Spatio-temporal dynamics of sulfur bacteria during oxic-anoxic regime shifts in a seasonally stratified lake

Diao, M.; Huisman, J.; Muyzer, G.

DOI
10.1093/femsec/fiy040

Publication date
2018

Document Version
Final published version

Published in
FEMS Microbiology Ecology

License
CC BY

Citation for published version (APA):

General rights
It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations
If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: https://uba.uva.nl/en/contact, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.

UvA-DARE is a service provided by the library of the University of Amsterdam (https://dare.uva.nl)
RESEARCH ARTICLE

Spatio-temporal dynamics of sulfur bacteria during oxic–anoxic regime shifts in a seasonally stratified lake

Muhe Diao, Jef Huisman and Gerard Muyzer*,
†

Department of Freshwater and Marine Ecology, Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, 1090 GE Amsterdam, The Netherlands

*Corresponding author: Gerard Muyzer, Department of Freshwater and Marine Ecology, Institute for Biodiversity and Ecosystem Dynamics, University of Amsterdam, Science Park 904, 1098 XH Amsterdam, The Netherlands, E-mail: g.muyzer@uva.nl

One sentence summary: The succession of sulfur bacteria during oxic–anoxic transitions in a seasonally stratified lake was investigated and summarized in a conceptual model.

Editor: Hendrikus Laanbroek
†Gerard Muyzer, http://orcid.org/0000-0002-2422-0732

ABSTRACT

Sulfate-reducing bacteria (SRB) and sulfur-oxidizing bacteria drive major transformations in the sulfur cycle, and play vital roles in oxic–anoxic transitions in lakes and coastal waters. However, information on the succession of these sulfur bacteria in seasonally stratified lakes using molecular biological techniques is scarce. Here, we used 16S rRNA gene amplicon sequencing to study the spatio-temporal dynamics of sulfur bacteria during oxic–anoxic regime shifts in Lake Vechten. Oxygen and sulfate were mixed throughout the water column in winter and early spring. Meanwhile, SRB, green sulfur bacteria (GSB), purple sulfur bacteria (PSB), and colorless sulfur bacteria (CSB) exclusively inhabited the sediment. After the water column stratified, oxygen and nitrate concentrations decreased in the hypolimnion and various SRB species expanded into the anoxic hypolimnion. Consequently, sulfate was reduced to sulfide, stimulating the growth of PSB and GSB in the metalimnion and hypolimnion during summer stratification. When hypoxia spread throughout the water column during fall turnover, SRB and GSB vanished from the water column, whereas CSB (mainly Arcobacter) and PSB (Lamprocystis) became dominant and oxidized the accumulated sulfide under micro-aerobic conditions. Our results support the view that, once ecosystems have become anoxic and sulfidic, a large oxygen influx is needed to overcome the anaerobic sulfur cycle and bring the ecosystems back into their oxic state.

Keywords: Sulfur-oxidizing bacteria; sulfate-reducing bacteria; green sulfur bacteria; purple sulfur bacteria; regime shifts; stratified lakes

INTRODUCTION

Oxygen depletion may lead to hypoxia and anoxia in lakes, coastal waters and the open ocean, which is detrimental to many aquatic organisms (Diaz and Rosenberg 1995; Vaquer-Sunyer and Duarte 2008; Breitburg et al. 2018). During the past decades, the frequency, intensity and duration of hypoxia and anoxia seem to have increased due to combined effects of eutrophication and global warming, resulting in an expanding area of ‘dead zones’ in eutrophied waters across the globe (Diaz and Rosenberg 2008; Middelburg and Levin 2009; Jenny et al. 2016). The transition from oxic to anoxic water is accompanied
by major changes in microbial community structure and bio-
geochemoical processes. However, a detailed understanding of the different biological and chemical feedbacks involved in oxic–
anoxic transitions is still lacking.

