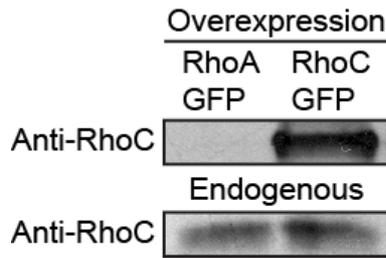


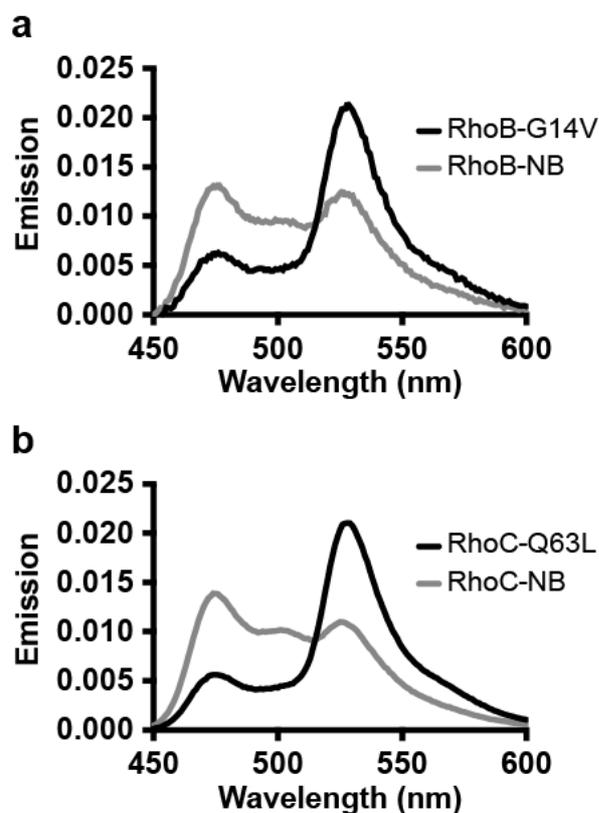
Supplementary materials:

Spatiotemporal analysis of RhoA/B/C activation in primary human endothelial cells

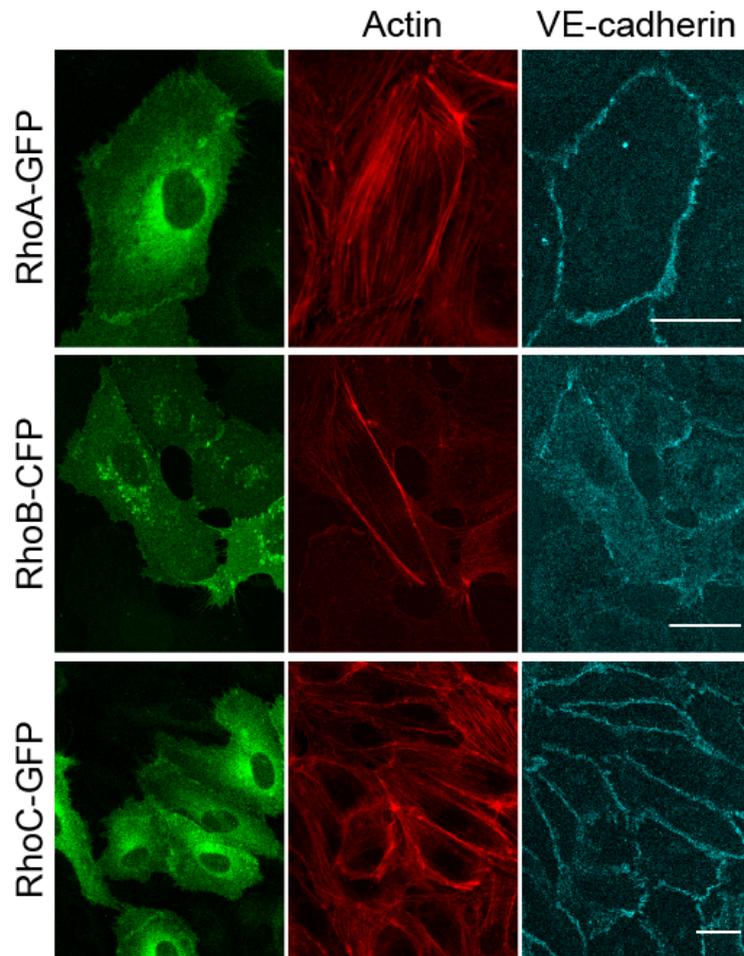
Nathalie R. Reinhard^{1,2}, Suzanne F. van Helden², Eloise C. Anthony², Taofei Yin³, Yi Wu³,
Joachim Goedhart¹, Theodorus W.J. Gadella Jr.¹, Peter L. Hordijk^{1,2,*} †



