



UvA-DARE (Digital Academic Repository)

Effects of rising CO₂ on the harmful cyanobacterium *Microcystis*

Sandrini, G.

Publication date

2016

Document Version

Final published version

[Link to publication](#)

Citation for published version (APA):

Sandrini, G. (2016). *Effects of rising CO₂ on the harmful cyanobacterium *Microcystis**. [Thesis, fully internal, Universiteit van Amsterdam].

General rights

It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations

If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: <https://uba.uva.nl/en/contact>, or a letter to: Library of the University of Amsterdam, Secretariat, P.O. Box 19185, 1000 GD Amsterdam, The Netherlands. You will be contacted as soon as possible.



References

- Allahverdiyeva Y, Ermakova M, Eisenhut M, Zhang P, Richaud P, Hagemann M, Cournac L, Aro EM. (2011). Interplay between flavodiiron proteins and photorespiration in *Synechocystis* sp. PCC 6803. *J Biol Chem* **286**: 24007–24014.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. (1990). Basic local alignment search tool. *J Mol Biol* **215**: 403–410.
- Anderson DM, Glibert PM, Burkholder JM. (2002). Harmful algal blooms and eutrophication: nutrient sources, composition, and consequences. *Estuaries* **25**: 704–726.
- Bade DL, Cole JJ. (2006). Impact of chemically enhanced diffusion on dissolved inorganic carbon stable isotopes in a fertilized lake. *J Geophys Res* **111**: C01014.
- Badger MR, Palmqvist K, Yu JW. (1994). Measurement of CO₂ and HCO₃⁻ fluxes in cyanobacteria and microalgae during steady-state photosynthesis. *Physiol Plant* **90**: 529–536.
- Badger MR, Price GD, Long BM, Woodger FJ. (2006). The environmental plasticity and ecological genomics of the cyanobacterial CO₂ concentrating mechanism. *J Exp Bot* **57**: 249–265.
- Balmer MB, Downing JA. (2011). Carbon dioxide concentrations in eutrophic lakes: undersaturation implies atmospheric uptake. *Inland Waters* **1**: 125–132.
- Bañares-España E, López-Rodas V, Salgado C, Costas E, Flores-Moya A. (2006). Inter-strain variability in the photosynthetic use of inorganic carbon, exemplified by the pH compensation point, in the cyanobacterium *Microcystis aeruginosa*. *Aquat Bot* **85**: 159–162.
- Barica J, Kling H, Gibson J. (1980). Experimental manipulation of algal bloom composition by nitrogen addition. *Can J Fish Aquat Sci* **37**: 1175–1183.
- Beaulieu M, Pick F, Gregory-Eaves I. (2013). Nutrients and water temperature are significant predictors of cyanobacterial biomass in a 1147 lakes data set. *Limnol Oceanogr* **58**: 1736–1746.
- Benjamini Y, Yekutieli D. (2001). The control of the false discovery rate in multiple testing under dependence. *Ann Statist* **29**: 1165–1188.
- Benschop JJ, Badger MR, Price GD. (2003). Characterisation of CO₂ and HCO₃⁻ uptake in the cyanobacterium *Synechocystis* sp. PCC6803. *Photosynth Res* **77**: 117–126.
- Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Lipman DJ, Ostell J, Sayers EW. (2013). GenBank. *Nucleic Acids Res* **41**: D36–42.
- Berner, RA. (1998) The carbon cycle and CO₂ over Phanerozoic time: the role of land plants. *Phil Trans R Soc Lond B Biol Sci* **353**: 75–81.
- Berry S, Esper B, Karandashova I, Teuber M, Elanskaya I, Rögner M, Hagemann M. (2013). Potassium uptake in the unicellular cyanobacterium *Synechocystis* sp. strain PCC 6803 mainly depends on a Ktr-like system encoded by *slr1509 (ntpJ)*. *FEBS Lett* **548**: 53–58.
- Bersanini L, Battchikova N, Jokel M, Rehman A, Vass I, Allahverdiyeva Y, Aro EM. (2014). Flavodiiron protein Flv2/Flv4-related photoprotective mechanism dissipates excitation pressure of PSII in cooperation with phycobilisomes in cyanobacteria. *Plant Physiol* **164**: 805–818.
- Binladen J, Gilbert MT, Bollback JP, Panitz F, Bendixen C, Nielsen R, Willerslev E. (2007). The use of coded PCR primers enables high-throughput sequencing of multiple homolog amplification products by 454 parallel sequencing. *PLoS One* **2**: e197.
- Bolstad BM, Irizarry RA, Åstrand M, Speed TP. (2003). A comparison of normalization methods for high density oligonucleotide array data based on variance and bias. *Bioinformatics* **19**: 185–193.
- Boopathi T, Ki JS. (2014). Impact of environmental factors on the regulation of cyanotoxin production. *Toxins* **6**: 1951–1978.
- Bradford MM. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* **72**: 248–254.

