A Tonoplast $P_{3B}$-ATPase Mediates Fusion of Two Types of Vacuoles in Petal Cells


DOI
10.1016/j.celrep.2017.05.076

Publication date
2017

Document Version
Final published version

Published in
Cell Reports

License
CC BY-NC-ND

Citation for published version (APA):

General rights
It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations
If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: https://uba.uva.nl/en/contact, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.

UvA-DARE is a service provided by the library of the University of Amsterdam (https://dare.uva.nl)
A Tonoplast P$_{3B}$-ATPase Mediates Fusion of Two Types of Vacuoles in Petal Cells

**Graphical Abstract**

**Highlights**
- Multiple vacuoles coexist and exchange material in plant petal cells
- The presence of additional vacuoles is controlled by regulators of pigmentation
- The P-ATPases and vacuolar SNAREs interact with each other
- Vacuolar P-ATPases and SNAREs control protein traffic from one vacuole to the other

**Authors**
Marianna Faraco, Yanbang Li, Shuangjiang Li, ..., Walter Verweij, Ronald Koes, Francesca M. Quattrocchio

**Correspondence**
r.koes@uva.nl (R.K.), f.quattrocchio@uva.nl (F.M.Q.)

**In Brief**
Faraco et al. describe vacuolinos, small vacuoles coexisting with the CV in petal epidermal cells. Vacuolino formation is co-regulated with pigmentation and vacuolar acidification. Vacuolar proteins traffic through vacuolinos, and their delivery to the CV requires balanced expression of vacuolar SNAREs and P-ATPases interacting on the tonoplast.

**Accession Numbers**
KY196467
KY196466
A Tonoplast P3B-ATPase Mediates Fusion of Two Types of Vacuoles in Petal Cells

Marianna Faraco, Yanbang Li, Shuangjiang Li, Cornelis Spelt, Gian Pietro Di Sansebastiano, Lara Reale, Francesco Ferranti, Walter Verweij, Ronald Koes, and Francesca M. Quattrocchio

1Plant Development and EpiGenetics, Swammerdam Institute for Life Science, University of Amsterdam, Science Park 904, 1098 XH Amsterdam, the Netherlands
2Section of Genetics, Department of Molecular Cell Biology, VU University, de Boelelaan 1087, 1081 HV Amsterdam, the Netherlands
3Department of Biotechnology and Environmental Sciences, University of Salento, via Monteroni, Centro Ecotekne, 73100 Lecce, Italy
4Department of Agricultural, Food, and Environmental Sciences, University of Perugia, Borgo XX Giugno 74, 06121 Perugia, Italy
5Present address: ENZA Zaden, Haling 1E, 1602 DB Enkhuizen, the Netherlands
6These authors contributed equally
7Lead Contact
*Correspondence: r.koes@uva.nl (R.K.), f.quattrocchio@uva.nl (F.M.Q.)
http://dx.doi.org/10.1016/j.celrep.2017.05.076

SUMMARY

It is known that plant cells can contain multiple distinct vacuoles; however, the abundance of multivacuolar cells and the mechanisms underlying vacuolar differentiation and communication among different types of vacuoles remain unknown. PH1 and PH5 are tonoplast P-ATPases that form a heteromeric pump that hyper-acidifies the central vacuole (CV) of epidermal cells in petunia petals. Here, we show that the sorting of this pump and other vacuolar proteins to the CV involves transit through small vacuoles: vacuolinos. Vacuolino formation is controlled by transcription factors regulating pigment synthesis and transcription of PH1 and PH5. Trafficking of proteins from vacuolinos to the central vacuole is impaired by misexpression of vacuolar SNAREs as well as mutants for the PH1 component of the PH1-PH5 pump. The finding that PH1-PH5 and these SNAREs interact strongly suggests that structural tonoplast proteins can act as tethering factors in the recognition of different vacuolar types.

INTRODUCTION

In Petunia hybrida and many other species, epidermal petal cells display color to attract pollinators (Galliot et al., 2006). Color depends on synthesis, chemical modification, vacuolar sequestration of anthocyanins and co-pigments (e.g., flavonols), and the pH of the vacuolar lumen (Koes et al., 2005). Petunia mutants with blue petals define seven loci (PH1 to PH7) required for vacuolar acidification in epidermal petal cells (Quattrocchio et al., 2006). PH3 and PH4 encode WRKY and myeloblastosis (MYB) transcription factors, respectively, that interact with basic-helix-loop-helix (bHLH) and WD40 transcription activators encoded by ANTHOCYANIN1 (ANT1) and AN11 to activate expression of ~40 genes (unpublished results), including PH1 and PH5. PH5 is a tonoplast P3B-ATPase proton pump that interacts with the P3B-ATPase PH1 to acidify the vacuolar lumen (Faraco et al., 2014; Li et al., 2016; Verweij et al., 2008).

In plants constitutively expressing 3SS:PH5-GFP, most of PH5-GFP resides in the tonoplast of the central vacuole (CV) in all cell types (Verweij et al., 2008). By studying the cellular localization of the components of the PH1-PH5 pump, we found a peculiar compartment that represent a station of transit for protein directed to the vacuole in petal epidermal cells: vacuolino. This compartment, and the collection of mutants affecting its formation or fusion to the CV, provide access to the regulation of formation of multiple vacuoles and the coexistence of distinct vacuoles in a single cell.

