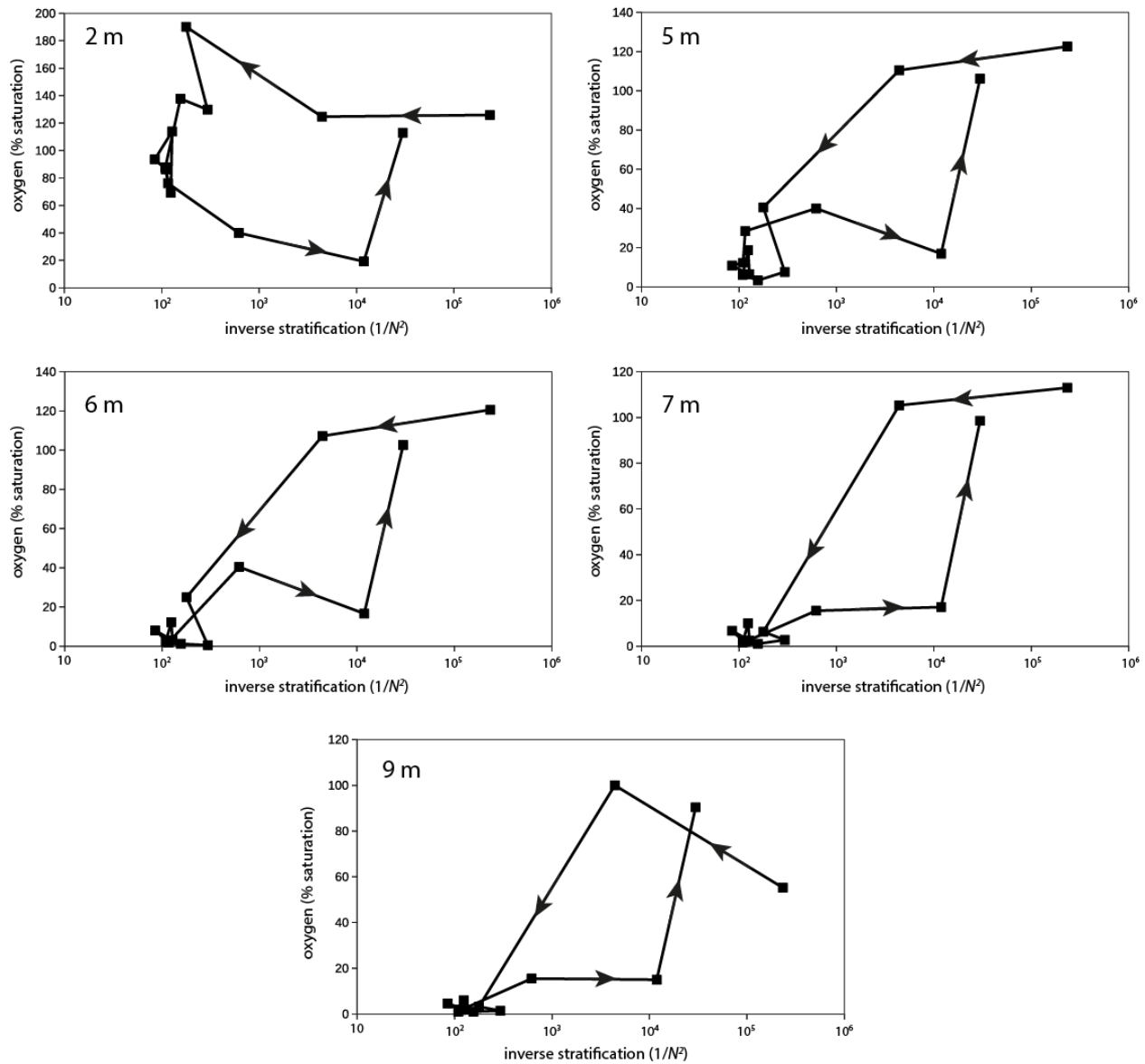
**General rights**

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

**Disclaimer/Complaints regulations**

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.