- Brauer VS, Stomp M, Huisman J. (2012). The nutrient-load hypothesis: patterns of resource limitation and community structure driven by competition for nutrients and light. *Am Nat* **179**: 721–740.
- Brauer VS, Stomp M, Rosso C, Van Beusekom SAM, Emmerich B, Stal LJ, Huisman J. (2013). Low temperature delays timing and enhances the cost of nitrogen fixation in the unicellular cyanobacterium *Cyanothece*. *ISME J* **7**: 2105–2115.
- Buckling A, Maclean RC, Brockhurst MA, Colegrave N. (2009). The Beagle in a bottle. *Nature* **457**: 824–829.
- Burnap RL, Hagemann M, Kaplan A. (2015). Regulation of CO₂ concentrating mechanism in cyanobacteria. *Life* **5**: 348–371.
- Burnap RL, Nambudiri R, Holland S. (2013). Regulation of the carbon-concentrating mechanism in the cyanobacterium *Synechocystis* sp. PCC6803 in response to changing light intensity and inorganic carbon availability. *Photosynth Res* **118**: 115–124.
- Butterwick C, Heaney SI, Talling JF. (2005). Diversity in the influence of temperature on the growth rates of freshwater algae, and its ecological relevance. *Freshw Biol* **50**: 291–300.
- Campbell D, Hurry V, Clarke AK, Gustafsson P, Öquist G. (1998). Chlorophyll fluorescence analysis of cyanobacterial photosynthesis and acclimation. *Microbiol Mol Biol Rev* **62**: 667–683.
- Cao H, Shimura Y, Masanobu K, Yin Y. (2014). Draft genome sequence of the toxic bloom-forming cyanobacterium *Aphanizomenon flos-aquae* NIES-81. *Genome Announc* **2**: e00044-14.
- Caraco NF, Miller R. (1998). Effects of CO₂ on competition between a cyanobacterium and eukaryotic phytoplankton. *Can J Fish Aquat Sci* **55**: 54–62.
- Carey CC, Ibelings BW, Hoffmann EP, Hamilton DP, Brookes JD. (2012). Eco-physiological adaptations that favour freshwater cyanobacteria in a changing climate. *Water Res* **46**: 1394–1407.
- Carmichael WW. (2001). Health effects of toxin producing cyanobacteria: the ‘CyanoHABS’. *Hum Ecol Risk Assess* **7**: 1393–1407.
- Chaturvedi P, Agrawal MK, Bagchi SN. (2015). Microcystin-producing and non-producing cyanobacterial blooms collected from the Central India harbor potentially pathogenic *Vibrio cholerae*. *Ecotoxicol Environ Saf* **115**: 67–74.
- Chen YW, Qin BQ, Teubner K, Dokulil MT. (2003). Long-term dynamics of phytoplankton assemblages: *Microcystis*-domination in Lake Taihu, a large shallow lake in China. *J Plankton Res* **25**: 445–453.
- Chorus I, Bartram J. (1999). *Toxic Cyanobacteria in Water: A Guide to their Public Health Consequences, Monitoring and Management*. London, UK: E&FN Spon.
- Chorus I, Falconer IR, Salas HJ, Bartram J. (2000). Health risks caused by freshwater cyanobacteria in recreational waters. *J Toxicol Environ Health B: Crit Rev* **3**: 323–347.
- Christiansen G, Goesmann A, Kurmayer R. (2014). Elucidation of insertion elements carried on plasmids and *in vitro* construction of shuttle vectors from the toxic cyanobacterium *Planktothrix*. *Appl Environ Microbiol* **80**: 4887–4897.
- Civitello DJ, Hite JL, Hall SR. (2014). Potassium enrichment stimulates the growth and reproduction of a clone of *Daphnia dentifera*. *Oecologia* **175**: 773–780.
- Clapham, ME. (2013). Extinction: End-Permian Mass Extinction. In: eLS. Chichester, UK: John Wiley & Sons.
- Codd GA, Morisson LF, Metcalf JS. (2005). Cyanobacterial toxins: risk management for health protection. *Toxicol Appl Pharmacol* **203**: 264–272.
- Cole JJ, Bade DL, Bastviken D, Pace ML, Van de Bogert M. (2010). Multiple approaches to estimating air-water gas exchange in small lakes. *Limnol Oceanogr Methods* **8**: 285–293.
- Cole JJ, Caraco NF. (2001). Carbon in catchments: connecting terrestrial carbon losses with aquatic metabolism. *Mar Freshw Res* **52**: 101–110.