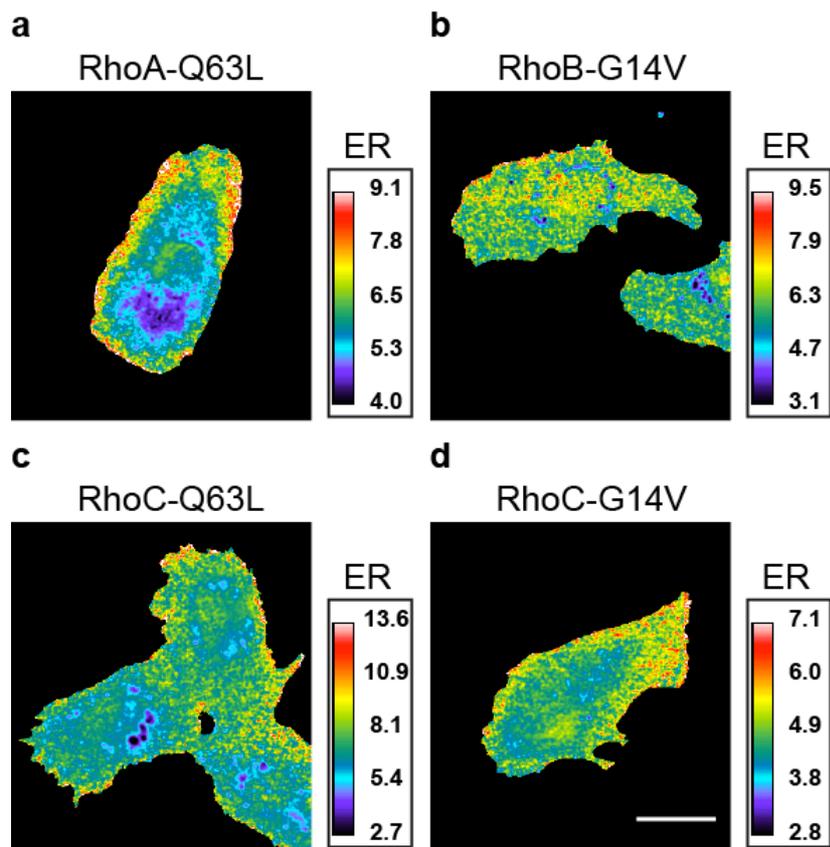
Supplemental Figure S1: RhoC antibody specifically detects RhoC and not RhoA. ECs were transfected with RhoA-GFP or RhoC-GFP and a RhoC antibody was used to check for its specificity towards RhoA and RhoC on Western blot. Endogenous RhoC detection in RhoA-GFP and RhoC-GFP positive cell lysates was included to demonstrate that endogenous RhoC was detected in both conditions.