**Supplementary Figure 1. Hysteresis loops for different depths in Lake Vechten.** The graphs plot the oxygen saturation level against the inverse of the stratification strength ( $1/N^2$ , where  $N^2$  is the squared buoyancy frequency). The inverse of the stratification strength provides a simple proxy of oxygen diffusivity across the thermocline (see Methods). A hysteresis loop is found irrespective of whether oxygen saturation is measured at a depth of **a**, 2 m; **b**, 5 m; **c**, 6 m; **d**, 7 m; **e**, 9 m. Data points are from March 2013 to March 2014; arrows indicate the direction of time.



**Supplementary Table 1.** Parameter values of the model.

Parameter	Meaning	Value	Reference
$g_{\max, \text{CB}}$	Maximum specific growth rate of CB	0.05 hr <sup>-1</sup>	1
$g_{\max, \text{PB}}$	Maximum specific growth rate of PB	0.07 hr <sup>-1</sup>	2
$g_{\max, \text{SB}}$	Maximum specific growth rate of SB	0.1 hr <sup>-1</sup>	3,4
$K_{\text{PB,SR}}$	Half-saturation constant of PB on reduced sulfur	10 μM	5
$K_{\text{SB,SO}}$	Half-saturation constant of SB on oxidized sulfur	5 μM	6
$K_{\text{CB,P}}$	Half-saturation constant of CB on phosphorus	0.2 μM	7
$K_{\text{PB,P}}$	Half-saturation constant of PB on phosphorus	0.5 μM	8
$K_{\text{SB,P}}$	Half-saturation constant of SB on phosphorus	0.5 μM	-
$H_{\text{CB,SR}}$	Half-inhibition constant of CB on reduced sulfur	300 μM	9
$H_{\text{PB,O}}$	Half-inhibition constant of PB on oxygen	100 μM	10
$H_{\text{SB,O}}$	Half-inhibition constant of SB on oxygen	100 μM	10
$y_{\text{SB}}^{\text{SO}}$	Yield of SB on oxidized sulfur	3.33×10 <sup>7</sup> cells μM <sup>-1</sup>	11
$y_{\text{PB}}^{\text{SR}}$	Yield of PB on reduced sulfur	1.25×10 <sup>7</sup> cells μM <sup>-1</sup>	9
$y_{\text{CB}}^{\text{P}}$	Yield of CB on phosphorus	1.67×10 <sup>8</sup> cells μM <sup>-1</sup>	12
$y_{\text{PB}}^{\text{P}}$	Yield of PB on phosphorus	1.67×10 <sup>8</sup> cells μM <sup>-1</sup>	-
$y_{\text{SB}}^{\text{P}}$	Yield of SB on phosphorus	1.67×10 <sup>8</sup> cells μM <sup>-1</sup>	-
$p_{\text{CB}}$	Production of oxygen per cyanobacterial cell	6×10 <sup>-9</sup> μM cell <sup>-1</sup>	13
$m_{\text{CB}}$	Mortality rate of CB	0.020 hr <sup>-1</sup>	-
$m_{\text{PB}}$	Mortality rate of PB	0.028 hr <sup>-1</sup>	-
$m_{\text{SB}}$	Mortality rate of SB	0.040 hr <sup>-1</sup>	-
$\alpha_{\text{S}}$	Diffusivity of sulfur	0.001 hr <sup>-1</sup>	-
$\alpha_{\text{O}}$	Diffusivity of oxygen	10 <sup>-6</sup> – 10 <sup>-2</sup> hr <sup>-1</sup>	-
$\alpha_{\text{P}}$	Diffusivity of phosphorus*	0.01 hr <sup>-1</sup>	-
$S_{\text{R,b}}$	Background concentration of reduced sulfur	300 μM	14
$S_{\text{O,b}}$	Background concentration of oxidized sulfur	300 μM	14
$O_{\text{b}}$	Background concentration of oxygen	300 μM	15
$P_{\text{b}}$	Background concentration of phosphorus	2 – 10 μM	16
$c$	Oxidation rate of reduced sulfur	4×10 <sup>-5</sup> μM <sup>-1</sup> hr <sup>-1</sup>	17,18

CB = cyanobacteria; PB = phototrophic sulfur bacteria; SB = sulfate-reducing bacteria

\*We assumed a higher diffusive influx for phosphorus than for sulfur and oxygen, because the phosphorus influx also includes phosphorus release from the sediment and remineralization from dead biomass.