- Cole JJ, Caraco NF, Kling GW, Kratz TK. (1994). Carbon dioxide supersaturation in the surface waters of lakes. *Science* **265**: 1568–1570.
- Collins S, Bell G. (2004). Phenotypic consequences of 1,000 generations of selection at elevated CO₂ in a green alga. *Nature* **431**: 566–569.
- Collins S, Sültemeyer D, Bell G. (2006). Changes in C uptake in populations of *Chlamydomonas reinhardtii* selected at high CO₂. *Plant Cell Environ* **29**: 1812–1819.
- Conley, DJ, Paerl HW, Howarth RW, Boesch DF, Seitzinger SP, Havens KE, Lancelot C, Likens GE. (2009). Controlling eutrophication: nitrogen and phosphorus. *Science* **323**: 1014–1015.
- Cox PA, Banack SA, Murch SJ. (2003). Biomagnification of cyanobacterial neurotoxins and neurodegenerative disease among the Chamorro people of Guam. *Proc Natl Acad Sci USA* **100**: 13380–13383.
- Crusius J, Wanninkhof R. (2003). Gas transfer velocities measured at low wind speed over a lake. *Limnol Oceanogr* **48**: 1010–1017.
- Cuypers Y, Vinçon-Leite B, Groleau A, Tassin B, Humbert JF. (2011). Impact of internal waves on the spatial distribution of *Planktothrix rubescens* (cyanobacteria) in an alpine lake. *ISME J* **5**: 580–589.
- Czerny J, Barcelo e Ramons J, Riebesell U. (2009). Influence of elevated CO₂ concentrations on cell division and nitrogen fixation rates in the bloom-forming cyanobacterium *Nodularia spumigena*. *Biogeosciences* **6**: 1865–1875.
- Daley SM, Kappell AD, Carrick MJ, Burnap RL. (2012). Regulation of the cyanobacterial CO₂-concentrating mechanism involves internal sensing of NADP⁺ and α -ketoglutarate levels by transcription factor CcmR. *PLoS One* **7**: e41286.
- Davis TW, Berry DL, Boyer GL, Gobler CJ. (2009). The effects of temperature and nutrients on the growth and dynamics of toxic and non-toxic strains of *Microcystis* during cyanobacteria blooms. *Harmful Algae* **8**: 715–725.
- Dittmann E, Fewer DP, Neilan BA. (2013). Cyanobacterial toxins: biosynthetic routes and evolutionary roots. *FEMS Microbiol Rev* **37**: 23–43.
- Dittmann E, Neilan BA, Erhard M, von Döhren H, Börner T. (1997). Insertional mutagenesis of a peptide synthetase gene that is responsible for hepatotoxin production in the cyanobacterium *Microcystis aeruginosa* PCC 7806. *Mol Microbiol* **26**: 779–787.
- Dodds WK, Bouska WW, Eitzmann JL, Pilger TJ, Pitts KL, Riley AJ, Schloesser JT, Thornbrugh DJ. (2008). Eutrophication of US freshwaters: analysis of potential economic damages. *Environ Sci Technol* **43**: 12–19.
- Dokulil MT, Teubner K. (2000). Cyanobacterial dominance in lakes. *Hydrobiol* **438**: 1–12.
- Doney SC, Fabry VJ, Feely RA, Kleypas JA. (2009). Ocean acidification: the other CO₂ problem. *Annu Rev Mar Sci* **1**: 169–192.
- Downing JA, Watson SB, McCauley E. (2001). Predicting cyanobacteria dominance in lakes. *Can J Fish Aquat Sci* **58**: 1905–1908.
- Du J, Förster B, Rourke L, Howitt SM, Price GD. (2014). Characterisation of cyanobacterial bicarbonate transporters in *E. coli* shows that SbtA homologs are functional in this heterologous expression system. *PLoS One* **9**: e115905.
- Dubois M, Giles KA, Hamilton JK, Rebers PA, Smith F. (1956). Colorimetric method for determination of sugars and related substances. *Anal Chem* **28**: 350–356.
- Dykhuizen DE, Hartl DL. (1983). Selection in chemostats. *Microbiol Rev* **47**: 150.
- Dziallas C, Pinnow S, Grossart HP. (2011). Quantification of toxic and toxinproducing cyanobacterial cells by RING-FISH in combination with flow cytometry. *Limnol Oceanogr Methods* **9**: 67–73.
- Dziallas C, Grossart H. (2011). Increasing oxygen radicals and water temperature select for toxic *Microcystis* sp. *PLoS One* **6**: 25569.