Recently, Bush et al. (2017) developed a mathematical model to investigate interactions between microbial community com-
position and the dissolved oxygen (DO) concentration. The model predicts that the transition from oxic to anoxic water may occur in the form of a regime shift. Regime shifts are defined as abrupt, large and persistent changes in the structure and func-
tion of ecosystems triggered by gradual changes in environmental conditions (Chavez et al. 2003; Scheffer and Carpenter 2003; Biggs, Carpenter and Brock 2009). When the oxygen influx is gradually reduced, at first oxygen-producing cyanobacteria and algae still persist and the lake remains in an oxic state. Once the oxygen concentration is suppressed below a critical threshold, sulfate-reducing bacteria (SRB) and photosynthetic sulfur bacte-
ria take over. The SRB cause an increase in sulfide concentration, which suppresses cyanobacterial growth, and hence the lake rapidly shifts from an oxic into an anoxic state. One of the implica-
tions predicted by the model is that this oxic–anoxic regime shift displays hysteresis. Once the water has turned anoxic, high sulfide concentrations maintained by SRB stabilize the anoxic conditions. As a consequence, a large oxygen influx is needed to bring the system back into its oxic state. These model pre-
dictions are supported by observations from a seasonally strati-
fied lake, which shows hysteresis in the transition between oxic and anoxic states with similar changes in microbial community composition as predicted by the model (Bush et al. 2017). Hysteresis effects have also been reported for anoxia in coastal ecosystems (e.g. Conley et al. 2009; Middelburg and Levin 2009; Zhang et al. 2010).

SRB and sulfur-oxidizing bacteria (SOB) thus play important roles in oxic–anoxic regime shifts, as they drive major transfor-
mations in the sulfur cycle (Muyzer and Stams 2008; Frigaard and Dahl 2009; Ghosh and Dam 2009; Muyzer, Kuenen and Robertson 2013). SRB use sulfite as electron acceptor to degrade organic compounds, producing sulfide as end product (Muyzer and Stams 2008). SOB comprise several groups, including color-
less sulfur bacteria (CSB), green sulfur bacteria (GSB) and pur-
ple sulfur bacteria (PSB). CSB oxidize reduced sulfur compounds with oxygen or nitrate as electron acceptor (Swerts et al. 1990; Muyzer, Kuenen and Robertson 2013). GSB and PSB can use sulfide, elemental sulfur and thiosulfate as electron donors in anoxygenic photosynthesis (Pfennig and Trüper 1989; Frigaard and Dahl 2009; Gregersen, Bryant and Frigaard 2011).

In the past, several excellent attempts at an overarching appraisal of sulfuretum communities have been made (Guerrer-
et al. 1985; Camacho, Vicente and Miracle 2000; Tonolla et al. 2003), but these studies were to some extent hampered by the inherent limitations of classic microscopic observations. With the advancement of modern molecular-genetic techniques, a much deeper understanding of the dynamics, structure and functioning of microbial communities can be obtained. Yet, although some bacterial groups have been studied in detail (Edwardson and Hollibaugh 2017; Llorens-Marés et al. 2017), comprehensive studies of the overall community dynamics of sulfur bacteria in stratified lakes have received less attention in recent years. For instance, Bush et al. (2017) provided data on the seasonal succession of cyanobacteria, phototrophic sulfur bac-
teria and SRB in the metalimnion, but a detailed analysis of the sulfur bacteria at a higher taxonomic resolution and across the entire depth range of the lake is lacking. Other studies have ana-
lyzed the overall composition of lake bacteria at a coarse taxo-
nomic resolution, but do not specifically zoom in at the spatio-
temporal dynamics of the different genera involved in the sulfur cycle (Eiler, Heinrich and Bertilsson 2012; Diao et al. 2017). As a consequence, the diversity, abundance and seasonal dynamics of sulfur bacteria during oxic–anoxic transitions in seasonally stratified lakes are not yet well understood.

Here, we investigated the spatio-temporal dynamics of dif-
ferent functional groups of sulfur bacteria (including SRB, GSB, PSB and CSB) in Lake Vechten, a seasonally stratified lake that is well mixed during winter, while it develops an anoxic and sulfidic hypolimnion during summer stratification (Steenbergen and Korthals 1982; Steenbergen and Verdouw 1982; Bush et al. 2017). Specific objectives of our study were (i) to determine the taxonomic composition, at the genus level, of the SRB, GSB, PSB and CSB in this seasonally stratified lake, (ii) to determine sea-
sonal changes in their diversity, distribution and relative abun-
dances, and (iii) to investigate how their successional trajec-
tories are related to seasonal changes between oxic and anoxic conditions. For this purpose, samples from different water lay-
ers were collected monthly over a period of 1 year, and 16S rRNA gene amplicon sequencing was applied to identify the diversity and composition of the sulfur bacteria. These results were cor-
related to seasonal changes in environmental parameters.