### Supplementary References

1. Paerl, H. W. & Huisman, J. Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. *Env. Microb. Rep.* **1**, 27–37 (2009).
2. Montesinos, E. Change in size of *Chromatium minus* cells in relation to growth rate, sulfur content, and photosynthetic activity: a comparison of pure cultures and field populations. *Appl. Environ. Microbiol.* **53**, 864–871 (1987).

3. Kalyuzhnyi, S., Fedorovich, V., Lens, P., Hulshoff Pol, L. & Lettinga, G. Mathematical modelling as a tool to study population dynamics between sulfate reducing and methanogenic bacteria. *Biodegradation* **9**, 187–199 (1998).
4. Dev, S., Roy, S. & Bhattacharya, J. Understanding the performance of sulfate reducing bacteria based packed bed reactor by growth kinetics study and microbial profiling. *J. Environ. Manag.* **177**, 101–110 (2016).
5. Van Gemerden, H. The sulfide affinity of phototrophic bacteria in relation to the location of elemental sulfur. *Arch. Microbiol.* **139**, 289–294 (1984).
6. Ingvorsen, K., Zehnder, A. J. B. & Jørgensen, B. B. Kinetics of sulfate and acetate uptake by *Desulfobacter postgatei*. *Appl. Environ. Microbiol.* **47**, 403–408 (1984).
7. Kromkamp, J., Van den Heuvel, A. & Mur, L. R. Phosphorus uptake and photosynthesis by phosphate-limited cultures of the cyanobacterium *Microcystis aeruginosa*. *Br. Phycol. J.* **24**, 347–355 (1989).
8. Bañeras, L., Ros-Ponsatí, M., Cristina, X. P., Garcia-Gil, J. L. & Borrego, C. M. Phosphorus deficiency and kinetics of alkaline phosphatase in isolates and natural populations of phototrophic sulphur bacteria. *FEMS Microbiol. Ecol.* **73**, 243–253 (2010).
9. De Wit, R., Van den Ende, F. P. & Van Gemerden, H. Mathematical simulation of the interactions among cyanobacteria, purple sulfur bacteria and chemotrophic sulfur bacteria in microbial mat communities. *FEMS Microbiol. Ecol.* **17**, 117–136 (1995).
10. Gerritse, J., Schut, F. & Gottschal, J. C. Modelling of mixed chemostat cultures of an aerobic bacterium, *Comamonas testosteroni*, and an anaerobic bacterium, *Veillonella alcalescens*: comparison with experimental data. *Appl. Environ. Microbiol.* **58**, 1466–1476 (1992).
11. Jin, Q. & Bethke, C. M. The thermodynamics and kinetics of microbial metabolism. *Am. J. Sci.* **307**, 643–677 (2007).
12. Saxton, M. A., Arnold, R. J., Bourbonniere, R. A., McKay, R. M. L. & Wilhelm, S. W. Plasticity of total and intracellular phosphorus quotas in *Microcystis aeruginosa* cultures and Lake Erie algal assemblages. *Front. Microbiol.* **3**, 3 (2012).
13. Gons, H. J. & Rijkeboer, M. The ‘true’ growth efficiency of phytoplankton as influenced by light attenuation and insolation: implications of the photosynthesis-irradiance relationship. *Hydrobiol.* **238**, 169–176 (1992).
14. Goldhaber, M. B. Sulfur-rich sediments. *Treatise on Geochemistry* **7**, 257–288 (2003).
15. Shaffer, G., Olsen, S. M. & Pedersen, J. O. P. Long-term ocean oxygen depletion in response to carbon dioxide emissions from fossil fuels. *Nat. Geosci.* **2**, 105–109 (2009).
16. Ruttenberg, K. C. The global phosphorus cycle. *Treatise on Geochemistry* **8**, 585–643 (2003).
17. Luther, G. W. *et al.* Thermodynamics and kinetics of sulfide oxidation by oxygen: a look at inorganically controlled reactions and biologically mediated processes in the environment. *Front. Microbiol.* **2**, 62 (2011).
18. Millero, F. J., Hubinger, S., Fernandez, M. & Garnett, S. Oxidation of H<sub>2</sub>S in seawater as a function of temperature, pH, and ionic strength. *Environ. Sci. Technol.* **21**, 439–443 (1987).