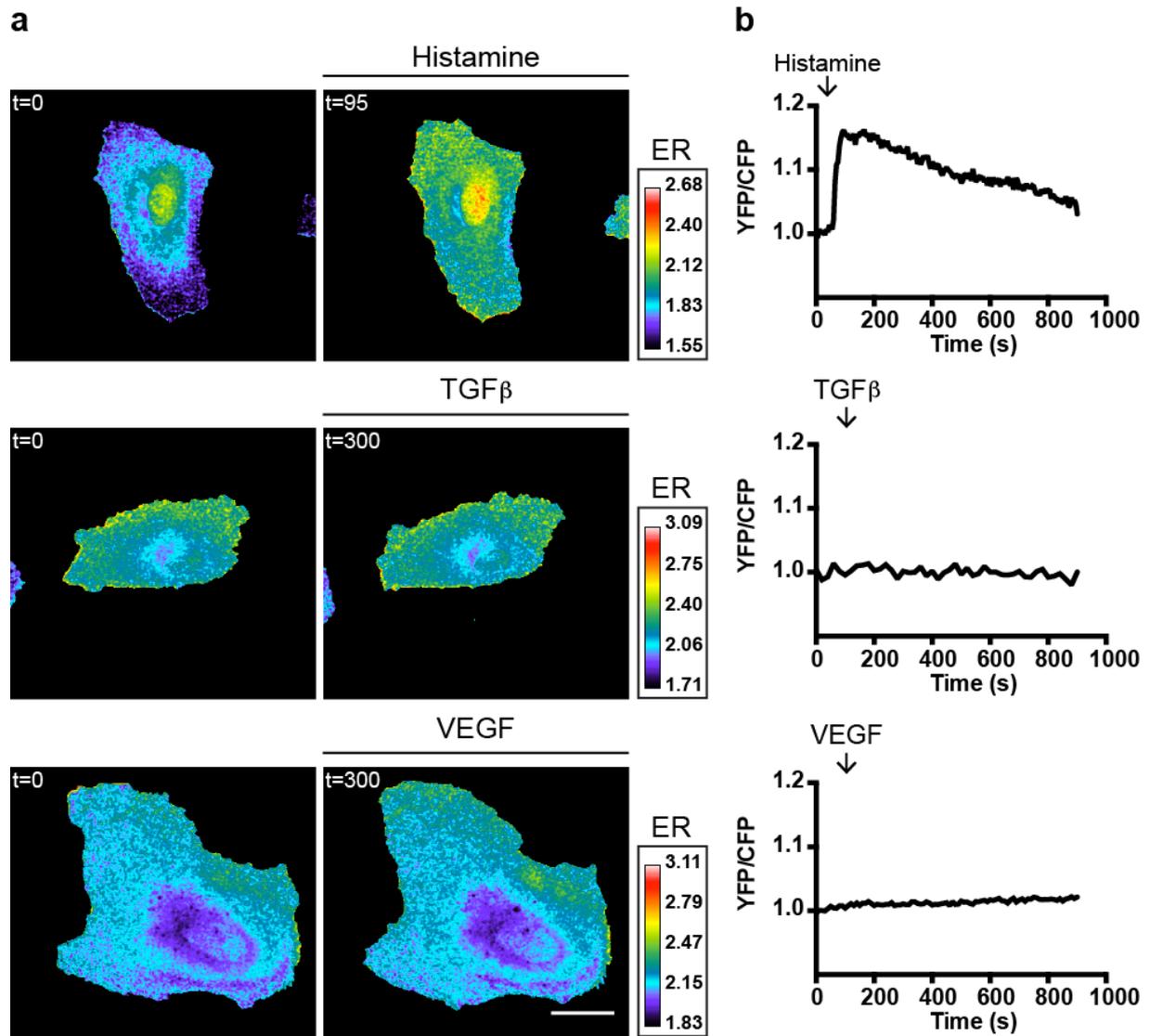
Supplemental Figure S2: Emission spectra of RhoB and RhoC FRET sensors. (a) Normalized emission spectra of the RhoB-G14V (black trace) and RhoB-DN (grey trace) measured in transiently transfected HEK293 cells. Spectra were normalized to the area under the curve. (b) Normalized emission spectra of the RhoC-Q63L (black trace) and RhoC-DN (grey trace) measured in transiently transfected HEK293 cells. Spectra were normalized to the area under the curve.