RESULTS

Vacuolar Proteins Are Sorted through Vacuolinos to the CV in Epidermal Petal Cells

In agroinfiltrated petunia leaves, PH5-GFP localized after 24 hr on the tonoplast of epidermal cells (Figure 1A) by the “canonical” pathway described in a variety of tissues and species. In epidermal petal cells, however, PH5-GFP localized within 24 hr in vacuole-like structures (“vacuolinos,” Italian for “small vacuoles”) with a diameter of 1–15 μm but was absent from the tonoplast of the CV containing the anthocyanins (Figure 1A). Only 48 hr after transformation did PH5-GFP reach the CV. PH1-GFP (GFP in a predicted cytoplasmic loop; Faraco et al., 2014) moved in a similar way (Figure 1A).

Within the petal epidermis of transgenic plants expressing the same 3SS:PH5-GFP, transgene expression remained lower than that of the PH5 endogene, both at mRNA and protein level, probably explaining why 3SS:PH5-GFP rescued the ph5 flower color only partially (Verweij et al., 2008). Epidermal petal cells of these plants also contain vacuolinos marked by PH5-GFP (Figure 1B), indicating that sorting through vacuolinos is the natural pathway by which proteins reach the CV and not the result of jamming of the sorting system or other artifacts.
To investigate whether this trafficking pathway was specific for PH5 and PH1, we studied the sorting of PhSYP22 and PhSYP51 (petunia orthologs of, respectively, the vacuolar QA-SNARE paralogs AtSYP21 and AtSYP22 (Jemura et al., 2004) and the Qc-SNARE paralogs AtSYP51 and AtSYP52 from Arabidopsis (De Benedictis et al., 2013; Figure S1A). Both are expressed in all plant parts independent from AN1, AN11, PH3, and PH4 (Figures S1B and S1C). In plant cells, GFP-PhSYP22 and GFP-PhSYP51 fusions remain largely intact and membrane-bound (Figure S1D); therefore, they are reliable localization markers. In epidermal petal cells, GFP and red fluorescent protein (RFP) fusions of PhSYP22 and PhSYP51, like PH5-GFP, localized on the membrane of vacuolinos 24 hr after agroinfiltration of intact petals or transformation of petal protoplasts and reached the tonoplast of the CV 48 hr after transformation (Figures 1A and 1B).

The heterologous proteins AtSYP52 (Jemura et al., 2004) and AtKCO1 (Czeminski et al., 2002) from Arabidopsis (Figures S2A and S2B) and the soluble protein ALEU-GFP (Di Sansebastiano et al., 2001) also trafficked via the vacuolinos in petal epidermal cells. ALEU-GFP, PhSYP51, and PhSYP22 labeled, in addition, some punctate structures (Figures 1A and 1C), probably prevacuolar compartments (PVCs), which have a diameter ranging from 100–250 nm (Marty, 1999; Paris et al., 1996), or endosomes en route to vacuolinos. However, the plasma membrane marker RFP-AtSYP122 (Assaad et al., 2004) reached the cell outer membrane within 24 hr after transformation (Figure 1A), indicating that the transit via vacuolinos is specific for vacuolar proteins.

Vacuolinos Are Petal Epidermis-Specific and Exist in Different Plant Species

Petal protoplasts are a mixture of epidermal cells (with anthocyanin-containing vacuoles) and mesophyll cells (upigmented; Faraco et al., 2011). In epidermal protoplasts, PH5-GFP (and other vacuolar proteins) mimics the trafficking in infiltrated petals (Figure 1C). In mesophyll cells, vacuolar proteins reached the CV within 24 hr, and no vacuolino-like structures were seen, indicating that the sorting via vacuolinos is cell type-specific (Figure 1C).

In freshly prepared petal protoplasts, bodies distinct from the central vacuole are visible (Figure 1D), indicating that vacuolinos are present before the introduction of transgenes. Similar structures are also found in epidermal protoplasts from rose (Rosa hybridra) petals (Figure S4A), and transiently expressed PH5-GFP localized after ~24 hr to these compartments before reaching the CV after ~48 hr (Figure 1E). Because roses (rosids) and petunias (asterids) belong to divergent clades of eudicots, this suggests that vacuolinos are widespread, at least among dicots.

Co-localization experiments in petunia epidermal petal cells and derived protoplasts showed that all vacuolar protein fusions co-localize in the same small compartment(s), indicating that vacuolinos constitute a homogeneous population (Figure 1F).

TIPs Are Not Sorted to Vacuolinos or the CV in Petal Cells

Tonoplast intrinsic proteins (TIPs) have been observed on different types of vacuoles (Jauh et al., 1999; Paris et al., 1996). To test whether TIPs differentially mark vacuolinos and the central vacuole, we used GFP and yellow fluorescent protein (YFP) fusions for α, δ, and γ TIPs from Arabidopsis that were previously used by others (Hunter et al., 2007) as well as fusions of homologous TIPs from petunia (PigTIP, Peinf101Scf02502 g03029;1; PiγTIP, Peinf101Scf00487 g13016;1; PiδTIP, Peinf101Scf02095 g01012.1), identified via searches of the petunia genome (Bom barely et al., 2016) and phylogenetic analyses (Figure S2C). In petal protoplasts, the Arabidopsis TIPs (ATIPs) marked the endoplasmic reticulum (ER) of epidermal and mesophyll cells after 24 and 48 hr, colocalizing with the ER marker GFP-KDEL, also on bright small vacuole-like compartments (2–3 μm). These also contained KDEL-RFP (Figure S2D) and may be similar to the α-TIP-positive PSV-like compartments observed in leaf cells (Park et al., 2004). The petunia TIPs (PTIPs) colocalized with their Arabidopsis homologs in the ER and the PVC-like compartments but reduced the number of these PSV-like structures substantially.

