



UvA-DARE (Digital Academic Repository)

Standardization procedure for plasma biomarker analysis in rat models of epileptogenesis: Focus on circulating microRNAs

van Vliet, E.A.; Puhakka, N.; Mills, J.D.; Srivastava, P.K.; Johnson, M.R.; Roncon, P.; Das Gupta, S.; Karttunen, J.; Simonato, M.; Lukasiuk, K.; Gorter, J.A.; Aronica, E.; Pitkänen, A.

DOI

[10.1111/epi.13915](https://doi.org/10.1111/epi.13915)

Publication date

2017

Document Version

Other version

Published in

Epilepsia

License

Article 25fa Dutch Copyright Act (<https://www.openaccess.nl/en/policies/open-access-in-dutch-copyright-law-taverne-amendment>)

[Link to publication](#)

Citation for published version (APA):

van Vliet, E. A., Puhakka, N., Mills, J. D., Srivastava, P. K., Johnson, M. R., Roncon, P., Das Gupta, S., Karttunen, J., Simonato, M., Lukasiuk, K., Gorter, J. A., Aronica, E., & Pitkänen, A. (2017). Standardization procedure for plasma biomarker analysis in rat models of epileptogenesis: Focus on circulating microRNAs. *Epilepsia*, *58*(12), 2013–2024. <https://doi.org/10.1111/epi.13915>

General rights

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, P.O. Box 19185, 1000 GD Amsterdam, The Netherlands. You will be contacted as soon as possible.

UvA-DARE is a service provided by the library of the University of Amsterdam (<https://dare.uva.nl>)

Supplementary table 1. Summary of blood sampling protocols in studies using rat plasma for miRNA detection

Disease or phenomena	Sampling site	Blood collection	Plasma separation/storage	Plasma QC	Plasma volume/RNA extraction	RNA QC /spike-ins	miRNA platform	Validation	Normalization / method	Calibrator/ other controls	Ref
Acute kidney injury	Abdominal aorta	EDTA			miRcute, mirVana without phenol	Bioanalyzer 2100	GoScript with stemloop primers		miR-16/ $2^{-\Delta\Delta CT}$		1
Non-identical miRNA profiles between arterial and venous plasma	abdominal aorta or vena cava	EDTA + RNAase inhibitor	1600 g, 10 min; 16,000 g, 4°C, filtering 0.22 μ m filter		miRNeasy		Agilent Rat miRNA V21.0 Microarray	RT-qPCR	Cel-miR-39-3p/ $2^{-\Delta\Delta CT}$		2
Diabetic nephropathy	Tail vein				TRIzol	Synthetic oligonucleotide	Hairpin-it miRNA RT-qPCR		U6-snRNA/ $2^{-\Delta\Delta CT}$	miR-130b	3
Chronic sucrose ingestion associated inflammation		K+EDTA	3000 rpm, 15 min, 4°C/-70°C		100 μ l/ miRNeasy, exoRNeasy	Cel-miR-39, bacterial ribosomal RNA	TaqMan RT-qPCR		Cel-miR-39/ $2^{-\Delta\Delta CT}$		4
Type 2 diabetes	Tail vein				150 μ l/ miRNeasy	Bioconductor package HTqSeq v1.16.0	Taqman miRNA Array, Card A V.2.0		Global mean normalization		5
Antigen-induced pulmonary inflammation	Abdominal aorta	Citrate	1800 g, 20 min, 4°C/-80°C	Coagulated samples excluded, hemolysis avoided	200 μ l/ Tri reagent BD	NanoDrop 2000c /Cel-miR-39 spike-in	iQ5 system		Cel-miR-39/ $2^{-\Delta\Delta CT}$	Non-spike-in control, non-template control, non-RT control, custom control pool	6
Heparinase treatment of	Aorta	EDTA, < 30	1600 g, 4°C,		200 μ l/ TRIzol	Bioanalyzer 2100 /E. coli	TaqMan RT-		Absolute quantification		7

