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**GASTRIC CANCER**

Comparison of cyclooxygenase 2 expression in adenocarcinomas of the gastric cardia and distal oesophagus

C J Buskens, A Sivula, B P van Rees, C Haglund, G J A Offerhaus, J J B van Lanschot, A Ristimäki

*Gut* 2003;52:1678–1683

**Background:** Adenocarcinomas of the gastric cardia and distal oesophagus are at present often considered as one clinical entity because of their comparable increasing incidence, prognosis, and optimal treatment options. However, it is still a matter of debate whether these malignancies have the same pathogenesis and genotype.

**Aims:** The aim of this study was to analyse expression of cyclooxygenase 2 (COX-2) in cardia carcinomas, and correlate this expression with clinicopathological parameters and survival. The results were compared with the prognostic value of COX-2 found for Barrett carcinomas.

**Methods:** Tumour sections of 134 consecutive patients undergoing potentially curative surgery for an adenocarcinoma of the gastric cardia and substantially invading the distal oesophagus were immunohistochemically stained using a COX-2 monoclonal antibody. Specimens were blindly scored based on intensity and extent of COX-2 immunopositivity.

**Results:** COX-2 expression was negative to weak in 59% (‘‘COX-2 low’’) and moderate to strong in 41% (‘‘COX-2 high’’) of tumours. This was significantly lower than in Barrett carcinomas (*p*<0.0001). COX-2 expression was not correlated with any clinicopathological parameter. A correlation between elevated COX-2 expression and reduced survival, as described for Barrett carcinomas, was not identified for cardiac carcinomas.

**Conclusions:** There is a difference in COX-2 expression with respect to intensity and prognostic significance between adenocarcinomas of the gastric cardia and distal oesophagus. This suggests a different pathogenesis and different genetic constitution of these two cancers. Based on these findings, the role of selective COX-2 inhibitors in the treatment of adenocarcinomas of the gastric cardia is less promising than in Barrett carcinomas.

**Abbreviations:** COX-2, cyclooxygenase 2
PATIENTS AND METHODS

Patients

Between 1 January 1993 and 31 December 2000, 306 patients underwent oesophageal resection with proximal gastrectomy for adenocarcinoma of the oesophagus, gastro-oesophageal junction, or gastric cardia (invading the distal oesophagus) with curative intent (that is, locally resectable disease without distant metastases). Data from these 306 patients were prospectively collected in a database.

A total of 151 patients (of whom six were excluded during the immunohistochemical analyses) with a distal oesophageal adenocarcinoma developed in a histologically proven Barrett’s oesophagus were analysed previously. The pathology reports of the remaining 155 patients were reviewed for the purpose of the present study. All patients were included who presented with an adenocarcinoma arising from the gastric cardia and substantially invading the distal oesophagus. The tumour was considered to be cardiac when no Barrett metaplasia was identified and when the epicentre was in the gastric cardia, defined as the area at and immediately below the gastro-oesophageal junction. Carcinomas with the epicentre of the mass located in the tubular oesophagus but without a Barrett segment were excluded, to prevent inclusion of cannibalised Barrett tumours (n = 15). Tumours arising from the fundus or the corpus of the stomach and infiltrating the gastric cardia or distal oesophagus were also excluded (n = 4), as were another two cases with an adenocarcinoma of the cardia/metaplastic oesophagus. The tumour was considered to be cardiac when no Barrett metaplasia was identified and when the epicentre was in the gastric cardia, defined as the area at and immediately below the gastro-oesophageal junction. Therefore, 134 patients remained for further analysis.

For all patients, preoperative workup consisted of endoscopy with histological biopsy, external ultrasonography of the abdomen and neck, chest x ray, endosonography, and indirect laryngoscopy. In 98 patients (73.1%) resection was performed by a transhiatal approach without thoracotomy and extended lymph node dissection. Thirty six patients (26.9%) underwent oesophagectomy through a right sided thoracotomy followed by a laparotomy in combination with two field lymph node dissection. Patients were followed until death or 1 July 2002, ensuring a minimal potential follow up of 18 months. Median actual follow up was 18 months (range 15 days to 7.8 years). They were seen on a regular basis for five years in the outpatient clinic. In the first two years patients were seen at 3–4 month intervals, and afterwards at six month intervals. For the present study, patients and/or their family practitioners were contacted by phone to assess their current status when they had been discharged by the surgeon after five years. No patient was lost to follow up.

