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How Can Early Stress Influence Later Alzheimer's Disease Risk? Possible Mediators and Underlying Mechanisms

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ABSTRACT

Alzheimer's disease (AD) is a progressive, age-related neurodegenerative disorder to which genetic mutations and risk factors contribute. Evidence is increasing that environmental and lifestyle-related factors, such as exercise, nutrition, education, and exposure to (early-life) stress modify the onset, incidence, and progression of AD. Here, we discuss recent preclinical findings on putative substrates that can explain or contribute to the effects of stress early in life on the risk of developing AD. We focus in particular on stress hormones, neural networks, synapses, mitochondria, nutrient and lipid metabolism, adult neurogenesis, engram cell ensembles, and neuroinflammation. We discuss the idea that stress exposure early in life can alter these processes, either combined or in isolation, thereby reducing the capacity of the brain to resist deleterious consequences of, for example, amyloid- β accumulation, thereby accelerating cognitive decline and progression of Alzheimer-related changes in model systems of the disease. A better understanding of whether experiences early in life also modify trajectories of cognitive decline and pathology in AD and how the substrates discussed translate to humans may help develop novel preventive and/or therapeutic strategies to mitigate the consequences of stressors early in life and increase resilience to developing dementia.

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Alzheimer's disease (AD) is a neurodegenerative disorder characterized by progressive, age-related impairments in cognition that impacts many people worldwide. Next to classical hallmarks, such as amyloid- β (A β) plaques and neurofibrillary tangles, brain atrophy, angiopathy, astro-/microgliosis, neuronal hyperexcitability, and synapse loss characterize the AD brain. Mutations in the amyloid precursor protein (*APP*) and *PSEN1* and *PSEN2* genes, which cause familial AD, comprise only a small percentage of all dementia cases. Most patients suffer from sporadic AD, a multifactorial disorder, with *APOE* $\epsilon 4$ as the best-known genetic risk factor. Large AD genome-wide association studies (1,2) have identified additional genetic risk factors at 108 genomic loci, each with limited prevalence and penetrance, apart from the *APOE* locus (3,4). Expansion of these studies captures more rare variants with larger effects (5). Interestingly, genome-wide association study loci aggregate in specific disease pathways [related to A β plaque and neurofibrillary tangle formation, cholesterol metabolism, endocytosis/phagocytosis, and the innate immune system (3,6)].

Epigenetic mechanisms, such as DNA methylation, histone modifications, and noncoding RNAs, can exacerbate or rescue dysregulation of these pathways, and while results of recent studies were not always consistent, global increases in methylation/hydroxymethylation have been described in AD,

and hypomethylation of the *APP* gene has, e.g., been linked to increased *APP* expression (7,8). Also, epigenetic alterations in *APOE* or insulin signaling can cause insulin resistance and increase AD risk. Thus, epigenetic changes may alter susceptibility, but their exact role in AD requires more study.

The aberrant processing of the *APP* gene and/or the gradual accumulation of A β with age has been associated with neuronal spine loss, neurofibrillary tangles, vascular changes, and glial activation that together may underlie the behavioral and cognitive deficits in AD (9). Between patients with AD, there is substantial heterogeneity in disease onset, progression, and neuropathology, variation that is often discussed in the context of brain/cognitive reserve (10–15). This may be related to early-life factors (16–19) (see below) and helps conceptualize why some individuals are more prone to developing AD than others.

ENVIRONMENTAL FACTORS

Besides age and (epi)genetic factors, environmental factors, including stressful experiences, modulate dementia risk. In the Framingham Heart Study, temporal trends were described in the incidence of dementia over 3 decades, and a consistent reduction was found in the cumulative hazard rates for dementia among participants, while the incidence rate steadily declined during the second, third, and fourth epochs (20).

How Can Early Stress Influence Later Alzheimer's Disease Risk?

Because genetic changes were unlikely to be involved, these studies highlight lifestyle as a potential contributor to AD heterogeneity and risk. Nevertheless, which aspects of lifestyle, such as nutrition, exercise, smoking, stress, and education, are critical remains poorly understood (4,18,21). In individuals with higher educational attainment, IQ, or occupational attainment, a reduced risk for dementia has been found; reduced risk for dementia has also been found in those engaged in more social and leisure activities, bilingualism, and lifelong learning (although such conditions cannot be separated from differences in income and health care). Also, postmortem studies have substantiated links between a higher education level and/or participation in cognitively stimulating activities and a reduced risk of AD neuropathology (14,22–25).

Thus, whereas positive lifestyle factors may contribute to resilience to AD (26,27), adverse environmental experiences in contrast have been linked to earlier development of AD. Poor nutrition, obesity, smoking, sleep deprivation, and heavy alcohol use have been associated with increases in neuroinflammation, neurodegeneration, and a higher risk of developing AD (6,28). Also, stressful life events may lower the age of onset in patients with familial AD, while major depression, a disorder with a strong stress-related component, can increase sporadic AD risk (29,30). Furthermore, hypothalamic-pituitary-adrenal axis activity and glucocorticoid levels are commonly elevated already early in AD, highlighting an interaction between stress and AD risk (31,32). In addition, elderly individuals who are prone to psychological distress are more likely to develop AD than age-matched, non-stressed individuals (29–34). This is supported by preclinical studies in rodents that have shown that glucocorticoids may

enhance AD pathology (29,35–38), while targeting glucocorticoids can mitigate AD pathology (35,39).

AIM OF THIS REVIEW

Many aspects of adult brain function are determined early in life (40), and the early perinatal period represents a sensitive time window during which pronounced effects on later cognition and disease risk can be exerted (18,41). While the outcome of negative early-life experiences may depend on the match/mismatch with the context later in life (42–45), early adversity is commonly linked to deleterious consequences for brain function and higher risk for psychopathology (46). However, surprisingly little is known about the early-life period as risk factor for AD. Because early prevention of AD is critical, it is important to understand the substrates affected by stress early in life (early-life stress [ELS]) and how they contribute to AD risk.

Here, we discuss recent preclinical findings on putative mediators of ELS effects on later AD risk. We acknowledge that none of the existing AD mouse models encompasses the full complexity of human AD pathology. However, these models remain indispensable because they allow us to study how ELS modulates specific cellular and neuropathological features in the context of AD. Because some of them can be manipulated experimentally, they are of considerable value for future therapeutic/preventive approaches (see Figure 1).

ELS: CLINICAL STUDIES

The long time window between early-life and AD manifestation is challenging for (prospective) clinical studies. Consequently,