To address whether the ATIP-positive compartments were (a subclass of) vacuolinos, we co-expressed TIP-RFP fusions and PH5-GFP. In mesophyll cells, PH5-GFP was on the vacuole within 24 hr with little or no overlap with the TIPs. In epidermal petal cells, expression of the TIP-RFP fusions affected the sorting of PH5-GFP in a major fraction of the cells. In most cells, PH5-GFP resided in vacuolinos after 24 hr, which, in many of these TIP-RFP-expressing cells, appeared to be very small.
(<1–3 μm), and reached the CV after 48 hr in only a limited number of cells (2 of 7 cells co-expressing AtiTI1I, 4 of 15 cells co-expressing AtI7TII, and 9 of 15 cells co-expressing AtIzTI1I). This might be due to adverse effects of the aquaporin activity of TIPs, as observed previously (Oufattole et al., 2005), or indicate a role for TIPs in trafficking from vacuolinos to the CV.

Although PH5-GFP and TIP-RFP patterns suggest co-localization in (some) small vacuolino-like structures, we observed no co-localization in vacuolinos of normal size or in the CV. Petal mesophyll cells, which lack vacuolinos, contain TIP-RFP-positive PSV-like structures similar to those observed in epidermal cells. This indicates that the PSV-like structures labeled by the analyzed TIPs are distinct from vacuolinos (Figures 1G and 1H).

**an1, ph3, and ph4 Mutations Abolish Vacuolino Formation but ph1 and ph5 Do Not**

We also investigated vacuolar trafficking in ph mutant petals. Mutations in PH3 and PH4 block expression of the tonoplas pumps PH1 and PH5, affecting vacuolar acidity and petal color (Verweij et al., 2008). After agroinfiltration of ph3 and ph4 petals or transformation of ph3 and ph4 petal protoplasts, GFP-SYP51, PH5-GFP, and PH1-GFP moved in epidermal cells to the CV within 24 hr, and vacuolinos were never seen (Figure 2A; Figures S4B and S4C), resembling the canonical sorting of vacuolar proteins in leaf and petal mesophyll cells. Light and electron microscopy revealed that epidermal cells from wild-type petals contain, within the conical tip, numerous membrane compartments (1–10 μm in size) squeezed between the plasma membrane and the CV. These structures correspond to the vacuolinos because they are absent in ph3 and ph4 mutants (Figures 2B and 2C). This implies that AN1, PH3, and PH4 are required for the formation of vacuolinos, explaining why these small vacuoles are specific to epidermal petal cells.

Among the ~40 genes regulated by the AN1-PH4-PH3 complex are PH1 and PH5 (Faraco et al., 2014; Verweij et al., 2008, 2016). Co-expression of PH1 and PH5 is sufficient to restore vacuolar acidification and (reddish) petal color in ph3 and ph4 mutants (Faraco et al., 2014). However, it did not rescue the formation of and the protein sorting via vacuolinos because transiently expressed GFP-PhSYP51 localized in epidermal cells of ph3 35S:PH1 35S:PH5 petals within 24 hr in the CV and vacuolinos was not seen (Figure 2D), as in ph3 petals. Furthermore, a ph5 loss-of-function mutation did not abolish the formation of vacuolinos or the sorting of transiently expressed GFP-PhSYP51 to these compartments (Figures 2A–2C). These findings imply that the hyper-acidification of the CV or precursor vacuoles by the PH1/PH5 pump is neither necessary nor sufficient for the formation of and protein trafficking via vacuolinos.

**PH1 Is Necessary for Traffic of Proteins from the Vacuolinos to the CV**

Because no isogenic PH1 controls are available for ph1 mutants, we analyzed ph1 mutants in two different genetic backgrounds. In both backgrounds, GFP-PhSYP51 and PH5-GFP localized on vacuolinos in epidermal petal cells 24 hr after agroinfection, but did not reach the CV after 48 hr or more and remained instead on vacuolinos, which had increased in size and number (Figure 3A). We observed the same in ph1 petal epidermis protoplasts transiently expressing GFP-PhSYP22 or PH5-GFP, whereas, in mesophyll protoplasts, vacuolar proteins moved to the CV within 24 hr (Figure 3B), as in the wild-type (Figure 1B). This implies that PH1 is essential for the trafficking of proteins from vacuolinos to the CV in epidermal petal cells but not for the “direct” transport that, in other cells, delivers proteins to the CV within 24 hr. We confirmed that the trafficking defect in epidermal cells is caused by ph1 by generating isogenic ph1V23 lines with a 35S:PH1 transgene. 35S:PH1 rescued color and vacuolar acidification in the petal epidermis, similar to previous results (Faraco et al., 2014), as well as the trafficking of PH5-GFP to the CV (Figure 3B; Figure S4E). Light and electron microscopy confirmed that the tips of ph1 petal epidermal cells were crowded by vacuolinos (Figures 3C and 3D). This indicates that PH1 is involved in trafficking from the vacuolinos to the CV, possibly by mediating the fusion of the two types of vacuoles.