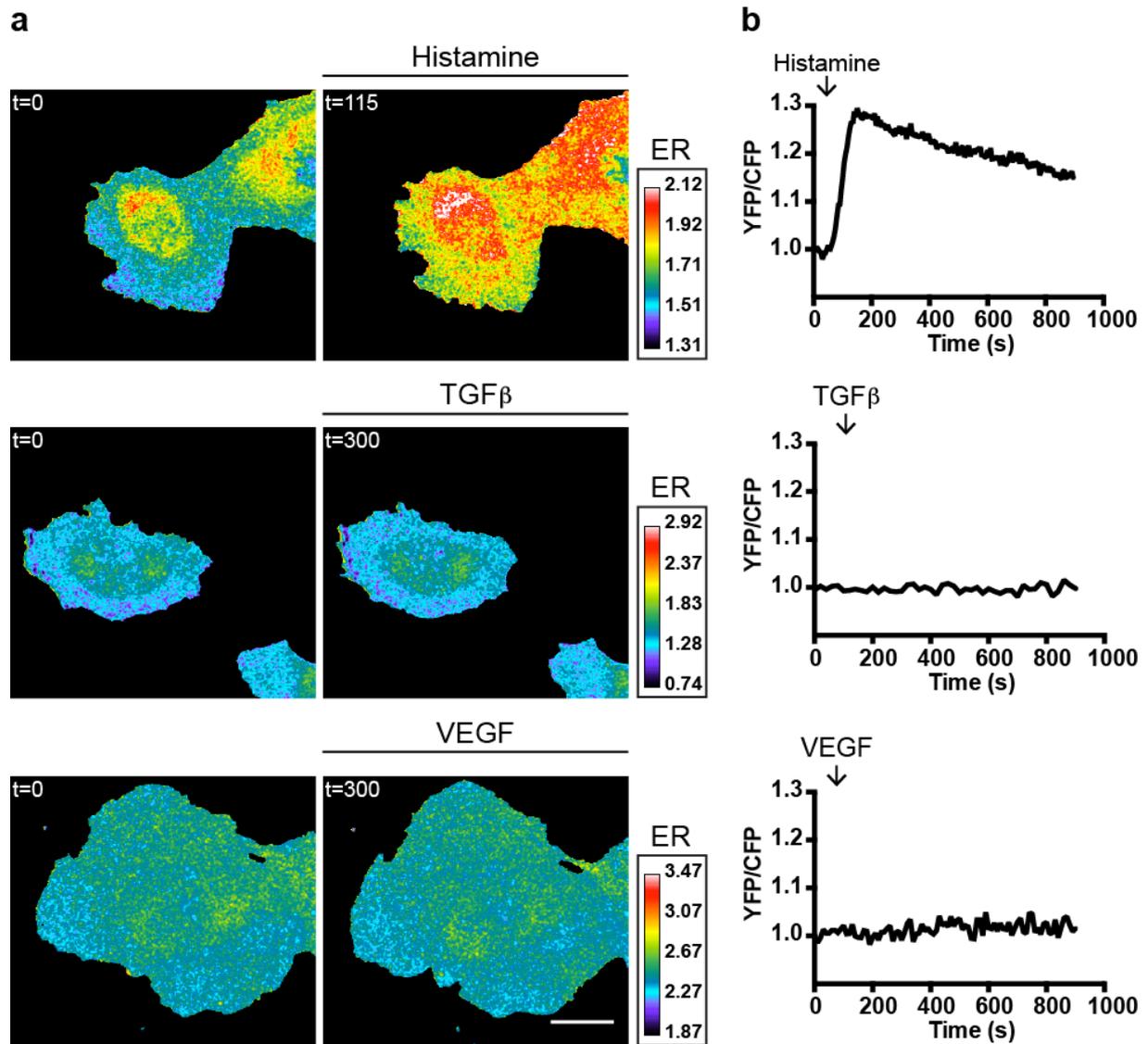
Supplemental Figure S3: Localization of FP-tagged RhoA/B/C constructs. ECs were transfected with RhoA/C-GFP or RhoB-CFP and co-stained for F-actin and VE-cadherin. Scale bar = 25 μ m.



