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Short-limbed dwarfism with bowing, combined immune deficiency, and late onset aplastic anaemia caused by novel mutations in the RMRP gene

T W Kuijpers, M Ridanpää, M Peters, I de Boer, J M J J Vossen, S T Pals, I Kaitila, R C M Hennekam

Key points

- A female patient was diagnosed with kyphomelic dysplasia in infancy, because of her short limbed dwarfism and kyphomelia, especially of the femurs.
- She developed a combined aplastic anaemia and immunodeficiency of late onset that responded well to allogeneic bone marrow transplantation, although growth remained extremely retarded.
- Clinical and radiological symptoms gradually changed and became more typical for cartilage-hair hypoplasia, as was confirmed by two novel mutations in the RMRP gene (insT195/C63T).
- Kyphomelic dysplasia constitutes a heterogeneous group of different disorders, which includes cartilage-hair hypoplasia.

Histochemistry

Bistochemistry and immunophenotypical analysis of the thymus and lymph nodes was carried out with the use of standard staining procedures and a streptavidin-biotin complex method on paraffin embedded sections, and a three step indirect immunoperoxidase method with AEC as a substrate on frozen sections. Primary antibodies used were CD3 (pan T cell), CD4 and CD8 for T cells (thymocytes and mature T helper cells/cytotoxic T cells), CD68 (pan macrophage), CD20 and CD79A (mature B cells), CD25 (IL-2Rα, thymocytes and activation of mature T cells), Ki67 (proliferating cells) and immunoglobulin heavy chains (IgA, IgM and IgG).

Molecular studies

Genomic DNA from a fibroblast cell line established from the patient's cells was extracted using standard methods. The

Abbreviations: BMT, bone marrow transplantation; CHH, cartilage-hair hypoplasia; MRP, mitochondrial RNA processing

METHODS

Immunological studies

Lymphocyte phenotyping

Absolute numbers of B cell (CD19+), T cell (CD2+, CD3+, CD4+, CD8+), and NK cell (CD3−, CD16+, CD56+) subsets were determined by standard FACScan procedures, with MAbs being produced by the Central Laboratory of the Netherlands Red Cross Blood Transfusion Service. CD3, CD4, CD8, CD19, TcRαβ and TcRγδ MAbs conjugated with FITC and PerCP were all purchased from Becton-Dickinson (San Jose, CA, USA). Using PE conjugated CD45RA and CD45RO MAbs from Coulter Immunology (Halcath, FL, USA) for staining, gating on CD3+ lymphocytes allowed further subtyping into functional CD4+ and CD8+ T cell subpopulations, and naive and memory cells, respectively. After lysing the erythrocytes, lymphocytes were fixedate using Cellfix (Becton-Dickinson) and analysed on a FACScan (Becton-Dickinson) with Cellquest software, using a lymphocyte gate.

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whole RNA coding region of RMRP (Genbank accession number M29916) was amplified in two separate PCRs using primers RMF 5’-CCAACCTTCTACCCCTAACC-3’ and RMR 5’-AGGGCGAAGACACCGGTAAA-3’. PCR products were run on 1% SeaKem (FMC BioProducts) gels, purified using Qiagen Gel Extraction Kit (Qiagen, Valencia, USA) and sequenced automatically (ABI3100; Applied Biosystems, Foster City, LA, USA).

CLINICAL REPORT
The proband is the first child of a non-consanguineous couple, with no family history of congenital abnormalities. Her 26 year old mother is white and her 39 year old father is of African-American descent; both are healthy. She has a healthy younger brother. Intrauterine growth retardation was observed from the 24th week of gestation. Ultrasound examination showed skeletal abnormalities, including shortening and bowing of the limbs. Campomelic dysplasia was suspected. The girl was born by vaginal delivery after 40 weeks and 4 days gestation, birth weight 2790 g (10th percentile), length 38 cm (<3rd percentile), OFC 35 cm (10th percentile). APGAR scores after 1 and 5 min were 9 and 10, respectively. Physical examination showed abnormal, short, curved limbs, most striking in the femur, and a small chest. Hands and feet appeared normal at that time. Radiographs showed short, flared ribs, vertebral flattening, and bowed femur, humerus, tibia and radius. No firm diagnosis was made at that time. The child was re-evaluated at 1 year of age, when the diagnosis of kyphomelic dysplasia was made (figs 1A and B).

During the first 2 years, she suffered from recurrent upper respiratory infections and diarrhoea, twice necessitating admission. She recovered following rehydration and antibiotics. Laboratory findings were normal at that time.