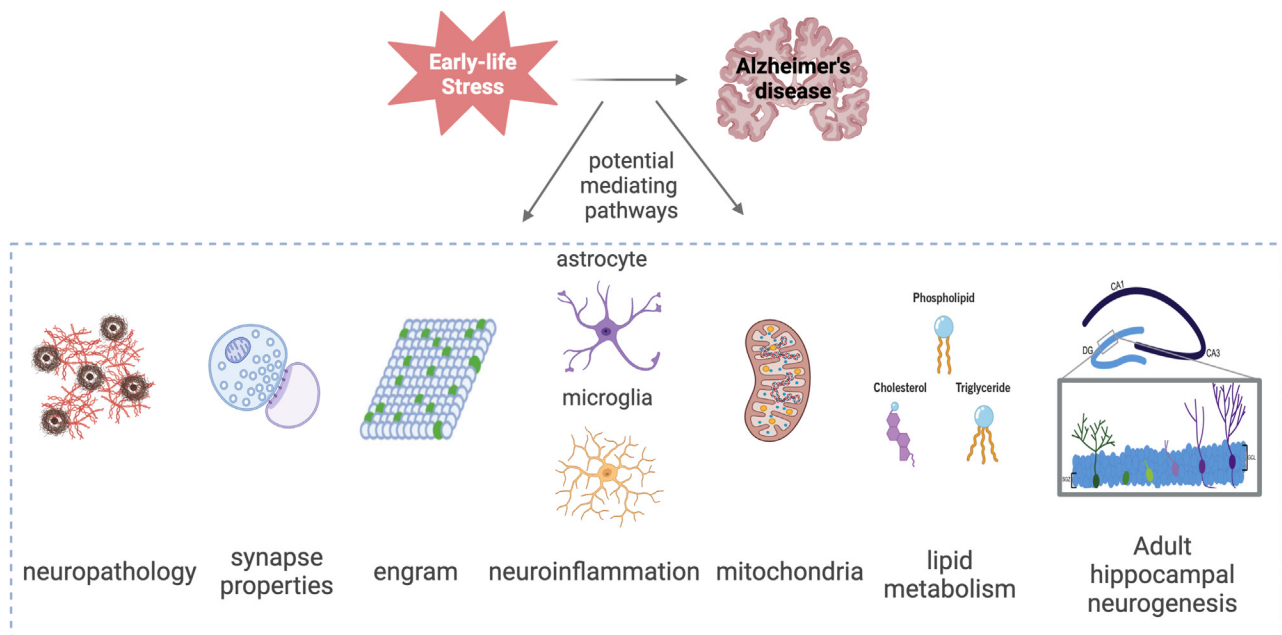


Figure 1. Summary of the different mechanisms through which early-life stress may alter development of later Alzheimer's pathology. Rodent studies provide evidence that early-life stress alters the progression and severity of Alzheimer's-related A β as well as tau pathology and cognitive dysfunction. At the cellular level, these changes may be mediated by alterations in synaptic function, stress responsivity, neural networks (and the excitation/inhibition balance within neural networks), neuroinflammation (including priming of microglia), mitochondrial function, lipid metabolism, and hippocampal neurogenesis. DG, dentate gyrus. (Figure created with BioRender.com.)

the clinical evidence that ELS affects AD risk is limited to retrospective studies on early exposure to trauma that was associated with compromised physical and mental health later in life (47–49). Studies on exposure, during early gestation, to the Dutch famine in 1944 to 1945 is associated with poorer cognitive function later in life. In particular, exposed men have shown evidence of advanced brain aging (16,50,51). Also, prenatal exposure to the Chinese famine has been associated with a higher prevalence of dementia (52). Adverse childhood events, such as losing a parent at a young age, increase dementia risk (48,53–55), and AD risk for people who experienced 3 or more adverse childhood events doubled (56). In addition, early social deprivation, sexual/physical abuse, and living in an orphanage have all been linked to a higher AD risk or impaired cognition as an adult (47,57–59). Recent systematic reviews have also linked the prenatal environment and childhood experiences to increased dementia risk (18,53,60).

ELS: PRECLINICAL STUDIES

While human studies underscore the relevance of focusing on early life, animal models provide mechanistic insight. Some of the most frequently found consequences of stress exposure during early life are accelerated development and altered stress responsivity (61–63). Notably, ELS is associated with accelerated maturation of brain regions involved in memory, from the synaptic to the network level (62,64–66). While such accelerated maturation may be beneficial in the short run, it may come at a cost for longevity (67), and increased stress sensitivity can modulate specific (epi) genetic programs (11) and/or render the brain more susceptible to AD-related pathology (18,39,54,68).

Cognitive decline is an important readout altered during the course of AD and is relatively easy to measure. In wild-type mice, exposure to ELS from postnatal days 2 to 9 induced prominent cognitive decline (62,69) but did not cause amyloid or tau pathology per se. The fact that no such accumulations occurred might have depended on genetic background, which was not taken into account here. When exposing AD transgenic mice to ELS, aggravated amyloid accumulation was observed in APP^{swE}/PS1^{dE9} (APP/PS1) mice (38,70,71), and cognitive flexibility and memory were impaired (18,39,68,72). Furthermore, hippocampal A β plaque load, soluble A β levels, and BACE1 levels were increased in the hippocampus of 6- and 12-month-old APP/PS1 mice exposed to ELS, while, surprisingly, the opposite was found after postnatal handling (71).

In 12-month-old APP/PS1 mice, the impaired cognitive flexibility could be rapidly rescued by a brief treatment with a glucocorticoid receptor antagonist, indicating an important role for stress hormones in these deficits (39). The same ELS paradigm further decreased hippocampal cell-associated (i.e., not plaque-related) A β levels in APP/PS1 mice at 4 months old, an age at which APP/PS1 mice start to show cognitive deficits (65,68,73); however, ELS exacerbated hippocampal A β plaque load at 10 months of age (68), demonstrating that ELS affects different forms of A β load in an age-dependent manner.

Behavioral characterization of young adult APP/PS1 mice exposed to ELS showed an impairment in reversal learning already at 3 months of age, particularly when the task criteria were increased (65). Also, hippocampal spatial learning was

impaired at 15 weeks of age (74). Although no plaques were observed at this age in these studies, some amyloid deposition has been reported by others in the neocortex at 6 weeks of age and at very low levels in the hippocampus at 3 months of age. Thus, relatively subtle cognitive impairments might have gone undetected with previously used behavioral paradigms and suggest that it is the preplaque A β deposition and/or soluble A β levels that may be involved in impaired cognitive flexibility. This may, to some extent, mimic the mild cognitive impairment that precedes the onset of full-blown AD. Thus, the early-life period modulates later cognition (69,75,76), with ELS enhancing pathological features characteristic of AD (18,25,38,39,53,59,77).

SYNAPSES

Next to these effects of ELS on A β and cognition in young APP/PS1 mice, ELS increased the synaptic strength of dentate gyrus granular cells and decreased the population of calretinin-expressing interneurons in the dentate gyrus at this age (65,66). This suggests that ELS could potentially change excitatory/inhibitory (E/I) balance in the hippocampus at an early age, thereby contributing to later cognitive decline. Parvalbumin-positive (PV⁺) interneurons are particularly sensitive to (neonatal) stress, and their maturation depends on synapse formation, axon myelination, and the acquisition of fast-spiking properties during the first postnatal weeks in mice (78).

In mouse models of AD, several studies have highlighted PV⁺ neuron dysfunction as a causal contributor to cognitive impairment. Hypoactivity of PV⁺ cells has been observed at relatively late disease stages in these mice, when A β plaques are present and optogenetic stimulation of PV⁺ cells, PV⁺ cell transplantation, or polysensory stimulation at 40 Hz (the typical firing frequency of PV⁺ cells) rescue cognitive deficits (78,79).

Interestingly, in 3- to 4-month-old APP/PS1 mice, i.e., before A β plaque deposition, hippocampal hyperexcitable PV⁺ cell states are observed that precede the hypoactivity at 6 months of age (74,80). Early chemogenetic inhibition of hippocampal PV⁺ cells also resulted in long-lasting restoration of E/I balance and cognitive performance and reduced the accumulation of A β plaques. Conversely, inducing an early hyperexcitable cell state of PV⁺ cells in wild-type mice increased sensitivity toward the neuronal network-disrupting effects of A β , and subthreshold concentrations of A β oligomers that are normally not toxic induced PV⁺ cell dysfunction, E/I imbalance, and cognitive impairment (74,80). Together, this suggests that early changes in hippocampal PV⁺ cell maturation, which is sensitive to ELS, could enhance AD pathology at an earlier age.

Hippocampal A β production is promoted by excitatory synaptic transmission. Hippocampal E/I balance disturbed by ELS can thereby accelerate A β accumulation and render APP/PS1 mice more vulnerable to later AD pathology, consistent with the disturbed network activity and symptoms of epilepsy frequently observed in several amyloid-based models (78,79,81–84) and in people developing AD (85,86). Long-term potentiation is suppressed by A β (87) and after ELS (70), which is mitigated by postweaning administration of the glutamate release inhibitor riluzole, consistent with the hypothesis that hyperexcitability may play a crucial role in ELS effects on later network disturbances relevant for AD.

Hippocampal synaptic dysfunction and spine loss are early features of AD neuropathology (88) and correlate well with cognitive decline. These changes are thought to be mediated by excess A β triggering hippocampal synaptic depression and loss in AD models (89,90). Notably, not all synapses appear equally vulnerable to A β . In the hippocampus, at the post-synaptic site, A β oligomers interact with NMDA receptors (87), particularly those that contain subunit GluN2B (91). Upon exposure to A β , mainly GluN2B-containing synapses are lost or weakened (92). GluN2B-containing NMDA receptors that bind scaffolding protein PSD-95 are thereby protected from synaptic depression (92). Although speculative, because ELS reduces PSD-95 and GluN2B content in synaptosomes (66), the resulting decreases in GluN2A/GluN2B ratio may thus render synapses more vulnerable to A β toxicity.