Membrane fusion relies on SNARE complexes that bridge the membranes. Excess of one of the components of such a complex may titrate away other partners, resulting in non-fusogenic complexes (Burian et al., 2016; Di Sansebastiano, 2013; Weimbs et al., 2003). Wild-type petals agroinfiltrated with a single plasmid co-expressing PH5-GFP with either PhSYP51 or PhSYP22 phenocopied ph1 petals. In these cells, PH5-GFP localized after 24 and 48 hr in the vacuolinos, whereas little or none reached the tonoplast of the CV (Figure 3E). In the same cells, trafficking of RFP-AtSYP122 to the plasma membrane was not affected. Thus, overexpression of these vacuolar SNAREs impairs, or at least strongly delays, fusion of the vacuolinos to the CV, just like ph1.

We tested whether PH1 might interact with PhSYP22 and PhSYP51 by a split ubiquitin-based yeast two-hybrid system (Obrdlik et al., 2004). Co-expression of PH1-Cub (fusion to the C-terminal part of ubiquitin) and Nub-PhSYP22 weakly activated the HIS and lacZ reporter genes, whereas co-expression of PH1-Cub and Nub-PhSYP51 resulted in a stronger response (Figure S5F). Expression of PH5-Cub alone gave some background growth, reduced by addition of methionine to the medium. Co-expression of PH5-Cub and Nub-SYP122 or Nub-SYP51 under the same conditions induced activity of the LacZ reporter. The same holds true for the already reported interactions of PH5...
with PH1 and with itself (Faraco et al., 2014) and the dimerization of AtKAT1 (Obrdlik et al., 2004). Yeast cells expressing Cub or Nub alone or in combination with one of the aforementioned fusions did not significantly activate the reporter.

To study these interactions in planta, we used bimolecular fluorescence complementation (BiFC) (Kerppola, 2009). We expressed PH1, PH5, and the SNAREs as fusions to the N- or C-terminal parts of YFP (nYFP and cYFP, respectively) in petunia petal protoplasts and marked transformed cells by co-expressing RFP-AtSYP122. The nYFP was inserted in a cytoplasmic loop of PH1 (PH1-cYFPi) (Faraco et al., 2014). Expression of PH1-cYFPi alone or in combination with nYFP yielded no YFP fluorescence (Faraco et al., 2014). In nearly all cells co-expressing PH5-nYFP with either PH5-cYFP or PH1-cYFPi, we observed YFP fluorescence on the tonoplast of the CV (Figures 4A and 4B; Faraco et al., 2014). Co-expression of nYFP-PhSYP51 with PH1-cYFPi or PH5-cYFP resulted in YFP fluorescence on the tonoplast, confirming the interaction of PhSYP51 with both PH1 and PH5 (Figure 4). Co-expression of cYFP-PhSYP22 with PH5-nYFP or PH1-nYFPi (Figure 4) resulted in less bright fluorescence than that of PH1xPhSYP51 and PH5xSYP51, suggesting a weaker interaction, as in the yeast two-hybrid (Y2H) assay. All interactions were detectable in pigmented epidermal as well as mesophyll protoplasts (Figure 4A; Figure S5G). Co-expression

Figure 3. Mutation of PH1 and Expression of PhSYP51 or PhSYP22 Block Trafficking from Vacuolinos to the CV
(A) Confocal images of epidermal petal cells from a wild-type and a ph1<sup>123</sup> plant 48 hr after agroinfiltration with GFP-PhSYP51 or PH5-GFP (see also Figures S3B–S3D).
(B) Protoplasts originating from the petal epidermis (top) and mesophyll (bottom) of a ph1<sup>123</sup> and complemented transgenic clone expressing 35S:PH1 48 hr after transformation with a PH5-GFP construct. The insets show enlargements of the regions indicated by the arrows (see also Figure S3E).
(C and D) Light (C) and electron (D) micrographs showing epidermal cells from ph1<sup>123</sup> petals. V, vacuolinos (arrowheads); CV, vacuole. The tips of the conical cells are marked with red asterisks.
(E) Confocal micrographs of wild-type epidermal petal cells 48 hr after agroinfection with PH5-GFP, RFP-ATSYP122, and PhSYP51 or PhSYP22.
Scale bars are 5 and 1 μm in (D) and 20 μm in all other panels.
of nYFP-PhSYP51 with cYFP-PhSYP51 or cYFP-PhSYP22 did not result in YFP fluorescence, indicating that self-assembly of nYFP and cYFP was neglectable (Figure 4C). These results show that PH1 and PH5 interact with PhSYP51 and, to a lesser extent, with PhSYP22 on the membrane of the CV.

DISCUSSION

It has long been known that plant cells can contain multiple vacuoles that can differ in size, protein or ion content, and function, but it is unclear how common (or rare) multivacuolar cells are in plants (Frigerio et al., 2008) because they are only seen in protoplasts derived from unknown cell types in leaves, seeds, or root tips but not in intact plant tissues (Hunter et al., 2007; Olbrich et al., 2007), except for a few highly specialized cells types. Among these are motor cells in motile organs of some legumes (Fleurat-Lessard et al., 1997), leaf cells from a salt-stressed halophyte (Epimashko et al., 2004), senescing leaves (Otegui et al., 2005), and grape cells synthesizing anthocyanins (Gomez et al., 2011). Whether the latter are vacuolar compartments is unclear because no information is available about the proteins residing in these structures.