heparin-contaminated plasma		min	2x/-80°C		LS	tRNA, cel-miR-39-3p	qPCR		using calibration curves from synthetic miRNAs		
CNS impairments associated with Gulf War Illness	Cardiac puncture or right atrium	EDTA	1900 g, 10 min, 4°C; 16,000 g, 10 min / -80° C; 16,000 g, 5 min, 4°C		200 µl/ miRNeasy, ExoQuick	Bioanalyzer 2100/ Cel-miR-39, synthetic control miRNA	miScript RT-qPCR	Illumina HiSeq2000	Cel-miR-39/ ΔΔCT		8
Acute rejection post limb transplantation	Caudal vein				100 µl/ mirVana PARIS		TaqMan RT-qPCR		Synthetic ath-mir-159a/ ΔΔCT		9
Acute skeletal muscle damage	Exsanguination	K ₂ -EDTA	2000 g, 10 min, 4°C, 2x		100 µl/ mirVana PARIS	miR-103a-3p	Mouse&Rat panel IplI, V3.M (Exiqon), Customized qPCR plates, Pick-&Mix microRNA	miScript RT-qPCR	Geometric mean of several miRNAs, based on geNorm algorithm	Plasma pools enriched with RBC and platelet pellets to study effects of RBC and platelet enrichment.	10
Contrast-induced acute kidney injury		EDTA, < 1 hour, RT	820 g, 10 min, 4°C; 16,000 g, 10 min, 4°C / -80°C		mirVana	NanoDrop 1000 / cel-miR-39	Agilent microRNAs microarray v.10.1	TaqMan RT-qPCR	Cel-miR-39/ 2 ^{ΔΔCT}		11
Fibrotic liver disease			3000 rpm, 10 min, 4°C/ -80°C		miRNeasy	Cel-miR-39	TaqMan RT-qPCR		U6-snrRNA/ 2 ^{ΔΔCT}		12

Nephrotoxicity and hepatotoxicity	Lateral tail vein (non-terminal) or vena cava (terminal)	EDTA	4°C		200 µl/ Asuragen	RT-qPCR of 3 miRNAs	miR-seq using TruSeq Small RNA Library Kit		U6-snRNA/ ΔΔCT		13
Acute kidney injury	Abdominal aorta	EDTA, < 1 hour	3000 g, 15 min, 4°C; 12,000 g, 15 min, 4°C		mirVana PARIS	Cel-miR-39	rodent TaqMan low-density array assay	TaqMan RT-qPCR	Cel-miR-39/ 2 ^{ΔΔCT}		14
Comparison of three pulsed focused ultrasound regimes to release tumor-derived miRNA into circulation	Jugular vein via implanted polyurethane catheters for repeated sampling	K ₂ -EDTA gentle inversion of blood samples 20 times without shaking to avoid hemolysis	1300 g, 15 min, RT; 250 µL of plasma aspirated and immediately combined with 1.25 mL of Qiazol, lysed/ -80°C		250 µL/ miRNeasy	Cel-miR-39	Human panel I, V2.M RT-qPCR arrays (Exiqon) using miRCURY LNA Universal RT miRNA PCR kit (Exiqon)	TaqMan RT-qPCR,	Cel-miR-39		15
Diabetic nephropathy	Tail vein				TRIzol		miRScript RT-qPCR	Taqman RT-qPCR using the Hairpin-it miRNA qPCR kit	U6-snRNA / 2 ^{ΔΔCT}		16
Retinal toxicity	Tail vein	EDTA	1500 g, 15 min		100 µL/ miRNeasy	Cel-miR-39	Poly(A) tailing kit, MultiScribe, RT-qPCR		Cel-miR-39, miR-92, miR-181, miR-192		17
Cardiac injury	Abdominal aorta	EDTA tubes pre-cooled on ice, Rapidly centrifuged	2100 g, 10 min, 4°C/ -80°C		200 µL/ miRNeasy	Ath-miR-159a	TaqMan customized multiplexed miRNA assay	ath-miR159a	Calibration curves from pooled synthetic miRNA stocks, plus miR-16-15p and mR-192-5p		18

Perinatal glutamate exposure	Atrial catheters for repeated sampling and trunk blood				miRNeasy	260/280 ratio	miScript II, miRNA qPCR assay using Rat miFinder RT2 miRNA PCR 96-well plate array		6 normalization controls, and controls to assess recovery, RT and PCR performance/ $\Delta\Delta CT$		19
Drug-induced cardiac injury	Posterior vena cava	Li-heparin S-Monovette tubes		miR-451, miR-486	40 μ L/ Firefly Bioworks		high throughput miR-profiling technology from Firefly BioWorks		miR-92a-3p, miR-103-3p, miR-106b-5p		20
Contrast-induced acute kidney injury	Tail cutting				miRNeasy	Cel-miR-39	miScript RT-qPCR		Cel-miR-39/ $2^{-\Delta\Delta CT}$		21
Third-degree burn	Abdominal aorta	Lithium heparin	200 g, 10 min/ -80°C		TRIzol	TRIzol	Revert Aid, SsoFast EvaGreen RT-qPCR		U6-snRNA		22
Temporal lobe epilepsy	Intracardiac withdrawal	EDTA	1000 g, 2min; 2500 g, 2 min/ -80°C		200 μ L/ miRNeasy	Cel-miR-39	Rat miRNA MicroArray Kit, Human micro-RNA Microarray V3, #G4470C, Agilent	TaqMan RT-qPCR	Cel-miR-39/ ΔCT		23
Pulmonary arterial hypertension	Inferior vena cava	EDTA	1700 g, 10 min, 4°C; 11,000 g, 2 min, 4°C/ -80°C; 11,000 g, 5 min, 4°C		100 μ L/ miRNeasy	Cel-miR-39	miScript II RT-qPCR		miR-24-3p/ ΔCT		24