- Dzialowski AR, Wang SH, Lim NC, Spotts WW, Huggins DG. (2005). Nutrient limitation of phytoplankton growth in central plains reservoirs, USA. *J Plankton Res* **27**: 587–595.
- Eichner M, Rost B, Kranz SA. (2014). Diversity of ocean acidification effects on marine N₂ fixers. *J Exp Mar Biol Ecol* **457**: 199–207.
- Eichner M, Thoms S, Kranz SA, Rost B. (2015). Cellular inorganic carbon fluxes in *Trichodesmium*: a combined approach using measurements and modelling. *J Exp Bot* **66**: 749–759.
- Eisenhut M, Aguirre von Wobeser E, Jonas L, Schubert H, Ibelings BW, Bauwe H, Bauwe H, Matthijs HC, Hagemann M. (2007). Long-term response toward inorganic carbon limitation in wild type and glycolate turnover mutants of the cyanobacterium *Synechocystis* sp. strain PCC 6803. *Plant Physiol* **144**: 1946–1959.
- Elliott JA. (2010). The seasonal sensitivity of cyanobacteria and other phytoplankton to changes in flushing rate and water temperature. *Global Change Biol* **16**: 864–876.
- Emerson S. (1975). Chemically enhanced CO₂ gas exchange in a eutrophic lake: a general model. *Limnol Oceanogr* **20**: 743–753.
- Erdmann N, Fulda S, Hagemann M. (1992). Glucosylglycerol accumulation during salt acclimation of two unicellular cyanobacteria. *J Gen Microbiol* **138**: 363–368.
- Espie GS, Miller AG, Canvin DT. (1988). Characterization of the Na⁺-requirement in cyanobacterial photosynthesis. *Plant Physiol* **88**: 757–763.
- Everitt BS, Landau S, Leese M, Stahl D. (2011). Hierarchical clustering. In: *Cluster Analysis* (5th Edition). pp. 71–110. Chichester, UK: John Wiley & Sons.
- Falconer IR. (2005). *Cyanobacterial Toxins in Drinking Water Supplies: Cylindrospermopsins and Microcystins*. Boca Raton, FL, USA: CRC Press.
- Field CB, Behrenfeld MJ, Randerson JT, Falkowski P. (1998). Primary production of the biosphere: integrating terrestrial and oceanic components. *Science* **281**: 237–240.