We demonstrated that (pigmented) epidermal cells in mature petals of diverse species contain multiple small vacuoles (vacuolinos), which differ in size and content from the large CV, and other small TIP-positive PSV-like compartments (Park et al., 2004). The vacuolinos are defined and distinguished from other small vacuoles by the transcription factors (AN1, PH3, and PH4) that govern their formation and fusion to the central vacuole. AN1, PH3, and PH4 activate a limited number (about 40) of downstream genes (Verweij, 2007; unpublished data), which seems insufficient to operate an entire trafficking pathway, considering that several of those are involved in vacuolar hyper-acidification and the stabilization of anthocyanins within the vacuole (Passeri et al., 2016; Quattrocchio et al., 2006). We presume that the vacuolin pathway is an alternative form of a canonical ubiquitous default vacuolar trafficking pathway operating in other tissues, which becomes evident in epidermal petal cells only when AN1, PH3, or PH4 is mutated.

SNAREs, Rabs, and their effectors promote membrane recognition and fusion (Stenmark, 2009; Uemura and Ueda, 2014).
Data suggest that SNARE proteins can physically and/or genetically interact with certain membrane transporters (Honsbein et al., 2008; Martin-Moutot et al., 1996; Pagel et al., 2003; Sassera and Fratti, 2014; Sassera et al., 2012; Zhang et al., 2015). Some of these interactions were shown to regulate the membrane transporter (Honsbein et al., 2009; Zhang et al., 2015), and other data report the effect of transporters on membrane fusion (Sassera et al., 2012), but evidence for a direct role in membrane recognition/fusion is lacking. PH1 is required for trafficking from vacuolinos to the CV, most likely by mediating the fusion of vacuolinos with the CV. This role of PH1 appears to be independent from the proton pumping activity of the PH1-PH5 complex because mutation of PH5, which is essential for electrogenic activity of PH1 (Faraco et al., 2014), does not affect the trafficking from vacuolinos to the CV. This suggests that PH1 operates as a structural component in a tethering or fusogenic complex together with SNAREs. That ph5 did not reduce trafficking of proteins from vacuolinos to the CV, even though PH5 interacts, like PH1, with vacuolar SNAREs (Figure 2), might be explained by the nature of the ph5 alleles used (ph5A2209 and ph5V69), accumulating PH5 transcripts for a truncated protein (Verweij et al., 2008). Alternatively, the function of PH5 in the SNARE complex could be redundant because PH5 is one of 10–12 P3A-ATPases, which are involved in the formation of vacuolinos and/or their fusion to the CV can be identified by straightforward reverse genetic analyses. The viability of an1, ph3, and ph4 plants and the low number of genes misregulated in these mutants suggest that additional factors involved in the formation of vacuolinos and/or their fusion to the CV can be identified by straightforward reverse genetic analyses. Mutations in three additional AN1-PH3-PH4-regulated genes disrupt the vacuolino pathway (unpublished data). In two of these mutants transiently expressed vacuolar proteins remain stuck in puncta and do not reach the vacuolinos or the CV, with no effect on vacuolar trafficking in leaf or petal mesophyll cells. In the third mutant, vacuolar proteins also traffic directly to the CV in epidermal petal cells. This mutation probably affects a protein that operates at or close to the point of bifurcation between the vacuolino and canonical pathway. It might act to promote entrance of vesicles into the vacuolino pathway or to inhibit their entrance into the canonical pathway toward the CV.

**EXPERIMENTAL PROCEDURES**

**Plant Material**

All *Petunia hybrida* lines (from the Amsterdam petunia collection) were grown in a greenhouse at a temperature of 19 C/30 C (minimum/maximum) and a minimum of 16 hr of light in all seasons. Lines W225 (an1TIP22G), R149 (ph4V215S), and R159 (ph4A2209) were derived from R27 (wild-type) by transposon insertion and excisions. We used the (transformable) F1 hybrids M1xV30 (wild-type), R143xR144 (ph3A46B5B26E79P), and V69xR159 (ph5A2209V69P) and a homozygous ph1V231V23 individual of the F2 cross V23xV30 or a ph1T2164I6. Further details on these alleles can be found elsewhere (Faraco et al., 2014; Quattrociocchi et al., 2006; Spelt et al., 2000; Verweij et al., 2008, 2016).

**Transient Expression Assays**

Agrominification of intact tissues, transient transformation of leaf and petal protoplasts (Faraco et al., 2011), and generation of transgenic plants were described previously. Petals from flowers that had just opened (stage 6) were used for protoplast preparation and agro-infection.

**Gene Constructs**

The full-length PhSYSP51 sequence, amplified from petal RNA of line R27 with primers 4655 and 4675 (Table S1) was cloned in pENTRY/D-TOPO (Invitrogen) and recombined (Gateway, Invitrogen) into specific destination vectors (Karimi et al., 2002, 2005) to give all expression clones.

Primers (Table S1) were designed to amplify the full-size cDNA of the petunia PhSYSP22 from R27 petals and produce expression constructs as described above for PhSYSP51.