Also, AMPA receptor (AMPA) subunit composition determines whether synapses are sensitive to A β . AMPAR removal from synapses is a critical step through which A β oligomers mediate synaptic depression and synapse loss. A β oligomers weaken, or induce loss of, synapses from neurons that express the AMPAR subunit GluA3 (93). Whereas ELS affects AMPAR function (64), it initially decreases the GluA3 content at synapses, while increasing levels later in life (66). Consistent with this, increased GluA3 and GluN2B levels are associated with more severe cognitive dysfunction in people with mild cognitive impairment (94). Thus, ELS may render synapses even more vulnerable to synaptic A β toxicity by also altering their AMPAR subunit composition.

MITOCHONDRIA

Because synapses have high energy demands and are affected early in AD, there is growing interest in mitochondria and their oxidative phosphorylation capacity as possible substrates linking ELS to synaptic changes and AD pathogenesis (95–97). Genetic risk factors for AD, such as *APOE* ϵ 4, have been linked to mitochondrial function (98–101), and clear connections exist between mitochondria and stress also in the human brain (102). In wild-type mice, ELS has been shown to alter hippocampal expression of genes involved in mitochondrial fission, mitophagy, and antioxidant defenses (103). Also, in young APP/PS1 mice, alterations in oxidative phosphorylation, electron transport, and mitochondrial mass occur, and recent proteomic studies of the synaptosome of AD mice have revealed a clear resemblance in protein expression profiles to those of AD samples. Furthermore, specific alterations were found in APP/PS1 mice after ELS at early (preplaque) or later stages of A β pathology (104).

ELS in wild-type mice also triggers a similar synaptosomal profile as the one that is induced by early A β pathology in nonstressed APP/PS1 mice at 4 months of age. At a later (10 months of) age, ELS strongly affected the synaptic proteome of APP/PS1 mice, particularly the expression of astrocytic and mitochondrial proteins (while having minimal effects in wild-type mice). Thus, ELS and amyloidosis may share pathogenic pathways that involve synaptic mitochondrial dysfunction (104), and mitochondria may represent a shared substrate through which both ELS and A β can act to disturb energy homeostasis, synaptic physiology, and cognition.

NEUROINFLAMMATORY CHANGES AND MICROGLIA

Neuroinflammation and astro- and microglia activation are hallmarks of AD (6,28,105,106). Notably, both human and preclinical studies have shown that ELS impacts the immune system, and in animal models, ELS impacts the immune system long-term (41,68,107,108). In human cohorts, ELS has been shown to trigger low-grade systemic inflammation and impact inflammatory markers, including higher C-reactive protein and altered cytokine profiles (109–111). In addition, early-life adversity, so far comprising studies using a poor diet or stress during early life, has been demonstrated to prime microglia, the main immune cells of the brain (17,112), and to increase their response to amyloid in the hippocampus (68). Reemst *et al.* (112) further characterized the long-term impact of ELS on microglia in the hippocampus of adult males and found that ELS affected microglial morphology and altered gene expression of 186 genes, most notably increasing tumor necrosis factor α -responsive genes and genes implicated in cytoskeleton dynamics. Administration of an additional inflammatory challenge led to distinct changes in microglia, either isolated from the adult hippocampus of ELS mice or from control conditions. ELS reduced their capacity to phagocytose synaptosomes *ex vivo*, both early and later in life.

This revealed priming and persistent effects of ELS on microglia function and on the microglial response to a secondary immunological challenge. Activation- and age-related changes have also been described for hippocampal astroglia after ELS (107), suggesting that alterations in neuroimmune function may mediate ELS effects on AD risk.

In support of this idea, blocking microglia in their activity by minocycline treatment affected various cognitive and functional measures in AD mice when provided from an early age on, *i.e.*, before the onset of microgliosis (73). Soluble hippocampal A β levels or A β plaques deposition were not affected by minocycline treatment from 4 months of age, whereas these were reduced upon treatment at a later stage (6 months of age). A similar but different approach that selectively targets only plaque-associated microglia via systemic dendrimer administration is promising in this respect (113). Together, this highlights the need to intervene early with glial activation to reduce later A β pathology.

Most human risk genes for AD are highly, and several are selectively, expressed by microglia cells and astrocytes (2) that play key roles in neural circuit plasticity. Microglia can induce synapse loss and neuron loss in AD and contribute to impaired synaptic E/I balance, while astrocytes become reactive already early in AD and show impaired structural interactions with synapses (114) that may impair network activity. Remarkably, stimulation of hippocampal gamma oscillations, one of the indices of E/I balance, alters microglia activity, attenuates AD pathology, and improves cognition (115). Together, this positions microglia activation both upstream and downstream of E/I balance in AD pathology (see above).

LIPID METABOLISM

In addition to these physiological effects, increasing evidence highlights a possible role for lipid metabolism in neurodegeneration. Lipids are vital for brain structure, function, and metabolism, and the main lipids found in the brain

(phospholipids, polyunsaturated fatty acids [PUFAs], and cholesterol) show alterations in AD (116–119). Dysregulation of lipid metabolism has also been implicated in neuroinflammation and various aspects of AD (120), particularly regarding neuron-glia interactions (121). Moreover, polymorphisms of genes involved in lipid metabolism (like *APOE*) and in neuroinflammation (like *TREM2*) represent significant risk factors for AD, and the cognitive decline triggered by age-associated inflammation also involves specific lipids (122).

In particular, PUFAs from the n-6 and n-3 families and their derivatives, oxylipins, are important regulators of neuroinflammation (121). PUFAs are abundant in neurons and glial cells, including microglia, and decreased levels of n-3 PUFAs have been reported in the blood and brain of elderly participants with cognitive decline and AD (121,123). n-3 PUFA-derived specialized proresolving mediators, such as resolvins, maresins, and protectins, are crucial to resolve inflammation and enhance tissue restoration (121), and their levels are decreased in patients with mild cognitive impairment and AD (124) and in 5xFAD mice (125).

ELS alters various metabolic measures, including fat distribution and metabolic hormones like ghrelin and leptin, and changes brain lipid composition (76,112), which in the long-term can result in lipid dyshomeostasis. Interestingly, an early diet enriched in n-3 PUFAs could protect against ELS-induced cognitive deficits in wild-type mice, pointing toward opportunities for prevention (76). In fact, there is evidence that PUFA-based interventions can be effective in modulating AD-related pathologies in transgenic AD models (126–128) as well as in human trials (129). In addition, composite dietary interventions that include PUFA components have been promising (130,131). Additional investigation related to this exciting opportunity is needed to refine specific composition and optimal intervention times. Taken together, lipid dyshomeostasis may be a key biological substrate in mediating ELS-induced AD risk as well as a promising target for intervention.

NEUROGENESIS

The ELS-induced acceleration of brain development also affects structural plasticity, in particular neurogenesis, which is prominent during early brain development but persists into old age. Neurogenesis originates from stem cells that give rise to new neurons in the adult hippocampus and olfactory bulb. It is regulated by factors like exercise, inflammation, stress, and environment. Glucocorticoids are strong modulators (69,132–134) that, when they occur during early life, alter cortical and hippocampal neurogenesis and the response to stress for life (126,132,134,135). Exercise at later ages can in turn reduce stress, enhance learning, and promote neurogenesis, even in aged mice; however, no such rescue was reported after stress during early life (136,137).

Adult neurogenesis is involved in cognition and has been implicated in AD, based on both rodent and human studies (102,138). Reductions in neurogenesis occur notably already early in the disease process also in the human brain and in close correlation with cognitive status antemortem (102,138–140). The modulation of neurogenesis parallel to cognition by AD pathology as well as by ELS, exercise, and nutrients in various models (72,76,141) highlights a potential

role for neurogenesis in hippocampal cognition and AD etiology and offers promise to promote resilience to AD (26,102,134,137–139,142).

ENGRAMS

ELS hampers later synaptic function and learning and memory (18). Recent studies suggest that memories are encoded in the brain by sparsely distributed, learning-activated cells and that synaptic properties of these engram cell ensembles are relevant for memory (143). Engram activation can restore memory retrieval in mouse models of AD (144,145) indicating that, at least in the earlier stages, mouse models of AD display in particular deficits in memory retrieval. In adult animals, stress and stress hormones enlarge the engram, which in turn influences memory expression. In contrast, ELS reduces neuronal activation after learning and hampers retrieval (146). Thus, memories for which the encoding and consolidation were weaker as a consequence of ELS may be harder to retrieve over time, making them more susceptible to forgetting and thus particularly relevant for AD.