**MATERIALS AND METHODS**

**Lake, sampling and general analysis**

Lake Vechten (52° 04’N, 5° 05’E) is a small eutrophic lake near the city of Utrecht, in the center of The Netherlands. It consists of two connected basins with a total surface area of 4.7 ha, and has a maximum depth of 11.9 m (Steenbergen and Verdouw 1982). Temperature, DO, photosynthetically active radiation (PAR) and pH of lake water were measured with a multiprobe Hydrolab DataSonde 4a (Hydrolab Corporation, Austin, TX, USA). Water samples were collected biweekly to monthly from every meter in the western basin from March 2013 to April 2014. Water was pumped via a hose connected to the Hydrolab to make sure the samples matched the conditions measured at that particular depth. The samples were filtered through 0.20 μm polyethersulfone mem-
brane filters (Millipore, GNWP) to collect bacterial cells. The fil-
ters were frozen immediately and preserved in a −20°C freezer until further processing. Sediment samples (0–10 cm) were col-
lected monthly with a box-corer from the same location, trans-
ported to the laboratory in a dry shipper and stored at −20°C until further analysis. Dry weight of sediment was determined after drying for 2 days in a 60°C oven.

Concentrations of sulfate (SO₄²⁻), ammonium (NH₄⁺) and nitrate (NO₃⁻) in the filtered water samples were measured by an auto-analyzer (SAN⁺⁺, Skalar, The Netherlands). For sulfide, lake water was filtered through 0.20 μm polyethersulfone mem-
brane filters and fixed with zinc acetate (at a final concentration of 2% w/v) in the field. Afterwards, sulfide was measured in the laboratory according to the methylene blue method (Trüper and Schlegel 1964). Dissolved organic carbon (DOC) was measured by a total organic carbon analyzer (TOC-VCPH, Shimadzu, Japan). The data were visualized with Ocean Data View 4.7.8 (Schlitzer 2002).
DNA extraction

DNA was extracted from the collected bacterial cells using the PowerSoil DNA Isolation Kit according to the manufacturer’s instructions (Mo Bio, Laboratories Inc., USA). The concentration of extracted DNA was quantified with the Qubit dsDNA BR Assay Kit (Invitrogen, USA).

Amplon sequencing and operational taxonomic unit assignments

The PCR-amplified 16S rRNA genes of 189 water samples and 11 sediment samples were profiled by denaturing gradient gel electrophoresis (DGGE). Based on the DGGE profiles and measured vertical stratification pattern of Lake Vechten, we selected 51 samples from the water column and 4 samples from the sediment for 16S rRNA gene amplicon sequencing (Table S1, Supporting Information). Sequencing was performed on an Illumina MiSeq system by Research and Testing Laboratory (Lubbock, Texas, USA). The primer pair S-D-Bact-0341-b-S-17 (S′-CTACGGGNGGCWGCAG-3′) and S-D-Bact-0785-a-A-21 (S′-GACTACHVGGGTATCTAATCC-3′) was used to generate paired-end sequence reads, covering the V3-V4 region of the 16S rRNA gene (Herlemann et al. 2011).

The forward and reverse reads were taken in FASTQ format and were merged using the PEAR Illumina paired-end read merger (Zhang et al. 2014). Reads were run through an internally developed quality trimming algorithm. Prefix dereplication was performed using the USEARCH dereplication algorithm (Edgar 2010). Clustering at a 4% divergence was applied using the USEARCH clustering algorithm (Edgar 2010). Operational taxonomic units (OTUs, 97% similarity) were selected using the UPARSE OTU selection algorithm (Edgar 2013). Chimera checking was performed on the selected OTUs using the UCHIME chimera detection software executed in de novo mode (Edgar et al. 2011). All chimeric sequences were removed and the corrected sequences were then written to the output file. After quality filtering a total of 2934 111 sequences were obtained with an average sequence length of 420 bp.

The sequences were run through the USEARCH global alignment program along with a python program to determine the taxonomic information for each sequence (Bokulich et al. 2015), using a database of high-quality sequences derived from the NCBI database up to May 2014. From the top 6 sequence matches, a confidence value was estimated at each taxonomic level (phylum, class, order, family, genus and species). Once confidence values were assigned for each sequence, an RDP-formatted output file was generated for the final analysis in USEARCH. Subsequently, the data were entered into the diversity analysis program that takes the OTU/dereplication table output from sequence clustering along with the output generated during taxonomic identification and generates a new OTU table with the taxonomic information tied to each cluster.

The 16S amplicon sequences have been deposited as dataset SAMN06314865-SAMN06314918 in the Sequence Read Archive (SRA, NCBI).

Statistical analysis

We applied redundancy analysis (RDA; Zuur et al. 2009; Oksanen et al. 2013) to investigate associations between environmental parameters and the taxonomic composition of the sulfur bacteria. To avoid data with many zeros, we selected only those bacterial genera that were present in at least 15 water samples.