Phylogenetic analyses were done as previously described (Li et al., 2016). Other constructs are described elsewhere as follows: AKCO1-GFP (Assaad et al., 2004; Czempinski et al., 2002; Faraco et al., 2014; Fluckiger et al., 2003; Verweij et al., 2008, 2010), Aleu-GFP (Assaad et al., 2004; Czempinski et al., 2002; Faraco et al., 2014; Fluckiger et al., 2003; Verweij et al., 2008), RFP-ASIYP2122 (Assaad et al., 2004; Czempinski et al., 2002; Faraco et al., 2014; Fluckiger et al., 2003; Verweij et al., 2008), PH5-GFP (Assaad et al., 2004; Czempinski et al., 2002; Faraco et al., 2014; Fluckiger et al., 2003; Verweij et al., 2008), and Arabidopsis TIP-GFP fusions (Hunter et al., 2007).

The coding sequences of petunia αTIP, βTIP, and γTIP were amplified from genomic DNA of *P. inflata* (line S6) using the primers listed in Table S1 and recombined in pK7RWG2.0 (Karimi et al., 2002), in which the GFP coding sequence was replaced by the RFP coding sequence. RFP fusions for Arabidopsis TIPs were constructed in a similar way.

The accession numbers for the sequences of PtiTIP, PiiTIP, and PγTIP reported in this paper are Sol Genomics: Peinf101Scf02932 g03029.1, Peinf101Scf02909 g01012.1, and Peinf101Scf00487 g13016.1, respectively.

**Split Ubiquitin Assay**

Split ubiquitin assays were performed as described previously (Obrdlik et al., 2004). The constructs for PH1 and PH5 were described previously (Faraco et al., 2014).

B1 and B2 sequences were added to SNAP-αTIP with PcDNA III using the following primers: 5750 and 5751 to amplify PhSYP22 and 5752 and 5753 (Table S1) to amplify PhSYSP51. The PCR products were then recombined in split ubiquitin vectors (Obrdlik et al., 2004).

**Confocal Microscopy**

Protoplasts and intact tissue were imaged with a confocal laser microscope (Zeiss, LSM510) (Faraco et al., 2014).

**Microscopy of Petal Semi-thin Sections**

Petal limbs were fixed in 5% (w/v) glutaraldehyde in 75 mM sodium cacodylate buffer (pH 7.2) for 24 hr, washed four times for 15 min each in 75 mM cacodylate buffer (pH 7.2), post-fixed in 1% (w/v) OsO4 for 90 min, dehydrated in increasing concentrations of ethanol, and included in resin (Epon, 2-dodecenediosuccinic anhydride, and methylendiacryl anhydride mixture). Semi-thin sections (1–2 μm) were cut by ultramicrotome (OmU2, Reichert) equipped with a glass blade, stained with toluidine blue, and observed under a light microscope (DMLB, Leica Microsystems). Transmission electron microscopy of petal cells was performed as previously reported (Verweij et al., 2008).

**ACCESSION NUMBERS**

The accession numbers for the sequences of the Petunia hybrida vacuolar SNAREs PhSYSP22 and PhSYSP51 reported in this paper are GenBank: KY196467 and KY196466, respectively.

**SUPPLEMENTAL INFORMATION**

Supplemental information includes four figures and one table and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2017.05.078.
AUTHOR CONTRIBUTIONS

R.K. and F.M.Q. conceived and supervised the project. M.F., Y.L., R.K., and F.M.Q. wrote the paper with input from all other authors. M.F., Y.L. and S.L. performed most of the experiments. W.V. and G.P.D.S. performed the early localization studies, which revealed the existence of vacuolins. C.S. generated constructs and performed the Y2H analysis. L.R. and F.F. performed light and electron microscopy.

ACKNOWLEDGMENTS

We thank Erik Manders and Ronald Breedijk (Center of Advanced Microscopy, University of Amsterdam) for technical assistance with confocal microscopy, Daisy Kloos and Pieter Hoogeveen for plant care, and Lorenzo Frigerio for supplying the ATIPs-YFP fusions. This work was supported by short-term EMBOfellowships (to M.F. and W.V.), fellowships of the Chinese Scholarship Council (to Y.L. and S.L.), and project n 14 “Reti di Laboratori Pubblici di Ricerca, SELGE,” Regione Puglia (to G.P.D.S.).

REFERENCES


Supplemental Information

A Tonoplast P$_{3B}$-ATPase Mediates Fusion of Two Types of Vacuoles in Petal Cells

Marianna Faraco, Yanbang Li, Shuangjiang Li, Cornelis Spelt, Gian Pietro Di Sansebastiano, Lara Reale, Francesco Ferranti, Walter Verweij, Ronald Koes, and Francesca M. Quattrocchio
Figure S1. Characterization of PhSY P22 and PhSY P51, Related to Figure 1.

(A) Phylogenetic tree of SNARE proteins from different plant species: Ph, Petunia hybrida; Pa, Petunia axillaris (Bombarely et al., 2016) At, Arabidopsis thaliana; Sc, Saccharomyces cerevisiae; St, Solanum tuberosum; Os, Oryza sativa; Sl, Solanum lycopersicum; Nt, Nicotiana tabacum; Cr, Capsella rubella, Bn, Brassica napus. The two petunia vacuolar SNAREs PhSY P22 and PhSY P51 are marked in red. Sequence alignments were generated with MUSCLE and phylogenetic trees were constructed with Maximum likelihood (PhyML) after curation of the alignments by the G-blocks tool, and then rendered by TreeDyn on line (http://phylogeny.lirmm.fr/phylo_cgi/index.cgi). Branch support is expressed as percentage of 300 bootstraps.