- Figge RM, Cassier-Chauvat C, Chauvat F, Cerff R. (2001). Characterization and analysis of an NAD(P)H dehydrogenase transcriptional regulator critical for the survival of cyanobacteria facing inorganic carbon starvation and osmotic stress. *Mol Microbiol* **39**: 455–469.
- Finkel ZV, Beardall J, Flynn KJ, Quigg A, Rees TAV, Raven JA. (2010). Phytoplankton in a changing world: cell size and elemental stoichiometry. *J Plankton Res* **32**: 119–137.
- Fiore MF, Alvarenga DO, Varani AM, Hoff-Rissetti C, Crespin E, Ramos RT, Silva A, Schaker PD, Heck K, Rigonato J, Schneider MP. (2013). Draft genome sequence of the Brazilian toxic bloom-forming cyanobacterium *Microcystis aeruginosa* strain SPC777. *Genome Announc* **1**: e00547–13.
- Flombaum P, Gallegos JL, Gordillo RA, Rincón J, Zabala LL, Jiao N, Karl DM, Li WK, Lomas MW, Veneziano D, Vera CS, Vrugt JA, Martiny AC. (2013). Present and future global distributions of the marine cyanobacteria *Prochlorococcus* and *Synechococcus*. *Proc Natl Acad Sci USA* **110**: 9824–9829.
- Frangoul L, Quillardet P, Castets AM, Humbert JF, Matthijs HCP, Cortez D, Tolonen A, Zhang CC, Gribaldo S, Kehr JC, Zilliges Y, Ziemert N, Becker S, Talla E, Latifi A, Billault A, Lepelletier A, Dittmann E, Bouchier C, de Marsac NT. (2008). Highly plastic genome of *Microcystis aeruginosa* PCC 7806, a ubiquitous toxic freshwater cyanobacterium. *BMC Genomics* **9**: 274.
- Fu FX, Mulholland MR, Garcia NS, Beck A, Bernhardt PW, Warner ME, Sañudo-Wilhelmy SA, Hutchins DA. (2008). Interactions between changing pCO₂, N₂ fixation, and Fe limitation in the marine unicellular cyanobacterium *Crocospaera*. *Limnol Oceanogr* **53**: 2472–2484.
- Fu FX, Warner ME, Zhang Y, Feng Y, Hutchins DA. (2007). Effects of increased temperature and CO₂ on photosynthesis, growth, and elemental ratios in marine *Synechococcus* and *Prochlorococcus* (Cyanobacteria). *J Phycol* **43**: 485–496.
- Gallon JR. (1992). Reconciling the incompatible: N₂ fixation and O₂. *New Phytol* **122**: 571–609.