Furthermore, ELS-activated cells are involved in stress sensitivity later in life, resulting in greater reactivation of ELS-activated ensembles during adult stress (147). This may increase susceptibility to adult-life stressors, thereby increasing cumulative, lifelong stress exposure and accompanying vulnerability to AD pathophysiology. Such effects may be due at least in part to changes in the synaptic properties of engram cell ensembles (148), and similar processes could be involved in relation to ELS and AD pathology (144). Additionally, PV⁺ interneuron function and their critical ability to govern engram ensemble formation and memory processing are age dependent (149) and are notably impaired by ELS. Weakened PV⁺ activity following ELS may thus disrupt engram ensemble formation and/or accessibility and thereby impair memory processing.

CONCLUSIONS

Various (pre)clinical lines of evidence suggest that negative experiences early in life can modulate the trajectory of cognitive decline and the onset, incidence, and progression of (aspects of) AD. This emphasizes the importance of better understanding the mechanisms by which ELS can alter AD features.

The substrates discussed help us understand how such long-lasting effects are mediated and can possibly be prevented. They include alterations in E/I balance, synapse function, mitochondria, lipid metabolism, neurogenesis, and neuroinflammation and may render brain cells and networks less able to adapt to AD pathology, thereby decreasing resilience and advancing AD onset.

Preclinical models have helped identify several putative substrates, but human data on similar mechanisms remain scarce. Whereas the importance of the early-life period for psychopathologies, such as depression, is generally recognized, research should also focus on this critical period for the brain in relation to AD.

ELS effects on brain health and AD risk involve different substrates and pathways. Therefore, a multifaceted and/or lifestyle-based approach is also likely required to target the early phase of AD. Also, the optimal timing of such interventions needs to be determined. Because there is evidence that early diet-based

Box 1. Outstanding Questions

- What are the critical periods in perinatal (human) development that are most sensitive to environmental alterations and bear relevance to increasing either resilience or vulnerability to Alzheimer disease (AD)?
- How does early-life stress affect AD pathology and cognitive decline? This may involve (epi)genetic factors, altered stress responsivity, amyloid clearance, glymphatics, neurogenesis, mitochondria, and (micro)glia priming. A better understanding is needed of how early-life stress changes (pro- and anti-inflammatory) properties of glia (subtypes) and how this can affect clearance and pruning of (excitatory and inhibitory) synapses and network function.
- Does early-life stress influence inhibitory neuron function and network imbalances in AD? Can restoring alterations in inhibitory neurons rescue network function and cognitive deficits in patients with AD?
- How is neurogenesis linked to AD, and can its stimulation, or its programming by early-life factors, modulate cognitive decline in AD?
- What is the link between lipid metabolism, neuroinflammation, and later cognition? Early nutritional interventions, focusing on, e.g., polyunsaturated fatty acids and specialized proresolving mediators, could be tested in relation to later AD features.
- Which of the substrates discussed exerts the largest contribution and to what extent are they interconnected? Also important is which positive lifestyle factors during early life can stimulate cognitive/brain reserve, mitigate cognitive decline, delay disease onset, and ultimately promote resilience to AD.

interventions can counteract later cognitive decline after ELS (94,111,112,123,125,128,129,142,150), future studies should assess whether later life dietary interventions are effective as well.

Next to identifying and targeting mechanisms that increase AD risk and/or reduce its age of onset (as summarized in Figure 1), more research should be devoted to identifying protective factors that can promote resilience to AD (12,14,18,22,26,42), preferably at the earliest age possible (see Box 1).

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ARTICLE INFORMATION

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REFERENCES

1. Bellenguez C, Küçükali F, Jansen IE, Klei L, Moreno-Grau S, Amin N, *et al.* (2022): New insights into the genetic etiology of Alzheimer's disease and related dementias. *Nat Genet* 54:412–436.
2. Wightman DP, Jansen IE, Savage JE, Shadrin AA, Bahrami S, Holland D, *et al.* (2021): A genome-wide association study with 1,126,563 individuals identifies new risk loci for Alzheimer's disease. *Nat Genet* 53:1276–1282.
3. Andrews SJ, Renton AE, Fulton-Howard B, Podlesny-Drabiniok A, Marcora E, Goate AM (2023): The complex genetic architecture of Alzheimer's disease: Novel insights and future directions. *EBioMedicine* 90:104511.
4. Scheltens P, De Strooper B, Kivipelto M, Holstege H, Chételat G, Teunissen CE, *et al.* (2021): Alzheimer's disease. *Lancet* 397:1577–1590.
5. Holstege H, Hulsman M, Charbonnier C, Grenier-Boley B, Quenez O, Grozeva D, *et al.* (2022): Exome sequencing identifies rare damaging variants in ATP8B4 and ABCA1 as risk factors for Alzheimer's disease. *Nat Genet* 54:1786–1794.
6. Chen X, Holtzman DM (2022): Emerging roles of innate and adaptive immunity in Alzheimer's disease. *Immunity* 55:2236–2254.
7. Coppieters N, Dieriks BV, Lill C, Faull RLM, Curtis MA, Dragunow M (2014): Global changes in DNA methylation and hydroxymethylation in Alzheimer's disease human brain. *Neurobiol Aging* 35:1334–1344.
8. Wei X, Zhang L, Zeng Y (2020): DNA methylation in Alzheimer's disease: In brain and peripheral blood. *Mech Ageing Dev* 191:111319.
9. Karran E, De Strooper B (2022): The amyloid hypothesis in Alzheimer disease: New insights from new therapeutics. *Nat Rev Drug Discov* 21:306–318.
10. Arenaza-Urquijo EM, Przybelski SA, Machulda MM, Knopman DS, Lowe VJ, Mielke MM, *et al.* (2020): Better stress coping associated with lower tau in amyloid-positive cognitively unimpaired older adults. *Neurology* 94:e1571–e1579.
11. Christensen H, Anstey KJ, Parslow RA, Maller J, Mackinnon A, Sachdev P (2007): The brain reserve hypothesis, brain atrophy and aging. *Gerontology* 53:82–95.
12. De Vries LE, Huitinga I, Kessels HW, Swaab DF, Verhaagen J (2024): The concept of resilience to Alzheimer's disease: Current definitions and cellular and molecular mechanisms. *Mol Neurodegener* 19:33.
13. Groot C, Van Loenhoud AC, Barkhof F, Van Berckel BNM, Koene T, Teunissen CC, *et al.* (2018): Differential effects of cognitive reserve and brain reserve on cognition in Alzheimer disease. *Neurology* 90:e149–e156.
14. Stern Y (2012): Cognitive reserve in ageing and Alzheimer's disease. *Lancet Neurol* 11:1006–1012.
15. Van den Bergh BRH, van den Heuvel MI, Lahti M, Braeken M, de Rooij SR, Entringer S, *et al.* (2020): Prenatal developmental origins of behavior and mental health: The influence of maternal stress in pregnancy. *Neurosci Biobehav Rev* 117:26–64.
16. De Rooij SR (2022): Are brain and cognitive reserve shaped by early life circumstances? *Front Neurosci* 16:825811.