(B) RT–PCR analysis of PhSY P22 and PhSY P51 mRNA in wild type petunia (line V30) in different tissues and different developmental stages. “Throat” is the part of the corolla at the junction of the petal limb and the tube.

(C) RT–PCR analysis of PhSY P22 and PhSY P51 in petals of opening flower buds (stage 5) from different genotypes. GAPDH (GLYCLERALDEHYDE 3-PHOSPHATE DEHYDROGENASE) was used as a constitutively expressed control. None of the two genes seems to be regulated by any of the regulatory genes involved in petal pigmentation or vacuolar acidification.

(D) Western blot analysis of protein fractions (M; membrane fraction, S; soluble fraction) obtained from GFP-PhSY P22 and GFP-PhSY P51 expressing petal protoplasts of wild type. Proteins were detected using an anti-GFP serum. Arrows indicate free GFP, which is in low amount and only visible in the soluble fraction.
Figure S2. Intracellular trafficking of AtKCOI, PhSYP51 and TIP proteins from the α-, β- and γ-subfamilies, Related to Figure 1.

(A) GFP-AtSYP52 localization in epidermal cells of intact petals 24 and 48 hours after agroinfiltration. Fluorescence of GFP-AtSYP52 is shown in green and anthocyanin autofluorescence in blue.

(B) AtKCOI-GFP localization in leaf and petal epidermis 24 hours after agroinfiltration. In leaf epidermis GFP signal is on the tonoplast, while in the petal epidermal cells it is on the membrane of vacuolinos. AtKCOI-GFP fluorescence is shown in green and of anthocyanins in blue.

(C) Phylogenetic tree of α, γ and δ TIP proteins from different species (At: Arabidopsis thaliana, Pi: Petunia inflata, Sl: Solanum lycopersicum, St: Solanum tuberosum). Sequence alignments were generated with MUSCLE and phylogenetic trees were constructed with Maximum likelihood (PhyML). After curation of the alignments by G-blocks tool, rendered by TreeDyn on line (http://phylogeny.lirmm.fr/phylo_cgi/index.cgi). Branch support is the percentage of 300 bootstraps.

(D,E) Localization of FP-tagged TIPs from Arabidopsis and petunia in petunia protoplasts from petal epidermis and petal mesophyll. RFP-KDEL was co-expressed to mark the ER. Fluorescence of Arabidopsis TIP-YFP fusions is shown in green, that of RFP-KDEL and petunia TIP-RFP fusions in red and of anthocyanins and chlorophyll in blue. The rightmost panel (yellow frame) shows an enlargement of the marked part in the leftmost image taken with lower laser intensity to better distinguish the membranes surrounding small brightly fluorescent compartments. The composite panel shows the merged image from the red (RFP), green (YFP) and blue (anthocyanin and chloroplasts) channels. Scale bars equal 20 µm in all panels.
Figure S3. RFP fusions of d and gTIPs from Arabidopsis and petunia do not co-localize with PH5-GFP and affect its sorting to the vacuole in epidermal petal cells, Related to Figure1. PH5-GFP was cotransformed with Arabidopsis or petunia RFP-fusions of TIPs from the α, δ and γ families. Fluorescence of PH5-GFP is shown in green, that of TIP-RFP fusions in red and of anthocyanins and chlorophyll in blue. The composite panel shows the merged image from the red, green and blue channel. Scale bars equal 20µm in all panels.
Figure S4. Vacuolinos formation and fusion, Related to Figures 1, 3 and 4.
(A) Light and confocal micrographs of freshly prepared rose petal protoplasts. Vacuolinos are visible and the CV is recognizable from the anthocyanins in the lumen (blue fluorescence).
(B) Localization of
PH5-GFP in epidermal cells of an1, ph3, ph4 or ph1 petals 24 hours after agroinfiltration. (C) PH1-GFPi in petal protoplasts from ph3 and ph4 petals 24 hours after transformation. (D) GFP-PhSYP22 in ph1 protoplasts shows that the fusion protein is stuck on the membrane of the vacuolinos in epidermal pigmented cells, while it normally reaches the tonoplast of the CV in uncolored mesophyll cells (white arrowhead). (E) Localization of PH5-GFP in petunia protoplasts from petals of a ph1127 mutant and a derived transgenic line transformed with the 35S:PH1 construct. Magenta arrowheads indicate protoplasts derived from the petal epidermis, orange arrowheads indicate mesophyll protoplasts. Scale bars equal 20µm in all panels in A, B and C. (F) Split ubiquitin based Yeast 2 Hybrid assay. Cells expressing different combinations of Nub (N-terminal part of Ubiquitin) and Cub (C terminal part of ubiquitin) fusions were grown on distinct media to select for interactions. The addition of 75µM methionine lowers the strong background given by the PH5-Cub construct. (G) Bimolecular fluorescence complementation in protoplasts of petunia petals shows that all interactions are detectable in colored cells from the epidermis as well as in white cells from the other cell layers. RFP-AtSYP122 marks the plasma membrane and serves as marker of transformation. Scale bars are 10µm.
Table S1: Primers used to amplify and to recombine the full length *PhSYP51*, *PhSYP22* and petunia *TIP* cDNAs in Gateway vectors, Related to Experimental Procedure.