At the age of 2 years and 5 months, she was admitted for the third time because of loss of appetite while having chronic diarrhoea. There were no signs of infections or haemorrhagic diathesis. Examination on admission showed a pale, dystrophic and tachypnoeic girl (fig 2), with a weight of 7.3 kg (percentile ±4SD) and a height of 60 cm (percentile ±4SD). Tonsils were present; lymph nodes were not enlarged. There was hepatomegaly in the absence of splenomegaly or clinical jaundice. Haematological studies were compatible with aplastic anaemia, as described in table 1. No proof for a recent (viral) infection was obtained. Bone marrow was hypocellular without erythropoiesis, and the myeloid lineage was hypoplastic; number and morphology of megakaryocytes was normal. Clonal derangements or myelodysplasia were excluded by morphology, immunophenotyping, and cytogenetic studies for chromosomal abnormalities (such as monosomy 7); the chromosomes were 46XX.

Immunological studies showed defects as described in table 1. Lymphocyte proliferation tests were severely impaired upon activation with mitogens or combined CD3/CD28 receptor signalling (not shown). A normal sized thymus was present on computerised tomography scanning. Pathological studies showed a hypocellular biopsy specimen lacking Hassal’s bodies (fig 3A). Lymph node biopsy showed hypocellularity with depleted T cell areas, and some follicular structures with extremely small numbers of IgA and IgG positive plasma cells (fig 3B). It was concluded that the patient had a combined immune deficiency of late onset. An infection was ruled unlikely because of negative microbiological cultures and PCR tests for several viruses (cytomegalovirus, Epstein-Barr virus, HIV, parvovirus B19) in nose washings, faeces, urine, blood, and bone marrow.

The patient had borderline malabsorption, as indicated by the low serum levels of fat soluble vitamins A and E (0.7 nmol/L and 7.3 mol/L, respectively). However, 25OH-vitamin D was normal (21 nmol/L), as were the levels of water soluble folic acid and vitamin B12. From stool collection over 72 hrs, the fat absorption coefficient was calculated at 85% (low–normal), with unaltered pancreatic function as demonstrated by normal levels of faecal chymotrypsin (3.0 g/g faeces), and serum amylase and lipase. Faeces were negative...
for known pathogens, in particular *Giardia, Cryptosporidium*, and microsporidial species. Duodenal biopsies did not reveal any abnormality (such as mucosal flattening or absence of mucosal/intraepithelial lymphocytes). Liver and kidney functions were normal as determined by plasma levels of bilirubin, ALAT, ASAT, and creatinine. However, alkaline phosphatase, which had been normal at birth, fluctuated between 7000 and 11000 U/L in the presence of normal levels of parathormone (2.8 pmol/L), calcitonine (0.14 g/L), serum calcium (2.30 mmol/L) and phosphate (1.85 mmol/L). Further endocrinological studies excluded any defect in thyroid function or the GH/IGF-1 or hypothalamo-pituitary-adrenal axis.

At 3 years of age the patient was referred for allogeneic bone marrow transplantation (BMT) from her HLA identical brother. The conditioning regimen consisted of busulphan (20 mg/kg total dose) on days −9 to −6 and cyclophosphamide (200 mg/kg/dose) on days −5 to −2. The patient received $2.5 \times 10^8$ nucleated cells/kg (recipient body weight). Graft vs host disease prophylaxis consisted of methotrexate on day +1, +3, and +6 and ciclosporin A from day −1 to +180.

Post-transplantation supportive care consisted of total gastrointestinal decontamination in a strict protective (sterile) environment. Haematological engraftment was achieved on day +17; platelet transfusion independence on day +28. One day later, she was discharged. At 6 months post-transplantation, complete immunological engraftment, with normal T and B cell function, was present. Donor chimerism was complete. She has recently had chickenpox, as has her healthy brother; her condition up to now is excellent without medication.

Her growth remained unaltered by the BMT. A biochemical decrease in alkaline phosphatase was seen during the preconditioning period, reaching normal levels at day +10 post-BMT. Bone age has a considerable lag of about 4 years. She was last evaluated at the age of 8 years. She had mild constipation treated with dietary regimens but did not need any medication. Growth and implantation of her hair was considered to be normal considering her ethnic descent. In addition, eyebrows, eyelashes, and nails were normal. She had a short thorax, mild asymmetric pectus carinatum, prominent abdomen caused by her expressed hyperlordosis, and mild scoliosis. There were three café-au-lait spots on her thorax. Her hands, fingers, feet, and toes had become much shortened compared with earlier ages. Joint mobility was normal. The upper arms were mildly bowed, the upper legs were severely bowed. Radiologically, she had expressed metaphyseal widening and sclerosis, especially in the upper femur, and around the knees and ankles. The metaphyses
showed cystic radiolucencies, especially around the knees and ankles. There was mild bowing of the humerus and tibia, and extensive bowing of the femur. The tibia was larger than the fibula. The height of the vertebral bodies was normal, as was the caudal widening of the interpediculate distances. The hand radiographs showed short phalanges and metacarpals, sclerotic epiphyses of the distal phalanges, cupped epiphyses of all middle phalanges but not of the proximal phalanges, and irregular epiphyses and metaphyses of the distal radius and ulna.