The analysis was performed using R (version 3.0.3), with the measured environmental parameters as explanatory variables and the 16S-based relative abundances of the bacterial genera as response variables. All environmental parameters, except pH, were log (x + 1)-transferred prior to the analysis. Subsequently, the number of explanatory variables was reduced by stepwise removal of variables with a high variance inflation factor (VIF) using the R function VIF in the car package (Fox and Weisberg 2011), until only variables with a VIF < 10 remained. Finally, RDA was applied using forward selection with the Ordistep function in the R package vegan to select those explanatory variables that contributed significantly to the RDA model, while removing non-significant terms (Oksanen et al. 2013). Significance was based on a permutation test using the multivariate pseudo-F statistic and 9999 permutations (Zuur et al. 2009).

RESULTS

Environmental conditions

The temperature in Lake Vechten was almost uniform over the entire depth in early spring, while the water column was saturated with oxygen (Fig. 1A and B). When the surface temperature increased in April, the water column stratified. The euphotic zone remained oxic, whereas the hypolimnion became anoxic during summer stratification. The stratification persisted until the water column was mixed during fall turnover. The oxygen concentration in the surface layer decreased during fall turnover, creating hypoxia throughout the water column in November. This was followed by re-oxygenation of the water column during the winter period (Fig. 1B).

The sulfate concentration was 60–90 μM throughout the water column in early spring, while the sulfide concentration was negligible during this period (Fig. 1C and D). When the lake stratified, the sulfate concentration in the anoxic hypolimnion decreased to less than 10 μM, whereas sulfide increased to 10–20 μM. Interestingly, the relatively high sulfide concentration of the hypolimnion did not expand into the surface layer when hypoxia spread throughout the water column during fall turnover. Instead, sulfide vanished during fall turnover, while the sulfate concentration increased to 30–50 μM (Fig. 1D).

The nitrate concentration was relatively high in early spring, was depleted throughout the entire water column from May until December, and increased again when the lake was mixed during the winter period (Fig. S1A, Supporting Information). Conversely, both ammonium and DOC had low concentrations in the winter period, but accumulated in the anoxic hypolimnion during summer stratification (Fig. S1B and S1C, Supporting Information).

To investigate light penetration in Lake Vechten, we measured the euphotic depth (i.e. the depth at which irradiance in the PAR range is 1% of the surface irradiance), which indicates the maximum depth of the light zone suitable for oxygenic photosynthesis by cyanobacteria and eukaryotic phytoplankton. Because anoxicogenic phototrophs (in particular GSB) can grow at lower light levels than phytoplankton species (Overmann, Cypionka and Pfennig 1992; Manske et al. 2005), we introduce the term ‘bacterial euphotic depth’ (which we define as the depth at which the irradiance is 0.1% of the surface irradiance). Light penetrated quite deep into the water column in March 2013, but the euphotic depth and bacterial euphotic depth shifted upwards to <2.5 m and <3.5 m, respectively, during the phytoplankton spring bloom in April (Fig. 2). When the phytoplankton bloom in
Figure 1. Spatio-temporal dynamics of environmental variables in Lake Vechten. (A) Temperature; (B) dissolved oxygen; (C) sulfate; (D) sulfide.
the surface layer declined in June, presumably because of nutrient limitation (Diao et al. 2017), light could enter into the deeper water layers again and bacterial euphotic depth reached 10.6 m in July. In the subsequent spring, the euphotic depth and bacterial euphotic depth shallowed again during the phytoplankton spring bloom.

**Microbial community composition**

Amplicon sequencing of bacterial 16S rRNA genes showed that SRB were represented by Deltaproteobacteria, including members of the families Desulfovibulaceae, Desulfovibrionaceae, Desulfobacteraceae, Desulfuromonadaceae and Syntrophaceae (Table 1). Desulfobulbus was the most abundant SRB genus in the water column, while Desulfobacterium was the dominant SRB genus in the sediment. GSB belonged to the family Chlorobiaceae of the phylum Chlorobi, while PSB belonged to the family Chromatiaceae of the class Gammaproteobacteria. CSB in the water column were mainly from the class Epsilonproteobacteria (with Arcobacter as the most abundant genus), whereas CSB in the sediment were dominated by Betaproteobacteria (mostly Thiobacillus). In total, the number of genera belonging to the sulfur bacteria was higher in the water column than in the sediment.