17. Hoijmakers L, Lesuis SL, Krugers H, Lucassen PJ, Korosi A (2018): A preclinical perspective on the enhanced vulnerability to Alzheimer's disease after early-life stress. *Neurobiol Stress* 8:172–185.
18. Lesuis SL, Hoijmakers L, Korosi A, de Rooij SR, Swaab DF, Kessels HW, *et al.* (2018): Vulnerability and resilience to Alzheimer's disease: Early life conditions modulate neuropathology and determine cognitive reserve. *Alzheimers Res Ther* 10:95.
19. Schuurmans IK, Hoepel SJW, Cecil CAM, Hillegers MHJ, Ikram MA, Luik AI (2023): The association of life stress with subsequent brain and cognitive reserve in middle-aged women. *J Alzheimers Dis* 93:97–106.
20. Satizabal C, Beiser AS, Seshadri S (2016): Incidence of dementia over three decades in the Framingham heart study. *N Engl J Med* 375:93–94.
21. Trumpff C, Monzel AS, Sandi C, Menon V, Klein HU, Fujita M, *et al.* (2024): Psychosocial experiences are associated with human brain mitochondrial biology. *Proc Natl Acad Sci U S A* 121:e2317673121.
22. Latimer CS, Keene CD, Flanagan ME, Hemmy LS, Lim KO, White LR, *et al.* (2017): Resistance to Alzheimer disease neuropathologic changes and apparent cognitive resilience in the Nun and Honolulu-Asia aging studies. *J Neuropathol Exp Neurol* 76:458–466.
23. Riley KP, Snowdon DA, Desrosiers MF, Markesbery WR (2005): Early life linguistic ability, late life cognitive function, and neuropathology: Findings from the Nun Study. *Neurobiol Aging* 26:341–347.
24. Snowdon DA, Nun Study (2003): Healthy aging and dementia: Findings from the Nun Study. *Ann Intern Med* 139:450–454.
25. Swaab DF (1991): Brain aging and Alzheimer's disease, "wear and tear" versus "use it or lose it". *Neurobiol Aging* 12:317–324.
26. Nestler EJ, Russo SJ (2024): Neurobiological basis of stress resilience. *Neuron* 112:1911–1929.
27. Ornish D, Madison C, Kivipetto M, Kemp C, McCulloch CE, Galasko D, *et al.* (2024): Effects of intensive lifestyle changes on the progression of mild cognitive impairment or early dementia due to Alzheimer's disease: A randomized, controlled clinical trial. *Alzheimers Res Ther* 16:122.
28. Singhaarachchi PH, Antal P, Calon F, Culmsee C, Delpech JC, Feldotto M, *et al.* (2024): Aging, sex, metabolic and life experience factors: Contributions to neuro-inflammation in Alzheimer's disease research. *Neurosci Biobehav Rev* 162:105724.
29. Burke MR, Sotiropoulos I, Waites CL (2024): The multiple roles of chronic stress and glucocorticoids in Alzheimer's disease pathogenesis. *Trends Neurosci* 47:933–948.
30. Chan YE, Chen MH, Tsai SJ, Bai YM, Tsai CF, Cheng CM, *et al.* (2020): Treatment-Resistant depression enhances risks of dementia and Alzheimer's disease: A nationwide longitudinal study. *J Affect Disord* 274:806–812.
31. Zheng B, Tai R, Yang Z, Middleton L, Udeh-Momoh C (2020): Cortisol hypersecretion and the risk of Alzheimer's disease: A systematic review and meta-analysis. *Ageing Res Rev* 64:101171.
32. Ouanes S, Rabl M, Clark C, Kirschbaum C, Popp J (2022): Persisting neuropsychiatric symptoms, Alzheimer's disease, and cerebrospinal fluid cortisol and dehydroepiandrosterone sulfate. *Alzheimers Res Ther* 14:190.
33. Escher CM, Sannemann L, Jessen F (2019): Stress and Alzheimer's disease. *J Neural Transm (Vienna)* 126:1155–1161.
34. Mejia S, Giraldo M, Pineda D, Ardila A, Lopera F (2003): Nongenetic factors as modifiers of the age of onset of familial Alzheimer's disease. *Int Psychogeriatr* 15:337–349.
35. Canet G, Pineau F, Zussy C, Hernandez C, Hunt H, Chevallier N, *et al.* (2020): Glucocorticoid receptors signaling impairment potentiates amyloid-beta oligomers-induced pathology in an acute model of Alzheimer's disease. *FASEB J* 34:1150–1168.
36. Catania C, Sotiropoulos I, Silva R, Onofri C, Breen KC, Sousa N, Almeida OF (2009): The amyloidogenic potential and behavioral correlates of stress. *Mol Psychiatry* 14:95–105.
37. Green KN, Billings LM, Roozendaal B, McGaugh JL, LaFerla FM (2006): Glucocorticoids increase amyloid- β and tau pathology in a mouse model of Alzheimer's disease. *J Neurosci* 26:9047–9056.
38. Lesuis SL, Maurin H, Borghgraef P, Lucassen PJ, Van Leuven F, Krugers HJ (2016): Positive and negative early life experiences differentially modulate long term survival and amyloid protein levels in a mouse model of Alzheimer's disease. *Oncotarget* 7:39118–39135.
39. Lesuis SL, Weggen S, Baches S, Lucassen PJ, Krugers HJ (2018): Targeting glucocorticoid receptors prevents the effects of early life stress on amyloid pathology and cognitive performance in APP/PS1 mice. *Transl Psychiatry* 8:53.
40. Turini S, Wong B, Eldaief M, Press DZ, Sinclair DA, Koch G, *et al.* (2023): The multifactorial nature of healthy brain ageing: Brain changes, functional decline and protective factors. *Ageing Res Rev* 88:101939.
41. Nusslock R, Miller GE (2016): Early-life adversity and physical and emotional health across the lifespan: A neuroimmune network hypothesis. *Biol Psychiatry* 80:23–32.
42. Santarelli S, Zimmermann C, Kalideris G, Lesuis SL, Arloth J, Uribe A, *et al.* (2017): An adverse early life environment can enhance stress resilience in adulthood. *Psychoneuroendocrinology* 78:213–221.
43. Champagne DL, Bagot RC, van Hasselt F, Ramakers G, Meaney MJ, de Kloet ER, *et al.* (2008): Maternal care and hippocampal plasticity: Evidence for experience-dependent structural plasticity, altered synaptic functioning, and differential responsiveness to glucocorticoids and stress. *J Neurosci* 28:6037–6045.
44. Daskalakis NP, Oitzl MS, Schächinger H, Champagne DL, de Kloet ER (2012): Testing the cumulative stress and mismatch hypotheses of psychopathology in a rat model of early-life adversity. *Physiol Behav* 106:707–721.
45. Oomen CA, Soeters H, Audureau N, Vermunt L, van Hasselt FN, Manders EMM, *et al.* (2010): Severe early life stress hampers spatial learning and neurogenesis, but improves hippocampal synaptic plasticity and emotional learning under high-stress conditions in adulthood. *J Neurosci* 30:6635–6645.
46. Yu Z, Cao Y, Shang T, Li P (2024): Depression in youths with early life adversity: A systematic review and meta-analysis. *Front Psychiatry* 15:1378807.
47. Chugani HT, Behen ME, Muzik O, Juhász C, Nagy F, Chugani DC (2001): Local brain functional activity following early deprivation: A study of postinstitutionalized Romanian orphans. *Neuroimage* 14:1290–1301.
48. Conde-Sala JL, Garre-Olmo J (2020): Early parental death and psychosocial risk factors for dementia: A case-control study in Europe. *Int J Geriatr Psychiatry* 35:1051–1059.
49. Mueller SC, Maheu FS, Dozier M, Peloso E, Mandell D, Leibenluft E, *et al.* (2010): Early-life stress is associated with impairment in cognitive control in adolescence: An fMRI study. *Neuropsychologia* 48:3037–3044.
50. Boots A, Thomason ME, Espinoza-Heredia C, Pruitt PJ, Damoiseaux JS, Roseboom TJ, de Rooij SR (2022): Sex-specific effects of prenatal undernutrition on resting-state functional connectivity in the human brain at age 68. *Neurobiol Aging* 112:129–138.
51. De Rooij SR, Bleker LS, Painter RC, Ravelli AC, Roseboom TJ (2022): Lessons learned from 25 years of Research into long term Consequences of prenatal Exposure to the Dutch famine 1944–45: The Dutch famine Birth Cohort. *Int J Environ Health Res* 32:1432–1446.
52. Kang Y, Zhang Y, Feng Z, Liu M, Li Y, Yang H, *et al.* (2017): Nutritional deficiency in early life facilitates aging-associated cognitive decline. *Curr Alzheimer Res* 14:841–849.
53. Corney KB, West EC, Quirk SE, Pasco JA, Stuart AL, Manavi BA, *et al.* (2022): The relationship between adverse childhood experiences and Alzheimer's disease: A systematic review. *Front Aging Neurosci* 14:831378.
54. Norton MC, Smith KR, Østbye T, Tschanz JT, Schwartz S, Corcoran C, *et al.* (2011): Early parental death and remarriage of widowed parents as risk factors for Alzheimer disease: The Cache County study. *Am J Geriatr Psychiatry* 19:814–824.
55. Tani Y, Fujiwara T, Kondo K (2020): Association between adverse childhood experiences and dementia in older Japanese adults. *JAMA Netw Open* 3:e1920740.
56. Tanaka T, Hirai S, Hosokawa M, Saito T, Sakuma H, Saido T, *et al.* (2021): Early-life stress induces the development of Alzheimer's disease pathology via angiopathy. *Exp Neurol* 337:113552.