<table>
<thead>
<tr>
<th>name</th>
<th>gene</th>
<th>sequence 5' - 3' ori</th>
</tr>
</thead>
<tbody>
<tr>
<td>4520</td>
<td><em>PhSYP51</em></td>
<td>CATCGCCTGATCAAAAAGAGGT F</td>
</tr>
<tr>
<td>4521</td>
<td><em>PhSYP51</em></td>
<td>ATGGCTTCTTCTGGAGAAGCTCAGTGGATCCAGG F</td>
</tr>
<tr>
<td>4522</td>
<td><em>PhSYP51</em></td>
<td>AGCTTCTGGAAAGAAGGCTC T</td>
</tr>
<tr>
<td>4523</td>
<td><em>PhSYP51</em></td>
<td>TGGCTTGCAAAAGCAGACTCAT R</td>
</tr>
<tr>
<td>4533</td>
<td><em>PhSYP51</em></td>
<td>TTCTGGCATTGTGGTCTGC R</td>
</tr>
<tr>
<td>4655</td>
<td><em>PhSYP51</em></td>
<td>CACCATGGCTTCTTCTGGA F</td>
</tr>
<tr>
<td>4656</td>
<td><em>PhSYP51</em></td>
<td>CACCGGTCTTGACAATTCTG G</td>
</tr>
<tr>
<td>4657</td>
<td><em>PhSYP51</em></td>
<td>TTATCCACCCTTTGTGCGTC F</td>
</tr>
<tr>
<td>4658</td>
<td><em>PhSYP51</em></td>
<td>TTACATATACTTAACCAACAT TCCA R</td>
</tr>
<tr>
<td>4675</td>
<td><em>PhSYP22</em></td>
<td>TTACAGATACTTGATCAACATATATATTA CC R</td>
</tr>
<tr>
<td>4670</td>
<td><em>PhSYP22</em></td>
<td>ATGAGCTTTCAAGATCTAGA ATCGG F</td>
</tr>
<tr>
<td>4671</td>
<td><em>PhSYP22</em></td>
<td>CAGATGCTAAACTTGCTAAA GTATT F</td>
</tr>
<tr>
<td>4672</td>
<td><em>PhSYP22</em></td>
<td>CGTTCCCTCTATAATAGCTTC ATTAAAG R</td>
</tr>
<tr>
<td>4673</td>
<td><em>PhSYP22</em></td>
<td>AGCCATGATCGATGATATTG GTTC F</td>
</tr>
<tr>
<td>4674</td>
<td><em>PhSYP22</em></td>
<td>AGTTGCAGCATGAGAATTCT CA R</td>
</tr>
<tr>
<td>4794</td>
<td><em>PhSYP22</em></td>
<td>ATTTCCTTTAATGAAGCTAT TATAGAGG</td>
</tr>
<tr>
<td>4798</td>
<td><em>PhSYP22</em></td>
<td>GGGGACAAGTTTGTACAAAAAAGCAGGCTTTATGAGCT TCAAGATCTA</td>
</tr>
<tr>
<td>4799</td>
<td><em>PhSYP22</em></td>
<td>GGGGACCACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>4800</td>
<td><em>PhSYP51</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>4863</td>
<td><em>PhSYP22</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>5570</td>
<td><em>PhSYP22</em></td>
<td>ACAAGTTTGTACAAAAAAGCAGGCTTCTCACCACCAACCAT GAGCTTCTCACCACCACCATAGT</td>
</tr>
<tr>
<td>5571</td>
<td><em>PhSYP22</em></td>
<td>TCCGCCACCACCAACCATTTGTACACAAGATGTTACAAAGAAGGCTGGGT ATGATAACAATAAAGGGAGCAGGATTC R</td>
</tr>
<tr>
<td>5572</td>
<td><em>PhSYP51</em></td>
<td>ACAAGTTTGTACAAAAAAGCAGGCTTCTCACCACCAACCAT GAGCTTCTCACCACCACCATAGT</td>
</tr>
<tr>
<td>5573</td>
<td><em>PhSYP5</em></td>
<td>GAAAGCTGGGTAGT ACAATAAATGAAGGCTGGGTATGATAACAATAAAGGGAGCAGGATTC</td>
</tr>
<tr>
<td>7881</td>
<td><em>PhaTIP</em></td>
<td>GGGGACAAGTTTGTACAAAAAAGCAGGCTTCAAA TCAGGCTTCAAA</td>
</tr>
<tr>
<td>7883</td>
<td><em>PhaTIP</em></td>
<td>GGGGACCACCTTTGTACAAAAAAGCAGGCTTGAGTGATTACAAAGATGTTACAAAGGCTGGGTATGATAACAATAAAGGGAGCAGGATTC</td>
</tr>
<tr>
<td>7886</td>
<td><em>PhgTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>7887</td>
<td><em>PhgTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>7888</td>
<td><em>PhdTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAGGGAG R</td>
</tr>
<tr>
<td>8023</td>
<td><em>AtaTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAAGGGAG R</td>
</tr>
<tr>
<td>8024</td>
<td><em>AtaTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAAGGGAG R</td>
</tr>
<tr>
<td>8025</td>
<td><em>AtgTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAAGGGAG R</td>
</tr>
<tr>
<td>8026</td>
<td><em>AtgTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAAGGGAG R</td>
</tr>
<tr>
<td>8027</td>
<td><em>AddTIP</em></td>
<td>GGGGACACACTTTGTACAAAAAAGCAGGCTTGACTATGATA ACAATAAAGGGAG R</td>
</tr>
</tbody>
</table>

F indicates a forward orientation of the primer, relative to the orientation of the gene, and R reverse orientation.