Spatio-temporal dynamics of the most abundant genera of sulfur bacteria are shown in Fig. 3. Members belonging to the SRB genera Desulfobulbus and Desulfovibrio were present in the metalimnion and hypolimnion when the lake became stratified (Fig. 3A and B). Similar distributions were observed for the SRB genera Desulfovibulbaceae and Desulfomonile (Fig. S2, Supporting Information). GSB of the genus Chlorobi and other unknown genera were present in the metalimnion and hypolimnion only when the lake was stratified (Fig. 3C and D). PSB of the genus Thiodictyon reached high relative abundances in the metalimnion (Fig. 3E), where it accounted for more than 20% of the total bacterial community in late summer. Furthermore, Thiocystis was found in the hypolimnion in late spring, Thiorhodacoccus in the metalimnion in the summer and hypolimnion in the fall, and Lamprocystis in the hypolimnion in the fall (Fig. 3F and Fig. S3, Supporting Information). The CSB genera Arcobacter and Sulfurimonas were prevalent at 7–10 m depth in November and December (Fig. 3G and H). Especially Arcobacter was highly abundant, constituting almost 60% of the total bacterial community in the bottom water layer in December.

In the sediment, SRB were by far the most abundant sulfur bacteria throughout the seasons (Fig. 4). The relative abundance of PSB was quite high in spring, decreased sharply during summer stratification, and increased again after fall turnover. There were no clear seasonal changes in the relative abundances of GSB and CSB in the sediment (Fig. 4).

**Linking sulfur bacteria with environmental drivers**

Redundancy analysis was applied to correlate the relative abundances of sulfur bacteria (at genus level) in the water column with environmental variables. In total nine explanatory variables had a VIF < 10, including temperature, DO, PAR, NH₄⁺, NO₃⁻, PO₄³⁻, SO₄²⁻, DOC and sulfide. Forward selection revealed that four of these nine variables were significant in the redundancy analysis: sulfide, DO, SO₄²⁻ and NH₄⁺ (Table S2, Supporting Information).

The first and second axis of the RDA plot explained 27.5% and 4.8% of the variation in the composition of sulfur bacteria, respectively (Fig. 5). Relative abundances of SRB (Desulfovibulbus, Desulfovibrio, Desulfomonile, unknown Desulfobulbacaeae) and GSB (Chlorobi, unknown Chlorobiaceae) were positively correlated with the sulfide concentration, and negatively correlated with DO. Furthermore, some PSB (Thiodictyon), GSB (unknown Chlorobiaceae) and SRB (unknown Desulfobulbacaeae) were negatively correlated with the sulfate concentration, whereas other PSB (Lamprocystis) were positively correlated with the ammonium concentration.

**DISCUSSION**

Our results show that Lake Vechten is characterized by distinct seasonal changes in the oxygen, sulfate and sulfide concentrations and a marked seasonal succession of a wide diversity of sulfur bacteria. For a proper interpretation of these results, it is important to realize that sulfur bacteria do not only track changes in oxygen and sulfur availability, but are also important drivers of these changes as they play key roles in oxidation-reduction reactions of the sulfur cycle. We will first discuss our observations on the taxonomic composition and dynamics of different functional groups of sulfur bacteria, and then summarize these results into a conceptual model of the seasonal succession of sulfur bacteria during oxic-anoxic regime shifts in seasonally stratified lakes.

**Sulfate-reducing bacteria**

SRB are anaerobic bacteria that oxidize organic matter (and hydrogen) using sulfate as terminal electron acceptor (Muyzer and Stams 2008). During seasonal succession in Lake Vechten, SRB were preceded by denitrifying bacteria, which reached peak abundances in the hypolimnion at the onset of lake stratification in May (data not shown). Sulfate reduction has a lower energy yield than nitrate reduction. Hence, as expected, SRB replaced the denitrifying bacteria in the hypolimnion and metalimnion of the lake during the summer months, after nitrate had been depleted. SRB produced sulfide, and their presence coincided with the observed decline of the sulfate concentration and accumulation of sulfide in the hypolimnion of the lake.

Based on bacterial 16S rRNA gene amplicon sequencing, Desulfovibulbus, Desulfovibrio, Desulfobacterium, Desulfomonile and...
Table 1. Diversity of sulfur bacteria in Lake Vechten.a