None of the patients received chemo- and/or radiotherapy preoperatively, and no adjuvant treatment was administered postoperatively. A limited number of patients received palliative external radiotherapy for symptomatic tumour recurrence. The study was done in accordance with the guidelines of the local ethics committee.

COX-2 immunohistochemical staining

The COX-2 immunohistochemical staining procedure is described in detail elsewhere. Briefly, formalin fixed paraffin embedded specimens were sectioned (5 μm) and deparaffinised for antigen retrieval. Immunostaining was performed with a COX-2 specific mouse anti-human monoclonal antibody (160112; Cayman Chemical Co., Ann Arbor, Michigan, USA) at a dilution of 1:200. Every 20th sample of the trial series was a known colon adenocarcinoma specimen in which stromal cells at an area of ulceration were scored 3+, cancer cells from 2+ to 3+, and adjacent non-neoplastic epithelium 1+ (for scoring criteria see below). Specificity of the antibody was confirmed by restaining a randomly selected subset of specimens (every 10th sample, n = 13) with and without preadsorption of the primary antibody with a human COX-2 control peptide (10 μg/ml; Cayman Chemical) for one hour at room temperature prior to the staining procedure (that is, blocking controls).

COX-2 immunohistochemical staining was scored independently and in a blinded manner by two investigators (CB and AS). The following scoring criteria of tumour cells were agreed upon before the analysis: 0, no staining; 1+, weak diffuse cytoplasmic staining (may contain stronger intensity in less than 10% of cancer cells); 2+, moderate to strong granular cytoplasmic staining in 10–90% of cancer cells; 3+, over 90% of tumour cells stained with strong intensity. Scores 0 and 1 were categorised as “COX-2 low” and scores 2 and 3 as “COX-2 high” for the statistical analyses (see below). Allocation of tumours to the “COX-2 low” versus the “COX-2 high” category by the two investigators was similar (>90% of specimens were categorised identically). In cases of disagreement (n = 14) the slides were re-evaluated using a multi-headed microscope (CB, AS, and AR). These scoring criteria and previously described immunohistochemical control procedures are identical to those used in our previous report on COX-2 expression in oesophageal adenocarcinoma.

Statistical analysis

The association between demographic and clinicopathological features and COX-2 expression was analysed using the Student’s t test (continuous data) and the χ2 test (categorical data). Overall survival was estimated according to the Kaplan-Meier method and compared using the log rank test. The Cox proportional hazard model was used to evaluate various factors simultaneously.

A p value of <0.05 was considered statistically significant. All statistical analyses were performed using the Statistical Software Package version 9.0 (SPSS Inc., Chicago, Illinois, USA).

RESULTS

Patients

There were 113 males (84.3%) and 21 females (15.7%), with a median age of 64 years (range 39–83).

Expression of COX-2 protein in adenocarcinomas of the gastric cardia

COX-2 immunoreactivity was detected in 112 of 134 (86.7%) adenocarcinomas of the cardia whereas 22 tumours had no COX-2 expression. Moderate to strong staining (“COX-2 high”) with a granular cytoplasmic pattern was observed in 55/134 (41.0%) cases of which two were scored as strong and 53 as moderate. COX-2 expression was mainly localised in the neoplastic cells of the invading peripheral margin of the tumour. Superficial tumour cells towards the lumen were often less intense, and only weak or no staining was observed in stromal cells (connective tissue cells, smooth muscle cells, and blood vessels), except at sites of erosions and ulcerations (fig 1).

Correlation between COX-2 expression and pathological or clinical parameters

COX-2 expression was not significantly correlated with any clinicopathological parameter at the time of operation, although a possible trend was seen towards a positive association with the presence of lymph node metastases (p = 0.08) (table 1). Neither could any correlation be found between elevated COX-2 expression and the development of distant metastases or locoregional recurrences during follow up (table 1).

Kaplan-Meier curves for patient survival are depicted in fig 2A. As can be seen in the survival curves, there was no significant difference in survival between patients in the
Figure 1  Representative examples of cyclooxygenase 2 (COX-2) immunohistochemistry. (A) Weak (1+) immunoreactivity in tumour cells. This tumour was categorised as “COX-2 low” (200×). In the lower right corner, the blocking control is shown (see materials and methods for details). (B) Moderate (2+) immunoreactivity in tumour cells. This tumour was categorised as “COX-2 high” (200×). In the lower right corner the blocking control is shown. (C) Immunoreactivity was stronger in the invading peripheral margin of the tumour.