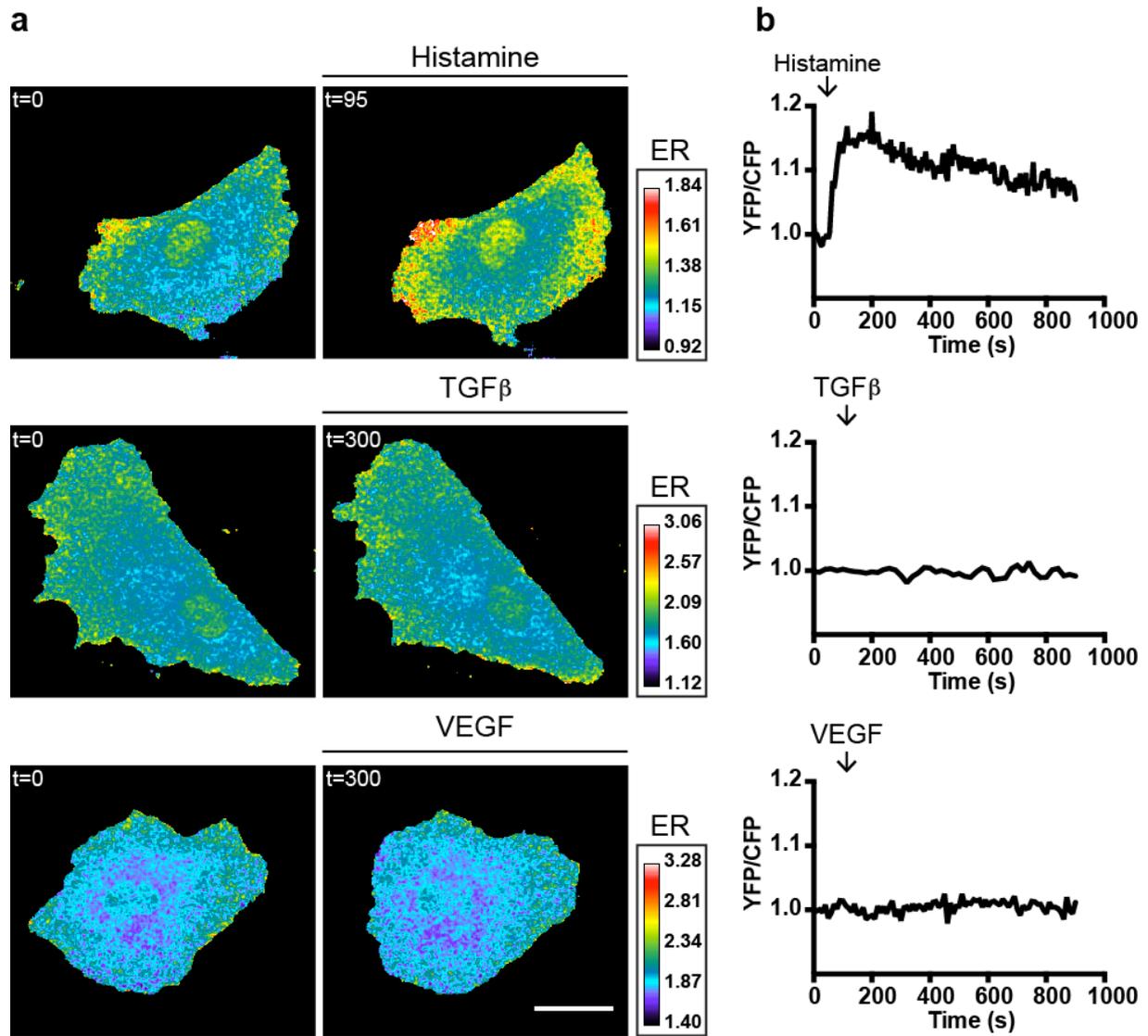
Supplemental Figure S4: Dominant positive RhoA/B/C FRET sensors. (a-d) Ratiometric images of ECs that were transfected with the (a) RhoA-Q63L- (b) RhoB-G14V- (c) RhoC-Q63L- or (d) RhoC-G14V FRET sensors. Warm colors represent high YFP/CFP ratios (Emission ratio (ER) on the right). Scale bar = 25 μ m.