How Can Early Stress Influence Later Alzheimer's Disease Risk?

57. Huang Z, Jordan JD, Zhang Q (2023): Early life adversity as a risk factor for cognitive impairment and Alzheimer's disease. *Transl Neurodegener* 12:25.
58. Kaplan GA, Turrell G, Lynch JW, Everson SA, Helkala EL, Salonen JT (2001): Childhood socioeconomic position and cognitive function in adulthood. *Int J Epidemiol* 30:256–263.
59. Majoka MA, Schimming C (2021): Effect of social determinants of health on cognition and risk of Alzheimer disease and related dementias. *Clin Ther* 43:922–929.
60. Wieggersma AM, Boots A, Langendam MW, Limpens J, Shenkin SD, Korosi A, *et al.* (2023): Do prenatal factors shape the risk for dementia?: A systematic review of the epidemiological evidence for the prenatal origins of dementia [published online Apr 8]. *Soc Psychiatry Psychiatr Epidemiol*.
61. Chen Y, Baram TZ (2016): Toward understanding how early-life stress reprograms cognitive and emotional brain networks. *Neuropsychopharmacology* 41:197–206.
62. Walker CD, Bath KG, Joels M, Korosi A, Larauche M, Lucassen PJ, *et al.* (2017): Chronic early life stress induced by limited bedding and nesting (LBN) material in rodents: Critical considerations of methodology, outcomes and translational potential. *Stress* 20:421–448.
63. Chan SY, Ngoh ZM, Ong ZY, The AL, Kee MZL, Zhou JH, *et al.* (2024): The influence of early-life adversity on the coupling of structural and functional brain connectivity across childhood. *Nature. Ment Health* 2:52–62.
64. Al-Chami A, Ross A, Hayley S, Sun H (2020): Early life stress facilitates synapse premature unsilencing to enhance AMPA receptor function in the developing hippocampus. *J Neurophysiol* 124:815–821.
65. Brosens N, Samouil D, Stolker S, Katsika EV, Weggen S, Lucassen PJ, Krugers HJ (2023): Early life stress enhances cognitive decline and alters synapse function and interneuron numbers in young male APP/PS1 Mice. *J Alzheimers Dis* 96:1097–1113.
66. Brosens N, Simon C, Kessels HW, Lucassen PJ, Krugers HJ (2023): Early life stress lastingly alters the function and AMPA-receptor composition of glutamatergic synapses in the hippocampus of male mice. *J Neuroendocrinol* 35:e13346.
67. D'Amico D, Amestoy ME, Fiocco AJ (2022): The mediating role of allostatic load in the relationship between early life adversity and cognitive function across the adult lifespan. *Psychoneuroendocrinology* 141:105761.
68. Hoeijmakers L, Ruigrok SR, Amelanchik A, Ivan D, van Dam A-M, Lucassen PJ, Korosi A (2017): Early-life stress lastingly alters the neuroinflammatory response to amyloid pathology in an Alzheimer's disease mouse model. *Brain Behav Immun* 63:160–175.
69. Naninck EFG, Hoeijmakers L, Kakava-Georgiadou N, Meesters A, Lazic SE, Lucassen PJ, Korosi A (2015): Chronic early life stress alters developmental and adult neurogenesis and impairs cognitive function in mice. *Hippocampus* 25:309–328.
70. Lesuis SL, Kaplick PM, Lucassen PJ, Krugers HJ (2019): Treatment with the glutamate modulator riluzole prevents early life stress-induced cognitive deficits and impairments in synaptic plasticity in APP^{swe}/PS1^{dE9} mice. *Neuropharmacology* 150:175–183.
71. Lesuis SL, van Hoek BACE, Lucassen PJ, Krugers HJ (2017): Early postnatal handling reduces hippocampal amyloid plaque formation and enhances cognitive performance in APP^{swe}/PS1^{dE9} mice at middle age. *Neurobiol Learn Mem* 144:27–35.
72. Hui J, Feng G, Zheng C, Jin H, Jia N (2017): Maternal separation exacerbates Alzheimer's disease-like behavioral and pathological changes in adult APP^{swe}/PS1^{dE9} mice. *Behav Brain Res* 318:18–23.
73. Kater MSJ, Huffels CFM, Oshima T, Renckens NS, Middeldorp J, Boddeke EWGM, *et al.* (2023): Prevention of microgliosis halts early memory loss in a mouse model of Alzheimer's disease. *Brain Behav Immun* 107:225–241.
74. Hijazi S, Heistek TS, Scheltens P, Neumann U, Shimshek DR, Mansvelder HD, *et al.* (2020): Early restoration of parvalbumin interneuron activity prevents memory loss and network hyperexcitability in a mouse model of Alzheimer's disease. *Mol Psychiatry* 25:3380–3398.
75. Pillai AG, Arp M, Velzing E, Lesuis SL, Schmidt MV, Holsboer F, *et al.* (2018): Early life stress determines the effects of glucocorticoids and stress on hippocampal function: Electrophysiological and behavioral evidence respectively. *Neuropharmacology* 133:307–318.
76. Yam KY, Schipper L, Reemst K, Ruigrok SR, Abbink MR, Hoeijmakers L, *et al.* (2019): Increasing availability of ω -3 fatty acid in the early-life diet prevents the early-life stress-induced cognitive impairments without affecting metabolic alterations. *FASEB J* 33:5729–5740.
77. Donley GAR, Lönnroos E, Tuomainen T-P, Kauhanen J (2018): Association of childhood stress with late-life dementia and Alzheimer's disease: The KIHd study. *Eur J Public Health* 28:1069–1073.
78. Hijazi S, Smit AB, van Kesteren RE (2023): Fast-spiking parvalbumin-positive interneurons in brain physiology and Alzheimer's disease. *Mol Psychiatry* 28:4954–4967.
79. Vico Varela EV, Etter G, Williams S (2019): Excitatory-inhibitory imbalance in Alzheimer's disease and therapeutic significance. *Neurobiol Dis* 127:605–615.
80. Hijazi S, Heistek TS, van der Loo R, Mansvelder HD, Smit AB, van Kesteren RE (2020): Hyperexcitable parvalbumin interneurons render hippocampal circuitry vulnerable to amyloid beta. *iScience* 23:101271.
81. Chung H, Park K, Jang HJ, Kohl MM, Kwag J (2020): Dissociation of somatostatin and parvalbumin interneurons circuit dysfunctions underlying hippocampal theta and gamma oscillations impaired by amyloid β oligomers in vivo. *Brain Struct Funct* 225:935–954.
82. Petrache AL, Rajulawalla A, Shi A, Wetzel A, Saito T, Saido TC, *et al.* (2019): Aberrant excitatory-inhibitory synaptic mechanisms in entorhinal cortex microcircuits during the pathogenesis of Alzheimer's disease. *Cereb Cortex* 29:1834–1850.
83. Schmid LC, Mittag M, Poll S, Steffen J, Wagner J, Geis H-R, *et al.* (2016): Dysfunction of somatostatin-positive interneurons associated with memory deficits in an Alzheimer's disease model. *Neuron* 92:114–125.
84. Van Loenhoud AC, Van Der Flier WM, Wink AM, Dicks E, Groot C, Twisk J, *et al.* (2019): Cognitive reserve and clinical progression in Alzheimer disease: A paradoxical relationship. *Neurology* 93:e334–e346.
85. Ranasinghe KG, Petersen C, Kudo K, Mizuiri D, Rankin KP, Rabinovici GD, *et al.* (2021): Reduced synchrony in alpha oscillations during life predicts post mortem neurofibrillary tangle density in early-onset and atypical Alzheimer's disease. *Alzheimers Dement* 17:2009–2019.
86. Vossel KA, Ranasinghe KG, Beagle AJ, Mizuiri D, Honma SM, Dowling AF, *et al.* (2016): Incidence and impact of subclinical epileptiform activity in Alzheimer's disease. *Ann Neurol* 80:858–870.
87. Kessels HW, Nabavi S, Malinow R (2013): Metabotropic NMDA receptor function is required for β -amyloid-induced synaptic depression. *Proc Natl Acad Sci U S A* 110:4033–4038.
88. Bilousova T, Melnik M, Miyoshi E, Gonzalez BL, Poon WW, Vinters HV, *et al.* (2019): Apolipoprotein E/amyloid-beta complex accumulates in Alzheimer disease cortical synapses via apolipoprotein E receptors and is enhanced by APOE4. *Am J Pathol* 189:1621–1636.
89. Kamenetz F, Tomita T, Hsieh H, Seabrook G, Borchelt D, Iwatsubo T, *et al.* (2003): APP processing and synaptic function. *Neuron* 37:925–937.
90. Wei W, Nguyen LN, Kessels HW, Hagiwara H, Sisodia S, Malinow R (2010): Amyloid beta from axons and dendrites reduces local spine number and plasticity. *Nat Neurosci* 13:190–196.
91. Taniguchi K, Yamamoto F, Amano A, Tamaoka A, Sanjo N, Yokota T, *et al.* (2022): Amyloid-beta oligomers interact with NMDA receptors containing GluN2B subunits and metabotropic glutamate receptor 1 in primary cortical neurons: Relevance to the synapse pathology of Alzheimer's disease. *Neurosci Res* 180:90–98.
92. Dore K, Carrico Z, Alfonso S, Marino M, Koymans K, Kessels HW, Malinow R (2021): PSD-95 protects synapses from β -amyloid. *Cell Rep* 35:109194.
93. Reinders NR, Pao Y, Renner MC, da Silva-Matos CM, Lodder TR, Malinow R, Kessels HW (2016): Amyloid- β effects on synapses and memory require AMPA receptor subunit GluA3. *Proc Natl Acad Sci U S A* 113:E6526–E6534.