<table>
<thead>
<tr>
<th>Group</th>
<th>Phylum/Class</th>
<th>Family</th>
<th>Water column</th>
<th>Sediment</th>
</tr>
</thead>
<tbody>
<tr>
<td>SRB</td>
<td>Deltaproteobacteria</td>
<td>Desulfobulbaceae</td>
<td>Desulfobulbus</td>
<td>Desulfobulbus</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Desulfovibrionaceae</td>
<td>unknown</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Desulfobacteraceae</td>
<td>Desulfobacterium</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Desulfomonadaceae</td>
<td>unknown</td>
<td>Desulfomonas</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Syntrophaceae</td>
<td>Desulfomonile</td>
<td>n.d.</td>
</tr>
<tr>
<td>GSB</td>
<td>Epsilonproteobacteria</td>
<td>Campylobacteraceae</td>
<td>Sulfurospirillumb</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td>Chlorobi</td>
<td>Chlorobiaceae</td>
<td>unknown</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Chlorobium</td>
<td>Desulfobacterium</td>
</tr>
<tr>
<td>PSB</td>
<td>Gammaproteobacteria</td>
<td>Chromatiaceae</td>
<td>Thiocystis</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Lamprocystis</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Thiocystis</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Thiorhodococcus</td>
<td>n.d.</td>
</tr>
<tr>
<td>CSB</td>
<td>Epsilonproteobacteria</td>
<td>Campylobacteraceae</td>
<td>Arcobacter</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Helicobacteraceae</td>
<td>Sulfurospirillumb</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td>Betaproteobacteria</td>
<td>Hydrogenophilaceae</td>
<td>Sulfurimonas</td>
<td>n.d.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Thiobacillus</td>
<td></td>
</tr>
</tbody>
</table>

aOnly genera representing >0.1% of the total bacterial community in at least one of the samples are shown; n.d. = not detected.
bSulfurospirillum can reduce elemental sulfur (Sorokin, Tourova and Muyzer 2013) and it can also oxidize sulfide with nitrate (Eisenmann et al. 1995). Therefore, it is listed in both the SRB and GSB.

Desulfuromonas were the dominant SRB genera in the metalimnion and hypolimnion of Lake Vechten, which was also found in previous studies for other freshwater lakes (Kondo et al. 2006; Kubo, Kojima and Fukui 2014).

Although we did not detect SRB in the water column in winter, SRB were present in the sediment during the winter period, when an influx of sulfate from the water column into the sediment may have stimulated their activity. Desulfobulbus and Desulfobacterium were found in the sediment throughout the year and in the anoxic hypolimnion during summer. Hence, most likely the sediment acted as a seed bank (sensu Lennon and Jones 2011) for these SRB, which initiated the succession of SRB in the anoxic hypolimnion during summer stratification.

Green sulfur bacteria

GSB are commonly found in the anoxic water layers of stratified lakes and marine ecosystems (Mori et al. 2013; Saarenheimo et al. 2016). They are often found below a layer of PSB, presumably because GSB are better adapted to very low light levels than PSB (Biebl and Pfennig 1978; Guerrero et al. 1985; Vila and Abella 1994). For instance, a strain isolated from the chemoclone of the Black Sea, at 100 m depth, could still photosynthesize at 0.015 μmol quanta m−2 s−1 and its photosynthetic rate reached light saturation at 1 μmol quanta m−2 s−1 (Overmann, Cyponka and Pfennig 1992; Manske et al. 2005), which is 0.05%–0.10% of the surface irradiance on a sunny day. Furthermore, GSB are obligate anaerobes, whereas many PSB can tolerate and sometimes even utilize oxygen. Hence, the anoxic hypolimnion of Lake Vechten provided suitable conditions for GSB during the summer period. Interestingly, GSB reached peak abundances in the hypolimnion from July onwards (Fig. 3C and D). At this time, light levels reaching the hypolimnion may have been sufficient for GSB photosynthesis, just after the development of the phytoplankton spring bloom that prevented light from reaching the hypolimnion in April and May (Fig. 2).

The GSB in our study showed a similar spatio-temporal distribution as the SRB. This may indicate that sulfide produced by SRB stimulated the proliferation of GSB, as has been observed in mixed laboratory cultures (Biebl and Pfennig 1978). Conversely, the photosynthetic activity of GSB and PSB produces oxidized sulfur species that can be utilized as terminal electron acceptor by SRB. This mutual reciprocity may result in a tight sulfur cycle in which sulfur is shuttled back and forth between the different functional groups, fueled by the photosynthetic activity of GSB and PSB and the degradation of organic matter by SRB (Hamilton et al. 2014; Bush et al. 2017).