Table 1  Correlation of clinicopathological findings and clinical outcome parameters with cyclooxygenase 2 (COX-2) expression

<table>
<thead>
<tr>
<th>COX-2 expression</th>
<th>Low (n = 79) (n [%])</th>
<th>High (n = 55) (n [%])</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Patient characteristics</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age (mean (SD)) (y)</td>
<td>64 (10)</td>
<td>62 (9)</td>
<td>0.3</td>
</tr>
<tr>
<td>Sex</td>
<td>Male 70 (88.6)</td>
<td>43 (78.2)</td>
<td>0.1</td>
</tr>
<tr>
<td>Female</td>
<td>9 (11.4)</td>
<td>12 (21.8)</td>
<td></td>
</tr>
<tr>
<td><strong>Tumour characteristics</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Depth of invasion*</td>
<td>T1 4 (5.1)</td>
<td>1 (1.8)</td>
<td>0.7</td>
</tr>
<tr>
<td>T2</td>
<td>6 (7.6)</td>
<td>3 (5.5)</td>
<td></td>
</tr>
<tr>
<td>T3</td>
<td>64 (81.0)</td>
<td>48 (87.3)</td>
<td></td>
</tr>
<tr>
<td>T4</td>
<td>5 (6.3)</td>
<td>3 (5.5)</td>
<td></td>
</tr>
<tr>
<td>Lymph node involvement†</td>
<td>N0 17 (21.5)</td>
<td>9 (16.4)</td>
<td>0.08</td>
</tr>
<tr>
<td>N1</td>
<td>37 (46.8)</td>
<td>27 (49.1)</td>
<td></td>
</tr>
<tr>
<td>N2</td>
<td>24 (30.4)</td>
<td>13 (23.6)</td>
<td></td>
</tr>
<tr>
<td>N3</td>
<td>1 (1.3)</td>
<td>6 (10.9)</td>
<td></td>
</tr>
<tr>
<td>Distant metastasis</td>
<td>M0 75 (94.9)</td>
<td>53 (94.6)</td>
<td>0.5</td>
</tr>
<tr>
<td>M1</td>
<td>4 (5.1)</td>
<td>2 (3.6)</td>
<td></td>
</tr>
<tr>
<td><strong>Differentiation grade</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Well</td>
<td>2 (2.5)</td>
<td>1 (1.8)</td>
<td>0.7</td>
</tr>
<tr>
<td>Moderate</td>
<td>29 (36.7)</td>
<td>17 (30.9)</td>
<td></td>
</tr>
<tr>
<td>Poor</td>
<td>48 (60.4)</td>
<td>37 (67.5)</td>
<td></td>
</tr>
<tr>
<td><strong>Tumour stage‡</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ia</td>
<td>3 (3.4)</td>
<td>0 (0.0)</td>
<td>0.7</td>
</tr>
<tr>
<td>Ib</td>
<td>1 (1.3)</td>
<td>2 (3.6)</td>
<td></td>
</tr>
<tr>
<td>II</td>
<td>16 (20.3)</td>
<td>10 (18.2)</td>
<td></td>
</tr>
<tr>
<td>IIIa</td>
<td>32 (40.5)</td>
<td>25 (45.5)</td>
<td></td>
</tr>
<tr>
<td>IIIb</td>
<td>23 (29.1)</td>
<td>16 (29.1)</td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>4 (5.1)</td>
<td>3 (6.6)</td>
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</tr>
<tr>
<td><strong>Operation type§</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>THE</td>
<td>58 (73.4)</td>
<td>40 (72.7)</td>
<td>0.5</td>
</tr>
<tr>
<td>TTE</td>
<td>21 (26.6)</td>
<td>15 (27.3)</td>
<td></td>
</tr>
<tr>
<td><strong>Radicality of resection¶</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>R0</td>
<td>58 (73.4)</td>
<td>40 (72.7)</td>
<td>0.5</td>
</tr>
<tr>
<td>R1</td>
<td>21 (26.6)</td>
<td>14 (25.5)</td>
<td></td>
</tr>
<tr>
<td>R2</td>
<td>0 (0.0)</td>
<td>1 (1.8)</td>
<td></td>
</tr>
<tr>
<td><strong>Clinical outcome</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Locoregional recurrence</td>
<td>No 41 (51.9)</td>
<td>31 (56.4)</td>
<td>0.6</td>
</tr>
<tr>
<td>Yes</td>
<td>38 (48.1)</td>
<td>24 (43.6)</td>
<td></td>
</tr>
<tr>
<td>Distant metastasis</td>
<td>No 45 (57.0)</td>
<td>31 (56.4)</td>
<td>0.9</td>
</tr>
<tr>
<td>Yes</td>
<td>34 (43.0)</td>
<td>24 (43.6)</td>
<td></td>
</tr>
</tbody>
</table>