Temporal lobe epileptogenesis	Trunk blood	K ₂ -EDTA, immediately	1500 g, 10 min, 4°C / -80°C		200 µL/ miRNeasy	Bioanalyzer 2100	TaqMan RT-qPCR		miR-23a/ 2 ⁻ ΔΔCT		32
Hypertension-induced heart failure	Catheters implanted in left femoral artery for repeated sampling. Sacrifice blood from vena cava	EDTA	1500 g, 15 min, 4°C / -80°C		100 µL/ TRIzol LS	Cel-miR-2, Cel-lin-4	TaqMan Array miRNA rodent card A	TaqMan RT-qPCR	Cel-miR-2, Cel-lin-4		33
Classification of muscular tissue	Inferior vena cava	EDTA	1600 g, 15 min, 4°C		mirVana PARIS		ABI TaqMan Rodent MicroRNA Array kit (Card A and B)	TaqMan RT-qPCR	Synthetic small RNA		34
Acute phased pancreatic injury	Inferior vena cava	EDTA	1600 g, 15 min, 4°C		mirVana PARIS				Synthetic small RNA		34
Congestive Heart Failure	Inferior vena cava	EDTA	1600 g, 15 min, 4°C		mirVana PARIS		ABI TaqMan Rodent MicroRNA Array kit (Card A)	TaqMan RT-qPCR	Synthetic small RNA		34
Drug-induced ocular toxicity	Exsanguination	EDTA			100 µL/ miRNeasy		Poly(A) tailing kit, MultiScribe, RT-qPCR		miR-192		35
Drug-induced hepatotoxicity	Sublingual veins				200 µL/ miRNeasy	Bioanalyzer 2100/ ath miRNA control	TaqMan RT-qPCR		ath miRNA control/ ΔCT		36
Acetaminophen-induced hepatotoxicity		Gel and clot activator tube	415 g, 5 min, 4°C		TRIzol, mirVana		miScript RT-qPCR		miR-16, miR-92a, let-7a, miR-103, 5S-snRNA, U6-snRNA using		37

									geNorm, BestKeeper, Normfinder/ ΔCT		
Hepatocellular Injury, Cholestasis, and Steatosis		EDTA, 30 min on ice	?/ -80°C		600 μL/ mirVana PARIS	Cel-miR-39	TaqMan Array Rodent MicroRNA A+B Cards Sets v2.0		3 endogenous controls/ ΔCT		38
Hypertension-induced heart failure					TRIzol LS	Cel-miR-2, Cel-lin-4	TaqMan RT-qPCR		Cel-miR-2, Cel-lin-4		39
Cerebral infarction	Right or left jugular vein	EDTA	1600 g, 15 min, RT		mirVana PARIS	miR-124	TaqMan RT-qPCR		miR-124	miR-451, 5S-snRNA	40
Pancreatic injury	Caval vein	K ₂ -EDTA, 30 min-2 h, RT	1200 g, 20 min, 4°C		100 μL/ TRI Reagent BD	Cel-miR-39	TaqMan RT-qPCR		Cel-miR-39		41
Middle cerebral artery occlusion	Vena cava	EDTA	±2000 g, 20 min, 4 °C/ -70°C		TRIzol	NanoDrop 1000	TaqMan RT-qPCR		Calibration curves of control miRNA diluted into yeast RNA		42
Myocardial Injury	Tail vein	EDTA	?/ -70°C		200 μL/ mirVana PARIS	³² P-miR208	TaqMan RT-qPCR		miR-208/ 2^(35-Ct)	5S-snRNA	43