94. Berchtold NC, Sabbagh MN, Beach TG, Kim RC, Cribbs DH, Cotman CW (2014): Brain gene expression patterns differentiate mild cognitive impairment from normal aged and Alzheimer's disease. *Neurobiol Aging* 35:1961–1972.
95. Ashleigh T, Swerdlow RH, Beal MF (2023): The role of mitochondrial dysfunction in Alzheimer's disease pathogenesis. *Alzheimers Dement* 19:333–342.
96. Clement A, Madsen MJ, Kastaniegaard K, Wiborg O, Asuni AA, Stensballe A (2022): Chronic stress induces hippocampal mitochondrial damage in APP/PS1 Model Mice and Wildtype Littermates. *J Alzheimers Dis* 87:1–14.
97. Wang W, Zhao F, Lu Y, Siedlak SL, Fujioka H, Feng H, *et al.* (2023): Damaged mitochondria coincide with presynaptic vesicle loss and abnormalities in Alzheimer's disease brain. *Acta Neuropathol Commun* 11:54.
98. Ning Z, Liu Y, Wan M, Zuo Y, Chen S, Shi Z, *et al.* (2024): APOE2 protects against A β pathology by improving neuronal mitochondrial function through ERR α signaling. *Cell Mol Biol Lett* 29:87.
99. Li W, Ali T, He K, Zheng C, Li N, Yu ZJ, Li S (2024): ApoE4 dysregulation incites depressive symptoms and mitochondrial impairments in mice. *J Cell Mol Med* 28:e18160.
100. Lee H, Cho S, Kim MJ, Park YJ, Cho E, Jo YS, *et al.* (2023): ApoE4-dependent lysosomal cholesterol accumulation impairs mitochondrial homeostasis and oxidative phosphorylation in human astrocytes. *Cell Rep* 42:113183.
101. Mahley RW (2023): Apolipoprotein E4 targets mitochondria and the mitochondria-associated membrane complex in neuropathology, including Alzheimer's disease. *Curr Opin Neurobiol* 79:102684.
102. Tobin MK, Musaraca K, Disouky A, Shetti A, Bheri A, Honer WG, *et al.* (2019): Human hippocampal neurogenesis persists in aged adults and Alzheimer's disease patients. *Cell Stem Cell* 24:974–982.e3.
103. Ruigrok SR, Yim K, Emmerzaal TL, Geenen B, Stöberl N, den Blaauwen JL, *et al.* (2021): Effects of early-life stress on peripheral and central mitochondria in male mice across ages. *Psychoneuroendocrinology* 132:105346.
104. Kotah JM, Kater MSJ, Brosens N, Lesuis SL, Tandari R, Blok TM, *et al.* (2024): Early-life stress and amyloidosis in mice share pathogenic pathways involving synaptic mitochondria and lipid metabolism. *Alzheimers Dement* 20:1637–1655.
105. Hoeijmakers L, Heinen Y, van Dam A-M, Lucassen PJ, Korosi A (2016): Microglial priming and Alzheimer's disease: A possible role for (early) immune challenges and epigenetics? *Front Hum Neurosci* 10:398.
106. Patani R, Hardingham GE, Liddelow SA (2023): Functional roles of reactive astrocytes in neuroinflammation and neurodegeneration. *Nat Rev Neurol* 19:395–409.
107. Abbink MR, Kotah JM, Hoeijmakers L, Mak A, Yvon-Durocher G, van der Gaag B, *et al.* (2020): Characterization of astrocytes throughout life in wildtype and APP/PS1 mice after early-life stress exposure. *J Neuroinflammation* 17:91.
108. Solarz A, Majcher-Maślanka I, Kryst J, Chocyk A (2023): Early-life stress affects peripheral, blood-brain barrier, and brain responses to immune challenge in juvenile and adult rats. *Brain Behav Immun* 108:1–15.
109. Baumeister D, Akhtar R, Ciufolini S, Pariante CM, Mondelli V (2016): Childhood trauma and adulthood inflammation: A meta-analysis of peripheral C-reactive protein, interleukin-6 and tumour necrosis factor- α . *Mol Psychiatry* 21:642–649.
110. Willemen FEM, van Zuiden M, Zantvoord JB, de Rooij SR, van den Born BH, Hak AE, *et al.* (2022): Associations between child maltreatment, inflammation, and comorbid metabolic syndrome to depressed mood in a multiethnic urban population: The HELIUS study. *Front Psychol* 13:787029.
111. Reemst K, Broos JY, Abbink MR, Cimetti C, Giera M, Kooij G, Korosi A (2022): Early-life stress and dietary fatty acids impact the brain lipid/oxylipin profile into adulthood, basally and in response to LPS. *Front Immunol* 13:967437.
112. Reemst K, Kracht L, Kotah JM, Rahimian R, van Irsen AAS, Congrains Sotomayor G, *et al.* (2022): Early-life stress lastingly impacts microglial transcriptome and function under basal and immune-challenged conditions. *Transl Psychiatry* 12:507.
113. Henningfield CM, Soni N, Lee RW, Sharma R, Cleland JL, Green KN (2024): Selective targeting and modulation of plaque associated microglia via systemic hydroxyl dendrimer administration in an Alzheimer's disease mouse model. *Alzheimers Res Ther* 16:101.
114. Kater MSJ, Badia-Soteras A, van Weering JRT, Smit AB, Verheijen MHG (2023): Electron microscopy analysis of astrocyte-synapse interactions shows altered dynamics in an Alzheimer's disease mouse model. *Front Cell Neurosci* 17:1085690.
115. Martorell AJ, Paulson AL, Suk H-J, Abdurrob F, Drummond GT, Guan W, *et al.* (2019): Multi-sensory gamma stimulation ameliorates alzheimer's-associated pathology and improves cognition. *Cell* 177:256–271.e22.
116. Feringa FM, van der Kant R (2021): Cholesterol and Alzheimer's disease; from risk genes to pathological effects. *Front Aging Neurosci* 13:690372.
117. Husain MA, Laurent B, Plourde M (2021): APOE and Alzheimer's disease: From lipid transport to physiopathology and therapeutics. *Front Neurosci* 15:630502.
118. Saher G (2023): Cholesterol metabolism in aging and age-related disorders. *Annu Rev Neurosci* 46:59–78.
119. Cisbani G, Bazinet RP (2021): The role of peripheral fatty acids as biomarkers for Alzheimer's disease and brain inflammation. *Prostaglandins Leukot Essent Fatty Acids* 164:102205.
120. Hansen SB, Wang H (2023): The shared role of cholesterol in neuronal and peripheral inflammation. *Pharmacol Ther* 249:108486.
121. Layé S, Nadjar A, Joffre C, Bazinet RP (2018): Anti-inflammatory effects of omega-3 fatty acids in the brain: Physiological mechanisms and relevance to pharmacology. *Pharmacol Rev* 70:12–38.
122. Minhas PS, Latif-Hernandez A, McReynolds MR, Durairaj AS, Wang Q, Rubin A, *et al.* (2021): Restoring metabolism of myeloid cells reverses cognitive decline in ageing. *Nature* 590:122–128.
123. Ooi KM, Vacy K, Boon WC (2021): Fatty acids and beyond: Age and Alzheimer's disease related changes in lipids reveal the neuro-nutritional potential of lipids in cognition. *Neurochem Int* 149:105143.
124. Do KV, Hjorth E, Wang Y, Jun B, Kautzmann MI, Ohshima M, *et al.* (2023): Cerebrospinal fluid profile of lipid mediators in Alzheimer's disease. *Cell Mol Neurobiol* 43:797–811.
125. Kantarci A, Aytan N, Palaska I, Stephens D, Crabtree L, Benincasa C, *et al.* (2018): Combined administration of resolvin E1 and lipoxin A4 resolves inflammation in a murine model of Alzheimer's disease. *Exp Neurol* 300:111–120.
126. Krontira AC, Cruceanu C, Dony L, Kyrousi C, Link MH, Rek N, *et al.* (2024): Human cortical neurogenesis is altered via glucocorticoid-mediated regulation of ZBTB16 expression. *Neuron* 112:1426–1443.e11.
127. Galeano P, de Ceglia M, Mastrogiovanni M, Campanelli L, Medina-Vera D, Campolo N, *et al.* (2023): The effect of fat intake with increased Omega-6-to-Omega-3 polyunsaturated fatty acid ratio in animal models of early and late Alzheimer's disease-like pathogenesis. *Int J Mol Sci* 24:17009.
128. Fang G, Shi B, Wu K, Chen S, Gao X, Xiao S, *et al.* (2019): The protective role of endogenous n-3 polyunsaturated fatty acids in Tau Alzheimer's disease mouse model. *Int J Neurosci* 129:325–336.
129. Nozaki S, Sawada N, Matsuoka YJ, Shikimoto R, Mimura M, Tsugane S (2021): Association between dietary fish and PUFA intake in midlife and dementia in later life: The JPHC Saku mental health study. *J Alzheimers Dis* 79:1091–1104.
130. Levak N, Lehtisalo J, Thunborg C, Westman E, Andersen P, Andrieu S, *et al.* (2024): Nutrition guidance within a multimodal intervention improves diet quality in prodromal Alzheimer's disease: Multimodal Preventive Trial for Alzheimer's disease (MIND-Admini). *Alzheimers Res Ther* 16:147.
131. Dai L, Lin X, Wang S, Gao Y, He F (2024): The Mediterranean-dietary approaches to stop hypertension diet intervention for neurodegenerative delay (MIND) diet: A bibliometric analysis. *Front Nutr* 11:1348808.