* T1, tumour limited to the submucosa; T2, tumour infiltrates muscularis propia, but not adventitia; T3, tumour infiltrates adventitia; T4, tumour infiltrates adjacent structures.
† N0, no lymph node metastasis; N1, 1–6 lymph node metastases; N2, 7–15 lymph node metastases; N3, more than 15 lymph node metastases.
‡ T1N0M0, Ib; T1N1M0, T2N0M0, II, T1N2-3M0; T2N1M0, T3N0M0, IIa, T2N2-3M0, T3N1M0, IIIb, T3N2-3M0, T4N1M0, IV, T4N2-3M0, anyTanyN1: 1 THE, transthoracic resection; TTE, transthoracic resection.
§ R0, microscopically radical; R1, microscopically non-radical; R2, macroscopically non-radical.
COX-2 expression in cardia carcinoma

“COX-2 low” category compared with the “COX-2 high” category, with a median survival of 22 months (95% confidence interval (CI) 13–31) and 25 months (95% CI 8–42), respectively (p = 0.50; log rank test). Subgroup analysis of the prognostic value of COX-2 per tumour stage did not show any significant difference in survival. Because this might be due to the low numbers of patients available per tumour stage, tumour stage was recoded into a dichotomous variable by combining categories with comparable prognosis (tumour stages Ia, Ib, II v IIIa, IIIb, IV). There was also no significant survival advantage for patients with “COX-2 low” expression in either group, although there was a slight indication that elevated COX-2 expression was a predictor for poor outcome in relatively early cancers (p = 0.17) (fig 2B).

Comparison with oesophageal Barrett carcinoma

The finding that COX-2 expression was not a prognostic variable for cardia carcinomas is different from results of distal oesophageal adenocarcinomas in which it was shown to be an independent prognostic variable together with tumour stage and radicality of resection. Comparison of the COX-2 staining results showed significantly less intense COX-2 staining in cardia carcinomas compared with Barrett carcinomas (p = 0.0001) (table 2).

Overall, there was no significant difference in survival between patients with a gastric cardia carcinoma and patients with a Barrett carcinoma (p = 0.22; log rank test), although a trend was seen towards a more favourable outcome for patients with a Barrett carcinoma (median 25 months (95% CI 18–33) v median 38 months (95% CI 16–60), respectively). This can be explained, at least in part, by a more favourable T and N stage in patients with a Barrett carcinoma (p<0.001).

DISCUSSION

This study shows that elevated COX-2 expression is present in 41% of patients (moderate to strong) with a cardia carcinoma. However, it did not correlate with prognosis. This finding is in contrast with our previous study in which we demonstrated that elevated COX-2 expression was an independent prognostic variable for Barrett carcinomas, and is surprising in view of the epidemiological and clinical similarities (that is, rapidly rising incidence, stage by stage prognosis, and optimal surgical treatment) shared by both distal oesophageal and gastric cardia adenocarcinomas. However, it is consistent with the results of a large epidemiological study in which a substantial protective effect of non-steroidal anti-inflammatory drugs was observed for adenocarcinomas arising at distal gastric or oesophageal sites but not for cardia carcinomas.

A possible explanation for the discrepancy in prognostic relevance of COX-2 that we identified between adenocarcinomas of the gastric cardia and distal oesophagus may be a difference in carcinogenesis for these two cancers. Apart from the significant difference in the intensity of COX-2 expression in carcinoma cells, there was a discrepancy in the pattern of expression. In cardia carcinomas, the strongest (heterogeneous) COX-2 expression was seen in the invading peripheral margin of the tumour, in contrast with the homogeneous and predominantly superficial luminal expression of COX-2 seen in Barrett carcinoma. This suggests that in cardia carcinomas, COX-2 upregulation is a relatively late event and occurs in invasive malignant cells that are often already infiltrating the lymphatic system or blood vessels, which could explain why COX-2 expression is not a prognostic factor.