References

1. Pu XY, Shen JY, Deng ZP, et al. Plasma-specific microRNA response induced by acute exposure to aristolochic acid I in rats. *Arch Toxicol* 2017;91:1473-1483.
2. Xu W, Zhou Y, Xu G, et al. Transcriptome analysis reveals non-identical microRNA profiles between arterial and venous plasma. *Oncotarget* 2017;8:28471-28480.
3. Bai X, Geng J, Zhou Z, et al. MicroRNA-130b improves renal tubulointerstitial fibrosis via repression of Snail-induced epithelial-mesenchymal transition in diabetic nephropathy. *Sci Rep* 2016;6:20475.
4. Brianza-Padilla M, Carbo R, Arana JC, et al. Inflammation Related MicroRNAs Are Modulated in Total Plasma and in Extracellular Vesicles from Rats with Chronic Ingestion of Sucrose. *Biomed Res Int* 2016;2016:2489479.
5. Delic D, Eisele C, Schmid R, et al. Characterization of Micro-RNA Changes during the Progression of Type 2 Diabetes in Zucker Diabetic Fatty Rats. *Int J Mol Sci* 2016;17.
6. Jiang C, Yu H, Sun Q, et al. Extracellular microRNA-21 and microRNA-26a increase in body fluids from rats with antigen induced pulmonary inflammation and children with recurrent wheezing. *BMC Pulm Med* 2016;16:50.
7. Kondratov K, Kurapeev D, Popov M, et al. Heparinase treatment of heparin-contaminated plasma from coronary artery bypass grafting patients enables reliable quantification of microRNAs. *Biomol Detect Quantif* 2016;8:9-14.

8. Pierce LM, Kurata WE, Matsumoto KW, et al. Long-term epigenetic alterations in a rat model of Gulf War Illness. *Neurotoxicology* 2016;55:20-32.
9. Oda H, Ikeguchi R, Yurie H, et al. Plasma microRNAs Are Potential Biomarkers of Acute Rejection After Hindlimb Transplantation in Rats. *Transplant Direct* 2016;2:e108.
10. Siracusa J, Koulmann N, Bourdon S, et al. Circulating miRNAs as Biomarkers of Acute Muscle Damage in Rats. *Am J Pathol* 2016;186:1313-1327.
11. Sun SQ, Zhang T, Ding D, et al. Circulating MicroRNA-188, -30a, and -30e as Early Biomarkers for Contrast-Induced Acute Kidney Injury. *J Am Heart Assoc* 2016;5.
12. Takeuchi-Yorimoto A, Yamaura Y, Kanki M, et al. MicroRNA-21 is associated with fibrosis in a rat model of nonalcoholic steatohepatitis and serves as a plasma biomarker for fibrotic liver disease. *Toxicol Lett* 2016;258:159-167.
13. Wolenski FS, Shah P, Sano T, et al. Identification of microRNA biomarker candidates in urine and plasma from rats with kidney or liver damage. *J Appl Toxicol* 2017;37:278-286.
14. Zhang L, Xu Y, Xue S, et al. Implications of dynamic changes in miR-192 expression in ischemic acute kidney injury. *Int Urol Nephrol* 2017;49:541-550.

15. Chevillet JR, Khokhlova TD, Giraldez MD, et al. Release of Cell-free MicroRNA Tumor Biomarkers into the Blood Circulation with Pulsed Focused Ultrasound: A Noninvasive, Anatomically Localized, Molecular Liquid Biopsy. *Radiology* 2017;283:158-167.
16. Hou X, Tian J, Geng J, et al. MicroRNA-27a promotes renal tubulointerstitial fibrosis via suppressing PPARgamma pathway in diabetic nephropathy. *Oncotarget* 2016;7:47760-47776.
17. Peng Q, Collette W, 3rd, Giddabasappa A, et al. Editor's Highlight: Plasma miR-183/96/182 Cluster and miR-124 are Promising Biomarkers of Rat Retinal Toxicity. *Toxicol Sci* 2016;152:273-283.
18. Thompson KL, Boitier E, Chen T, et al. Absolute Measurement of Cardiac Injury-Induced microRNAs in Biofluids across Multiple Test Sites. *Toxicol Sci* 2016;154:115-125.
19. Banerjee S, Das RK, Giffear KA, et al. Permanent uncoupling of male-specific CYP2C11 transcription/translation by perinatal glutamate. *Toxicol Appl Pharmacol* 2015;284:79-91.
20. Glineur SF, De Ron P, Hanon E, et al. Paving the Route to Plasma miR-208a-3p as an Acute Cardiac Injury Biomarker: Preclinical Rat Data Supports Its Use in Drug Safety Assessment. *Toxicol Sci* 2016;149:89-97.
21. Gutierrez-Escolano A, Santacruz-Vazquez E, Gomez-Perez F. Dysregulated microRNAs involved in contrast-induced acute kidney injury in rat and human. *Ren Fail* 2015;37:1498-1506.