- Garcia NS, Fu FX, Breene CL, Bernhardt PW, Mulholland MR, Sohm JA, Hutchins DA. (2011). Interactive effects of irradiance and CO₂ on CO₂ fixation and N₂ fixation in the diazotroph *Trichodesmium erythraeum* (Cyanobacteria). *J Phycol* **47**: 1292–1303.
- Garcia-Pichel F, Belnap J, Neuer S, Schanz F. (2003). Estimates of global cyanobacterial biomass and its distribution. *Arch Hydrobiol Algal Stud* **109**: 213–228.
- Gaudana SB, Zarzycki J, Moparthy VK, Kerfeld CA. (2015). Bioinformatic analysis of the distribution of inorganic carbon transporters and prospective targets for bioengineering to increase C_i uptake by cyanobacteria. *Photosynth Res* **126**: 99–109.
- Gehringer MM, Wannicke N. (2014). Climate change and regulation of hepatotoxin production in Cyanobacteria. *FEMS Microbiol Ecol* **88**: 1–25.
- Giordano M, Beardall J, Raven JA. (2005). CO₂ concentrating mechanisms in algae: mechanisms, environmental modulation, and evolution. *Ann Rev Plant Biol* **56**: 99–131.
- Gralla JD, Vargas DR. (2005). Potassium glutamate as transcriptional inhibitor during bacterial osmoregulation. *EMBO J* **25**: 1515–1521.
- Gu B, Schelske CL, Coveney MF. (2011). Low carbon dioxide partial pressure in a productive subtropical lake. *Aquatic Sciences* **73**: 317–330.
- Gulati RD, Van Donk E. (2002). Lakes in the Netherlands, their origin, eutrophication and restoration: state-of-the-art review. *Hydrobiologia* **478**: 73–106.
- Hackenberg C, Huege J, Engelhardt A, Wittink F, Laue M, Matthijs HCP, Kopka J, Bauwe H, Hagemann M. (2012). Low-carbon acclimation in carboxysome-less and photorespiratory mutants of the cyanobacterium *Synechocystis* sp. strain PCC 6803. *Microbiology* **158**: 398–413.
- Hagemann M. (2011). Molecular biology of cyanobacterial salt acclimation. *FEMS Microbiol Rev* **35**: 87–123.
- Hairston NG, Ellner SP, Geber MA, Yoshida T, Fox JA. (2005). Rapid evolution and the convergence of ecological and evolutionary time. *Ecol Lett* **8**: 1114–1127.
- Håkanson L, Bryhn AC, Hytteborn JK. (2007). On the issue of limiting nutrient and predictions of cyanobacteria in aquatic systems. *Sci Total Environ* **379**: 89–108.
- Hakkila K, Antal T, Gunnelius L, Kurkela J, Matthijs HCP, Tyystjärvi E, Tyystjärvi T. (2013). Group 2 sigma factor mutant $\Delta sigCDE$ of the cyanobacterium *Synechocystis* sp. PCC 6803 reveals functionality of both carotenoids and flavodiiron proteins in photoprotection of photosystem II. *Plant Cell Physiol* **54**: 1780–1790.
- Hall NS, Paerl HW, Peierls BL, Whipple AC, Rossignol KL. (2013). Effects of climatic variability on phytoplankton biomass and community structure in the eutrophic, microtidal, New River Estuary, North Carolina, USA. *Estuar Coast Shelf S* **117**: 70–82.
- Havaux M, Guedeny G, Hagemann M, Yermenko N, Matthijs HCP, Jeanjean R. (2005). The chlorophyll-binding protein IsiA is inducible by high light and protects the cyanobacterium *Synechocystis* PCC6803 from photooxidative stress. *FEBS Lett* **579**: 2289–2293.
- Hein M. (1997). Inorganic carbon limitation of photosynthesis in lake phytoplankton. *Freshw Biol* **37**: 545–552.
- Helman Y, Barkan E, Eisenstadt D, Luz B, Kaplan A. (2005). Fractionation of the three stable oxygen isotopes by oxygen-producing and oxygen-consuming reactions in photosynthetic organisms. *Plant Physiol* **138**: 2292–2298.
- Hihara Y, Kamei A, Kanehisa M, Kaplan A, Ikeuchi M. (2001). DNA microarray analysis of cyanobacterial gene expression during acclimation to high light. *Plant Cell* **13**: 793–806.
- Hillebrand H, Dürselen CD, Kirschtel D, Pollinger U, Zohary T. (1999). Biovolume calculation for pelagic and benthic microalgae. *J Phycol* **35**: 403–424.