Sequence analysis showed the patient to be compound heterozygous for two mutations in the transcript of RMRP. The first mutation was a base substitution of T for C at nucleotide 63. This mutation was previously identified in an Australian CHH patient (unpublished results). The other mutation was a novel duplication of T at nucleotide 195. Both mutations resided in evolutionarily conserved nucleotides and were not found in healthy controls. Molecular studies in the parents showed that the father had the insT195 and the mother to have the C63T mutation. Thus, molecular gene analysis to verify for CHH now demonstrated two novel mutations in the RMRP gene.

**DISCUSSION**

There are several short limb/skeletal dysplasias that can be associated with variable degrees of immune deficiency. In the present case the diagnosis kYPHOMELIC DYSPLASIA was made at the age of 1 year because of the expressed bowing of the femur, the platyspondyly, and only mild metaphyseal changes especially around the knees, cystic radiolucencies, and normal height of the vertebral bodies became clear, all pointing to CHH. However, several other radiolucencies, and normal height of the vertebral bodies were not present, while the femoral bowing was unusually severe. The mutations in the RMRP gene proved that CHH was the correct diagnosis.

Some of the earlier reports on cases diagnosed as kYPHOMELIC DYSPLASIA or reminiscent of the diagnosis may have been CHH instead, as they demonstrated haematological defects, a clear cut immunodeficiency, or mortality at an early age caused by infection. Molecular studies for mutations in the RMRP gene in other cases diagnosed as kYPHOMELIC DYSPLASIA are essential, as are careful analyses for other, similar entities (for instance, plasma levels of alkaline phosphatase and molecular studies of collagen type I mutations for hypophosphatasia and osteogenesis imperfecta, respectively). It remains uncertain whether all cases described as having kYPHOMELIC DYSPLASIA will be found to have symptoms fitting other known entities, or whether it will be confirmed as a separate entity characterised by short-limbed dwarfism and kYPHOMELIC DYSPLASIA.

The present case suffered from a highly unusual form of aplastic anaemia and combined immune deficiency, both of a late onset type, without the typical red cell abnormalities usually observed in CHH. The immune defect was most notable in her T cell compartment, similar to the immune defect most commonly found in CHH. Under normal conditions, the thymus generates sufficient numbers of naive CD45RA+ T lymphocytes (both CD4+ and CD8+) for early development and continuous maintenance of the immune system to enable the host to respond to newly encountered antigens during life. In this patient, thymic output had been progressively lost. First, circulating naive CD45RA+ T cells were no longer present. Secondly, the ability to show a memory response to tetanus toxoid by the remaining CD45RO+ T cells and B cells was still intact, but the ability to respond to a so-called primary or neo-antigen, keyhole limpet antigen (KLH; see table 1), was completely absent. The exhaustion of the thymic function was further indicated by a specimen of the thymus, showing a complete absence of the so-called Hassal’s bodies (fig. 3A). The normal size argues against a primary thymus dysplasia in our patient. Moreover, complete restoration upon HLA identical allogeneic BMT is further support for a normal thymus anlage and function. To our knowledge, these findings have not been reported previously in CHH.

The patient’s haematological and immunological course is highly unusual for its late onset. The symptoms could be