Purple sulfur bacteria

Bacterial 16S rRNA sequencing data revealed four different PSB genera in the meta- and hypolimnion of Lake Vechten. Thiocystis peaked just above the sediment in early May, soon after the onset of lake stratification. At that time, a dense phytoplankton spring bloom absorbed most of the incident light, and although the spectral distribution of the underwater light field was not measured, light penetration to 10 m depth was less than 0.1 μmol photons m−2 s−1 in the PAR range (cf. Fig. 2), which seems too low to support photoautotrophic growth of PSB. However, it is known that several PSB are not obligate photoautotrophs, but can also grow in the dark as chemolithoautotrophs oxidizing reduced sulfur compounds under micro-aerobic and sometimes even oxic conditions (Schaub and van Gemerden 1994; Camacho, Vicente and Miracle 2000; Casamayor, García-Cantizano and Pedrós-Alió 2008). Chemolithoautotrophic and chemoheterotrophic growth has been reported for several Thiocystis spp. (Kämpf and Pfennig 1980; Peduzzi et al. 2011), and offers a plausible explanation for the peak abundance of Thiocystis just above the sediment when the lake started to develop a sulfidic hypolimnion.

Thiocystis and to a lesser extent also Thiorhodococcus reached high relative abundances in the metalimnion during the summer months. Light levels reaching the metalimnion in the period...
July–September were roughly ∼1% of the surface light intensity, providing suitable light conditions for photoautotrophic growth of PSB. *Thiodictyon* cells are non-motile, but often form aggregates and contain large gas vesicles (Cohen-Bazire, Kunisawa and Pfennig 1969; Walsby 1994). Hence, *Thiodictyon* probably reached high relative abundances in the metalimnion through buoyancy regulation provided by its gas vesicles. *Thiorhodococcus* is a motile bacterium (Guyoneaud, Caumette and Imhoff 2015), which may have enabled its aggregate in the metalimnion.

*Lamprocystis* became dominant in the deeper parts of the lake in November. It is possible that it could benefit from the very low light levels at these depths, but some *Lamprocystis* species can also grow in the dark as chemolithoautotrophs under micro-aerobic conditions (Imhoff 2001). Hence, *Lamprocystis* may have contributed to the chemotrophic oxidation of sulfide when deep mixing brought low oxygen levels into the former hypolimnion during fall turnover.
Colorless sulfur bacteria

CSB are key players in the oxidation of reduced sulfur compounds, and often thrive under micro-aerobic and aerobic conditions. In Lake Vechten they were represented by Epsilonproteobacteria and Betaproteobacteria, similar to several other freshwater lakes (Biderre-Petit et al. 2011; Yang et al. 2013; Hamilton et al. 2014). The CSB genera Arcobacter and Sulfurimonas became abundant in deep water layers during the fall, where they most likely were involved in oxidation of the accumulated sulfide using the low oxygen levels that were mixed into the former hypolimnion during fall turnover. Arcobacter is a highly motile bacterium that can form swarms at the oxic–anoxic interface, and is able to grow at very low oxygen concentrations in comparison to other CSB (Sievert et al. 2007). Similar seasonal shifts from phototrophic sulfur bacteria to CSB have been reported in the chemocline of a meromictic lake in winter (Noguerola et al. 2015) and in a seasonally stratified marine lake after fall turnover (Pjevac et al. 2015), most likely because the increasing oxygen levels suppress GSB and to a lesser extent also PSB while favoring the oxidation activities of CSB.

Previous studies have reported that sulfur-oxidizing Beggiatoa were abundant in the mats on the sediment of Lake Vechten (Sweerts et al. 1990), but we have not detected Beggiatoa or close relatives in the water column or sediment of Lake Vechten. Instead, Thiobacillus was the dominant CSB in the sediment, particularly during the winter period.

Seasonal succession of sulfur bacteria

Our observations and those from previous studies can be summarized in a simple conceptual model of the seasonal succession of sulfur bacteria in eutrophic lakes (Fig. 6). In winter and early spring, the water column is oxic and rich in sulfate. At this time, the sediment contains various sulfur bacteria including SRB using sulfate as terminal electron acceptor, and CSB and PSB oxidizing the upward flux of sulfide. After the water column stratifies in spring, oxygen and nitrate in the hypolimnion are depleted and subsequently a variety of SRB species expand into the anoxic hypolimnion, reducing sulfate to sulfide. During the summer period, photosynthetically active PSB bloom in the metalimnion and GSB thrive in the hypolimnion underneath, stimulated by high sulfide concentrations. When the lake is mixed during fall turnover, low oxygen concentrations are brought into the former hypolimnion, and SRB and GSB disappear from the water column because they are inhibited by oxygen. A combination of CSB and chemolithoautotrophic PSB flourish under these micro-aerobic conditions, oxidizing sulfide into sulfate. In winter, when all sulfide in the water column has been removed andoxic conditions in the water column are fully restored, sulfur bacteria disappear from the water column, but remain active in the sediment.