How Can Early Stress Influence Later Alzheimer's Disease Risk?

132. Mirescu C, Peters JD, Gould E (2004): Early life experience alters response of adult neurogenesis to stress. *Nat Neurosci* 7:841–846.
133. Bassil K, Krontira AC, Leroy T, Escoto AIH, Snijders C, Pernia CD, *et al.* (2023): In vitro modeling of the neurobiological effects of glucocorticoids: A review. *Neurobiol Stress* 23:100530.
134. Lucassen PJ, Oomen CA, Naninck EFG, Fitzsimons CP, van Dam AM, Czeh B, Korosi A (2015): Regulation of adult neurogenesis and plasticity by (early) stress, glucocorticoids, and inflammation. *Cold Spring Harb Perspect Biol* 7:a021303.
135. Ruiz R, Roque A, Pineda E, Licona-Limón P, José Valdéz-Alarcón J, Lajud N (2018): Early life stress accelerates age-induced effects on neurogenesis, depression, and metabolic risk. *Psychoneuroendocrinology* 96:203–211.
136. Abbink MR, Naninck EFG, Lucassen PJ, Korosi A (2017): Early-life stress diminishes the increase in neurogenesis after exercise in adult female mice. *Hippocampus* 27:839–844.
137. Kannangara TS, Lucero MJ, Gil-Mohapel J, Drapala RJ, Simpson JM, Christie BR, van Praag H (2011): Running reduces stress and enhances cell genesis in aged mice. *Neurobiol Aging* 32:2279–2286.
138. Salta E, Lazarov O, Fitzsimons CP, Tanzi R, Lucassen PJ, Choi SH (2023): Adult hippocampal neurogenesis in Alzheimer's disease: A roadmap to clinical relevance. *Cell Stem Cell* 30:120–136.
139. Du Preez A, Lefèvre-Arbogast S, González-Domínguez R, Houghton V, de Lucia C, Lee H, *et al.* (2024): Association of dietary and nutritional factors with cognitive decline, dementia, and depressive symptomatology in older individuals according to a neurogenesis-centred biological susceptibility to brain ageing. *Age Ageing* 53:ii47–ii59.
140. Moreno-Jiménez EP, Flor-García M, Terreros-Roncal J, Rábano A, Cafini F, Pallas-Bazarra N, *et al.* (2019): Adult hippocampal neurogenesis is abundant in neurologically healthy subjects and drops sharply in patients with Alzheimer's disease. *Nat Med* 25:554–560.
141. Mishra R, Phan T, Kumar P, Morrissey Z, Gupta M, Hollands C, *et al.* (2022): Augmenting neurogenesis rescues memory impairments in Alzheimer's disease by restoring the memory-storing neurons. *J Exp Med* 219:e20220391.
142. Geertsema J, Kratochvil M, González-Domínguez R, Lefèvre-Arbogast S, Low DY, Du Preez A, *et al.* (2024): Coffee polyphenols ameliorate early-life stress-induced cognitive deficits in male mice. *Neurobiol Stress* 31:100641.
143. Rao-Ruiz P, Visser E, Mitrić M, Smit AB, van den Oever MC (2021): A synaptic framework for the persistence of memory engrams. *Front Synaptic Neurosci* 13:661476.
144. Lazarov O, Gupta M, Kumar P, Morrissey Z, Phan T (2024): Memory circuits in dementia: The engram, hippocampal neurogenesis and Alzheimer's disease. *Prog Neurobiol* 236:102601.
145. Perusini JN, Cajigas SA, Cohensedgh O, Lim SC, Pavlova IP, Donaldson ZR, Denny CA (2017): Optogenetic stimulation of dentate gyrus engrams restores memory in Alzheimer's disease mice. *Hippocampus* 27:1110–1122.
146. Sanguino-Gomez J, Huijgens S, den Hartog M, Schenk IJM, Kluck W, Versluis TD, Krugers HJ (2024): Neural correlates of learning and memory are altered by early-life stress. *Neurobiol Learn Mem* 19:107952.
147. Balouek JA, Mclain CA, Minerva AR, Rashford RL, Bennett SN, Rogers FD, Peña CJ (2023): Reactivation of early-life stress-sensitive neuronal ensembles contributes to lifelong stress hypersensitivity. *J Neurosci* 43:5996–6009.
148. Brosens N, Lesuis SL, Rao-Ruiz P, van den Oever MC, Krugers HJ (2024): Shaping memories via stress: A synaptic engram perspective. *Biol Psychiatry* 95:721–731.
149. Ramsaran AI, Wang Y, Golbabaei A, Aleshin S, de Snoo ML, Yeung BA, *et al.* (2023): A shift in the mechanisms controlling hippocampal engram formation during brain maturation. *Science* 380:543–551.
150. Naninck EFG, Oosterink JE, Yam KY, de Vries LP, Schierbeek H, van Goudoever JB, *et al.* (2017): Early micronutrient supplementation protects against early stress-induced cognitive impairments. *FASEB J* 31:505–518.