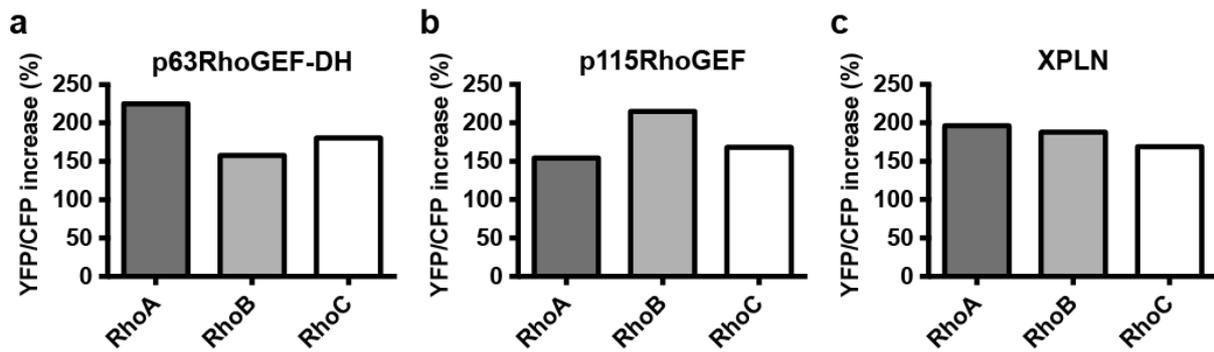
Supplemental Figure S5: Histamine, but not TGF β or VEGF, induces RhoA activation in human ECs. (a) Ratiometric images of ECs that were transfected with the RhoA FRET sensor and stimulated with either Histamine (100 μ M) at t=55 sec, VEGF (50ng/ml) at t=110 sec, or TGF β (10 ng/ml) at t=100 sec. Warm colors represent high YFP/CFP ratios (Emission ratio (ER) on the right). Scale bar=25 μ m. (b) Normalized mean YFP/CFP ratio changes of the cells in (a), which are representative examples of 2-7 cells.



Supplemental Figure S6: Histamine, but not TGF β or VEGF, induces RhoB activation in human ECs. (a) Ratiometric images of ECs that were transfected with the RhoB FRET sensor and stimulated with either Histamine (100 μ M) at $t=55$ sec, VEGF (50ng/ml) at $t=110$ sec, or TGF β (10 ng/ml) at $t=100$ sec. Warm colors represent high YFP/CFP ratios (Emission ratio (ER) on the right). Scale bar=25 μ m. (b) Normalized mean YFP/CFP ratio changes of the cells in (a), which are representative examples of 2-7 cells.



Supplemental Figure S7: Histamine, but not TGF β or VEGF, induces RhoC activation in human ECs. (a) Ratiometric images of ECs that were transfected with the RhoC FRET sensor and stimulated with either Histamine (100 μ M) at t=55 sec, VEGF (50ng/ml) at t=110 sec, or TGF β (10 ng/ml) at t=100 sec. Warm colors represent high YFP/CFP ratios (Emission ratio (ER) on the right). Scale bar=25 μ m. (b) Normalized mean YFP/CFP ratio changes of the cells in (a), which are representative examples of 2-7 cells.



Supplemental Figure S8: RhoGEF efficiencies towards RhoA/B/C. (a,b,c) Mean YFP/CFP ratios of RhoA/B/C FRET sensors with RhoGEF overexpression were divided by the mean YFP/CFP ratios of RhoA/B/C FRET sensors without RhoGEF overexpression to illustrate GEF efficiencies of (a) p63RhoGEF-DH (b) p115RhoGEF and (c) XPLN towards the different RhoA/B/C-WT FRET sensors. Mean values, used for the calculation, were obtained from Fig. 7a-c.

Supplemental Movie M1:

Thrombin induces RhoA/B/C activation

Primary HUVEC transfected with the RhoA (left panel); RhoB (middle panel); RhoC (right panel); FRET sensors were stimulated with 1U/ml Thrombin at t=110 sec. See also Fig. 3.

Supplemental Movie M1:

Nocodazole induces RhoA/B/C activation

Primary HUVEC transfected with the RhoA (left panel); RhoB (middle panel); RhoC (right panel); FRET sensors were stimulated with 5 μ M nocodazole to induce microtubule depolymerization at t=180 sec. See also Fig. 4.