To the best of our knowledge, this is the first study that investigates the succession of SRB, GSB, PSB and CSB during oxic–anoxic transitions at a high taxonomic resolution (genus level) and summarizes the succession into a conceptual model. Admittedly, this simple conceptual model can still be improved in many ways. In particular, it will be an interesting challenge to assess in future studies which of these aspects are specific for Lake Vechten and which will apply to the seasonal succession of sulfur bacteria in other aquatic ecosystems. As discussed above, the similarities with other lake studies are promising (e.g. Noguerola et al. 2015; Pjevac et al. 2015). Similarly, dynamics of nitrate and ammonium also showed seasonal patterns associated with the oxic–anoxic transitions (Fig. S1A and S1B, Supporting Information). A closer look at the 16S rRNA gene sequencing data revealed an increased relative abundance of denitrifiers during the first month of stratification in May (data not shown), when the hypolimnion became anoxic and nitrate concentrations in the hypolimnion were depleted. Furthermore, ammonium-oxidizing bacteria were detected in the sediment during fall turnover, accompanied by the accumulation of nitrate and decrease of ammonium. Meanwhile, methane-oxidizing bacteria (Methylbacterium) flourished in the water column during fall turnover (see Fig. 5C in Diao et al. 2017), indicative of methane oxidation.
Figure 6. Conceptual model of the seasonal dynamics of sulfur bacteria in the water column and sediment of Lake Vechten. The model shows the seasonal stratification and oxygen conditions. Line indicates the bacterial euphotic depth. The circles represent the different functional groups (colors) and genera (numbers). Grey circles represent sulfate-reducing bacteria (1, Desulfobulbus; 2, Desulfovibrio; 3, Desulfbacterium; 4, unknown Desulfbacteraceae; 5, Desulfuromonas; 6, Desulfomonile), purple circles represent purple sulfur bacteria (7, Thiolyction; 8, Lamprocystis; 9, Thiocyctis; 10, Thiorhodococcus), green circles represent green sulfur bacteria (11, unknown Chlorobiaceae; 12, Chlorobium) and white circles represent colorless sulfur bacteria (13, Arcobacter; 14, Sulfurimonas; 15, Thiobacillus). The size of the circles gives a rough indication of the relative abundances of the different bacteria.

Recently, we showed that the interplay between microbial community dynamics and oxidation–reduction processes result in oxic–anoxic regime shifts characterized by tipping point and hysteresis effects (Bush et al. 2017). More specifically, degradation of organic matter by SRB creates anoxic and sulfidic conditions (also known as euxinia), and once ecosystems have become euxinic a large oxygen influx will be required to suppress the SRB, to overcome the low reduction potential that they have generated, and to bring the ecosystems back into their oxic state. Our results support and extend these findings. We observed that SRB were involved in the development of euxinia, that SRB remained abundant and euxinia was sustained throughout the summer period and that the hypolimnion did not return to its oxic state immediately after fall turnover. Instead, hypoxia developed throughout the water column in the fall, presumably because oxidation of the accumulated sulfide by CSB (e.g. Arcobacter) and chemolithotrophic PSB (e.g. Lamprocystis) consumed oxygen and thereby kept the DO concentrations low. Similarly, microbial oxidation of ammonium and other reduced compounds (e.g. methane) accumulated in the hypolimnion will also have consumed part of the oxygen influx. These results indicate that once ecosystems have become anoxic, the former oxic conditions are not easily restored.

SUPPLEMENTARY DATA

Supplementary data are available at FEMSEC online.

ACKNOWLEDGEMENTS

MD was supported by the China Scholarship Council (CSC No. 201206220165). GM was supported by the Research Priority Area Systems Biology of the University of Amsterdam and the ERC Advanced Grant PARASOL (No. 322551). We thank Pieter Slot and Bas van Beusekom for assistance during the fieldwork, Cherei Balkema and Bart André Meijjs for assistance during lab work, Leo Hoitinga and Peter Serné for nutrient analyses, and the reviewers for constructive comments on the manuscript.

Conflict of interest. None declared.

REFERENCES


Edwardson CF, Hollibaugh JT. Metatranscriptomic analysis of prokaryotic communities active in sulfur and arsenic cycling in Mono Lake, California, USA. ISME J 2017;11:2195–208.


Sorokin DY, Tourova TP, Muyzer G. Isolation and characterization of two novel alkalitolerant sulfidogens from a Thiopaq bioreactor, Desulfonatronum alkalitolerans sp. nov., and Sulfospirillum alkalitolerans sp. nov. Extremophiles 2013;17:535–43.