This difference in COX-2 expression with respect to intensity, localisation, and prognostic significance is suggestive of a different pathogenesis and different genetic constitution of these two adenocarcinomas. To date, the strongest supportive evidence that Barrett and cardia carcinomas represent two separate clinical entities has come from an epidemiological study identifying symptomatic reflux as a strong risk factor for oesophageal adenocarcinomas, and only a relatively weak risk factor for adenocarcinomas of the gastric cardia.

Other evidence indicating that these tumours should be regarded as two different clinical entities comes from recent reports comparing intestinal metaplasia of the gastric cardia with that of the distal oesophagus. Intestinal metaplasia of the cardia is predominantly of the complete type and is associated with pathological features of the stomach, especially pangastritis. This is in contrast with the incomplete type of intestinal metaplasia in the distal oesophagus (that is, Barrett mucosa), which is characterised by the presence of goblet cells and non-secretory columnar cells and carries an increased risk of
dysplasia or cancer. Adenocarcinomas of the cardia were also reported to have a different oncogenic profile compared with distal oesophageal carcinomas. In particular, the prevalence of p53 mutations in cardia carcinomas is less than 50% (31–42%) whereas for oesophageal carcinomas a p53 mutation is the most frequent alteration identified (75–100%). As p53 mutations induce COX-2 transcription in vitro by disrupting binding of p53 to the promoter region of COX-2, this could be an explanation for the less pronounced COX-2 upregulation in cardia carcinomas in comparison with Barrett carcinomas.

Another explanation for the difference in prognostic value of COX-2 for adenocarcinomas of the cardia and adenocarcinomas of the distal oesophagus arising in a Barrett segment might be that elevated expression of COX-2 is only a prognostic variable for poor prognosis in relatively early carcinomas. In lung cancer, upregulation of COX-2 was demonstrated to be associated with worse overall survival in patients with stage I non-small cell lung cancer. This would be in line with the prognostic significance of COX-2 expression for patients with Barrett carcinomas who had significantly more favourable T and N stages than patients with cardia carcinomas. This is also supported by the finding that there seemed to be a trend for a prognostic role for COX-2 in early carcinomas of the gastric cardia compared with more advanced tumour stages.

In this report, we analysed the clinical significance of elevated COX-2 expression in adenocarcinomas of the gastric cardia. We could not demonstrate that COX-2 expression in epithelial tumour cells was related to stage or poorer long term outcome for patients with adenocarcinoma of the gastric cardia, which is in contrast with our previous results in Barrett carcinomas where COX-2 expression was an independent prognostic variable for patient survival. These results support the hypothesis that although these cancers might be considered as one clinical entity, they are two distinct pathological entities.

There are several ongoing chemoprevention and adjuvant chemotherapy trials with selective COX-2 inhibitors for gastrointestinal malignancies. Although the role of selective COX-2 inhibitors in the prevention and treatment of gastrointestinal cancer is promising, further investigations are needed before they can be incorporated into daily clinical practice. This study demonstrates that different tumours may require different treatment strategies. Therefore, it is important to gain further insight in the mechanisms by which COX-2 contributes to the carcinogenesis of various cancers. To date, these findings make the role of selective COX-2 inhibitors in the treatment of adenocarcinomas of the gastric cardia less promising than in Barrett cancers.

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Table 2
Comparison of cyclooxygenase 2 (COX-2) staining between adenocarcinomas of the gastric cardia (present study) and adenocarcinomas of the distal oesophagus developed in a Barrett segment (Buskens and colleagues)

<table>
<thead>
<tr>
<th>COX-2 expression</th>
<th>Cardia (n = 134) (n [%])</th>
<th>Oesophagus (n = 145) (n [%])</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>0 (24)</td>
<td>22 (16.4)</td>
<td>2 (1.4)</td>
<td>0.0001</td>
</tr>
<tr>
<td>1 (85)</td>
<td>57 (42.5)</td>
<td>28 (19.3)</td>
<td></td>
</tr>
<tr>
<td>2 (155)</td>
<td>53 (39.6)</td>
<td>102 (70.3)</td>
<td></td>
</tr>
<tr>
<td>3 (15)</td>
<td>2 (1.5)</td>
<td>13 (9.0)</td>
<td></td>
</tr>
</tbody>
</table>

In cardia carcinomas, significantly less (intense) COX-2 staining was found (p=0.0001).

REFERENCES
Cyclooxygenase 2 expression in cardia carcinoma