22. Hu D, Yu Y, Wang C, et al. microRNA-98 mediated microvascular hyperpermeability during burn shock phase via inhibiting FIH-1. *Eur J Med Res* 2015;20:51.
23. Roncon P, Soukupova M, Binaschi A, et al. MicroRNA profiles in hippocampal granule cells and plasma of rats with pilocarpine-induced epilepsy--comparison with human epileptic samples. *Sci Rep* 2015;5:14143.
24. Schlosser K, Taha M, Deng Y, et al. Discordant Regulation of microRNA Between Multiple Experimental Models and Human Pulmonary Hypertension. *Chest* 2015;148:481-490.
25. Wu Q, Li JV, Seyfried F, et al. Metabolic phenotype-microRNA data fusion analysis of the systemic consequences of Roux-en-Y gastric bypass surgery. *Int J Obes (Lond)* 2015;39:1126-1134.
26. Zhao R, Qian L, Jiang L. Identification of retinopathy of prematurity related miRNAs in hyperoxia-induced neonatal rats by deep sequencing. *Int J Mol Sci* 2014;16:840-856.
27. Nishimura Y, Kondo C, Morikawa Y, et al. Plasma miR-208 as a useful biomarker for drug-induced cardiotoxicity in rats. *J Appl Toxicol* 2015;35:173-180.
28. Takeuchi J, Sakamoto A, Takizawa T. Sevoflurane anesthesia persistently downregulates muscle-specific microRNAs in rat plasma. *Int J Mol Med* 2014;34:291-298.
29. Wang JF, Zha YF, Li HW, et al. Screening plasma miRNAs as biomarkers for renal ischemia-reperfusion injury in rats. *Med Sci Monit* 2014;20:283-289.

30. Werner W, Sallmon H, Leder A, et al. Independent effects of sham laparotomy and anesthesia on hepatic microRNA expression in rats. *BMC Res Notes* 2014;7:702.
31. Yamaura Y, Nakajima M, Tatsumi N, et al. Changes in the expression of miRNAs at the pericentral and periportal regions of the rat liver in response to hepatocellular injury: comparison with the changes in the expression of plasma miRNAs. *Toxicology* 2014;322:89-98.
32. Gorter JA, Iyer A, White I, et al. Hippocampal subregion-specific microRNA expression during epileptogenesis in experimental temporal lobe epilepsy. *Neurobiol Dis* 2014;62:508-520.
33. Dickinson BA, Semus HM, Montgomery RL, et al. Plasma microRNAs serve as biomarkers of therapeutic efficacy and disease progression in hypertension-induced heart failure. *Eur J Heart Fail* 2013;15:650-659.
34. Endo K, Naito Y, Ji X, et al. MicroRNA 210 as a biomarker for congestive heart failure. *Biol Pharm Bull* 2013;36:48-54.
35. Peng Q, Huang W, John-Baptiste A. Circulating microRNAs as biomarkers of retinal toxicity. *J Appl Toxicol* 2014;34:695-702.
36. Starckx S, Batheja A, Verheyen GR, et al. Evaluation of miR-122 and other biomarkers in distinct acute liver injury in rats. *Toxicol Pathol* 2013;41:795-804.
37. Wang Y, Tang N, Hui T, et al. Identification of endogenous reference genes for RT-qPCR analysis of plasma microRNAs levels in rats with acetaminophen-induced hepatotoxicity. *J Appl Toxicol* 2013;33:1330-1336.

38. Yamaura Y, Nakajima M, Takagi S, et al. Plasma microRNA profiles in rat models of hepatocellular injury, cholestasis, and steatosis. *PLoS One* 2012;7:e30250.
39. Montgomery RL, Hullinger TG, Semus HM, et al. Therapeutic inhibition of miR-208a improves cardiac function and survival during heart failure. *Circulation* 2011;124:1537-1547.
40. Weng H, Shen C, Hirokawa G, et al. Plasma miR-124 as a biomarker for cerebral infarction. *Biomed Res* 2011;32:135-141.
41. Kong XY, Du YQ, Li L, et al. Plasma miR-216a as a potential marker of pancreatic injury in a rat model of acute pancreatitis. *World J Gastroenterol* 2010;16:4599-4604.
42. Laterza OF, Lim L, Garrett-Engele PW, et al. Plasma MicroRNAs as sensitive and specific biomarkers of tissue injury. *Clin Chem* 2009;55:1977-1983.
43. Ji X, Takahashi R, Hiura Y, et al. Plasma miR-208 as a biomarker of myocardial injury. *Clin Chem* 2009;55:1944-1949.