**Table 1** Haematological and immunological blood values of the propositus

<table>
<thead>
<tr>
<th>Hematology</th>
<th>Birth (nt)</th>
<th>1 year</th>
<th>2 years</th>
<th>2½ years</th>
<th>3 years (BMT -5 days)</th>
<th>5 years (BMT -2 years)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hb (g/dL)</td>
<td>15.9</td>
<td>9.5</td>
<td>2.0</td>
<td>8.6</td>
<td>13.1</td>
<td></td>
</tr>
<tr>
<td>Ht [%]</td>
<td>0.53</td>
<td>0.33</td>
<td>0.75</td>
<td>0.40</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Leucocytes (10⁹/mL)</td>
<td>NT</td>
<td>3.2</td>
<td>&lt;0.1</td>
<td>0.40</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MCV (fl)</td>
<td>NT</td>
<td>90</td>
<td>79</td>
<td>83.7</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Platelets (10⁹/mL)</td>
<td>134</td>
<td>150</td>
<td>480</td>
<td>20†</td>
<td>379</td>
<td></td>
</tr>
<tr>
<td>Leukocytes (10⁷/mL)</td>
<td>18.9</td>
<td>9.4</td>
<td>1.1</td>
<td>0.1</td>
<td>8.1</td>
<td></td>
</tr>
<tr>
<td>% neutrophils</td>
<td>68</td>
<td>52</td>
<td>22</td>
<td>52</td>
<td></td>
<td></td>
</tr>
<tr>
<td>% monocytes</td>
<td>6</td>
<td>10</td>
<td>8</td>
<td>6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>% lymphocytes</td>
<td>22</td>
<td>35</td>
<td>67</td>
<td>42</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IgG (g/L)</td>
<td>11.5</td>
<td>9.6</td>
<td>5.2</td>
<td>11.3</td>
<td>13.1</td>
<td></td>
</tr>
<tr>
<td>IgA (g/L)</td>
<td>1.4</td>
<td>1.0</td>
<td>0.4</td>
<td>1.4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IgM (g/L)</td>
<td>1.0</td>
<td>0.7</td>
<td>0.3</td>
<td>1.3</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lymphocytes (10⁹/mL)</td>
<td>42</td>
<td>3.3</td>
<td>0.7</td>
<td>3.4</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CD3+ T cells</td>
<td>0.3</td>
<td>0.9</td>
<td>26</td>
<td>1.9</td>
<td>1.5</td>
<td></td>
</tr>
<tr>
<td>CD3+/4+ Th cells</td>
<td>0.06</td>
<td></td>
<td></td>
<td>1.03</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CD45RA</td>
<td>8%</td>
<td>81%</td>
<td>82%</td>
<td>81%</td>
<td>81%</td>
<td></td>
</tr>
<tr>
<td>CD45RO</td>
<td>84%</td>
<td>22%</td>
<td>42%</td>
<td>42%</td>
<td>42%</td>
<td></td>
</tr>
<tr>
<td>CD3+/8+ Tc cells</td>
<td>0.1</td>
<td>0.7</td>
<td>1.1</td>
<td>1.1</td>
<td>1.1</td>
<td></td>
</tr>
<tr>
<td>CD19+ B cells</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
<td>0.34</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CD16/56+ NK cells</td>
<td>0.2</td>
<td>0.2</td>
<td>0.2</td>
<td>0.34</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Outside the normal range of values.
†Post-BMT transfusion values.
compatible with a dramatic change at the level of the haematopoietic stem cell, resulting in a developmental arrest in all lineages, including the T cell development in the thymus. Megakaryocytic development was spared, and platelet counts in the blood remained relatively normal. Although unexplained, this phenomenon suggests that the bone marrow stroma was unaffected by the disease. Although several examples of murine models for late onset disease exist, they do not show a clear homology to the underlying disease in our patient.11–13 The endoribonuclease RNase MRP consists of an RNA molecule bound to several proteins. It has at least two functions, namely, cleavage of RNA in mitochondrial DNA synthesis and nucleolar cleaving of pre-rRNA.10 A role of RNase MRP in cell cycle progression at the end of mitosis has been suggested to explain several of the CHH symptoms.16 However, the late onset appearance in our patient is difficult to reconcile with this hypothesis. A clear phenotype-genotype relationship seems absent in CHH. Variation in clinical severity is wide, even within affected families. Only heterozygosity of insertion and duplication mutations has been detected, suggesting that expression of the RNA molecule is essential for life. To date, no mutations have been found in nucleotides 23–62, which were reported to be a nucleolar localisation region.18–20 Whether the phenotype depends on the most proximal and novel C63T mutation is as yet unclear.

In our patient, blood levels of alkaline phosphatase began to decline upon total decontamination of the digestive tract, followed by a complete normalisation within the first week after allogenic BMT. Liver function has always remained normal. If indeed derived from bone, AP represents a maturation marker of osteoblasts. The interplay between osteoblasts and activated osteoclasts may have been disturbed for some time in our patient, until a change in the bacterial flora reset the increased serum levels of AP. The levels of AP remained low.

Bone marrow transplantation has been used for several bone diseases such as osteogenesis imperfecta and forms of malignant osteopetrosis.21–24 The bone dysplasia in our patient did not improve upon BMT, as has been observed before in CHH.25–27 In conclusion, the female patient with short-limb dwarfism presented here shows an unusual late onset haematological and immunological disorder. Recessive metaphyseal dysplasia without hypotrichosis (MIM 250460), a disorder presenting with short stature and metaphyseal dysplasia similar to CHH, but lacking hair anomalies, immunodeficiency and other extra skeletal features, was recently suggested to be allelic to CHH.25 In our patient, the early presentation was most compatible with kYPHOMELIC dysplasia, but later CHH became more likely, and was proven by the mutated RMRP gene. The neonatal presentation and the immunological characteristics of CHH are therefore even more variable than earlier thought.25

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