- Holland DP, Pantorno A, Orr PT, Stojkovic S, Beardall J. (2012). The impacts of a high CO₂ environment on a bicarbonate user: the cyanobacterium *Cylindrospermopsis raciborskii*. *Wat Res* **46**: 1430–1437.
- Hopkinson BM, Young JN, Tansik AL, Binder BJ. (2014). The minimal CO₂ concentrating mechanism of *Prochlorococcus* MED4 is effective and efficient. *Plant Physiol* **166**: 2205–2217.
- Houghton RA. (2007). Balancing the global carbon budget. *Annu Rev Earth Planet Sci* **35**: 313–347.
- Huckauf J, Nomura C, Forchhammer K, Hagemann M. (2000). Stress responses of *Synechocystis* sp. strain PCC 6803 mutants impaired in genes encoding putative alternative sigma factors. *Microbiology* **146**: 2877–2889.
- Huisman J, Matthijs HCP, Visser PM (eds) (2005). *Harmful Cyanobacteria*. Berlin, Germany: Springer.
- Huisman J, Matthijs HCP, Visser PM, Balke H, Sigon CAM, Passarge J, Weissing FJ, Mur LR. (2002). Principles of the light-limited chemostat: theory and ecological applications. *Antonie Leeuwenhoek* **81**: 117–133.
- Huisman J, Sharples J, Stroom JM, Visser PM, Kardinaal WEA, Verspagen JMH, Sommeijer B. (2004). Changes in turbulent mixing shift competition for light between phytoplankton species. *Ecology* **85**: 2960–2970.
- Humbert JH, Barbe V, Latifi A, Gugger M, Calteau A, Coursin T, Lajus A, Castelli V, Oztas S, Samson G, Longin C, Medigue C, de Marsac NT. (2013). A tribute to the disorder in the genome of the bloom-forming freshwater cyanobacterium *Microcystis aeruginosa*. *PLoS One* **8**: e70747.
- Hutchins DA, Fu FX, Webb EA, Walworth N, Tagliabue A. (2013). Taxon-specific response of marine nitrogen fixers to elevated carbon dioxide concentrations. *Nature Geosci* **6**: 790–795.
- Hutchins DA, Fu FX, Zhang Y, Warner ME, Feng Y, Portune K, Bernhardt PW, Mulholland MR. (2007). CO₂ control of *Trichodesmium* N₂ fixation, photosynthesis, growth rates, and elemental ratios: implications for past, present, and future ocean biogeochemistry. *Limnol Oceanogr* **52**: 1293–1304.
- Ibelings BW, Backer LC, Kardinaal WEA, Chorus I. (2014). Current approaches to cyanotoxin risk assessment and risk management around the globe. *Harmful Algae* **40**: 63–74.
- Ibelings BW, Maberly SC. (1998). Photoinhibition and the availability of inorganic carbon restrict photosynthesis by surface blooms of cyanobacteria. *Limnol Oceanogr* **43**: 408–419.
- Igamberdiev AU, Lea PJ. (2006). Land plants equilibrate O₂ and CO₂ concentrations in the atmosphere. *Photosynthesis Res* **87**: 177–194.
- IPCC. (2012). Managing the risks of extreme events and disasters to advance climate change adaptation. In: *A Special Report of Working Groups I and II of the Intergovernmental Panel on Climate Change*. Eds: Field C, Barros V, Stocker, TF, Qin D, Dokken DJ, Ebi KL, Mastrandrea MD, Mach KJ, Plattner GK, Allen SK, Tignor M, Midgley PM. pp. 582. Cambridge, UK and New York, NY, USA: Cambridge University Press.
- IPCC. (2014). Climate change 2014: synthesis report. Contribution of working groups I, II and III to the fifth assessment report of the Intergovernmental Panel on Climate Change [Core writing team, Pachauri RK, Meyer LA (eds.)]. IPCC, Geneva, Switzerland, 151 pp.
- Ishiura M, Kutsuna S, Aoki S, Iwasaki H, Andersson CR, Tanabe A, Golden SS, Johnson CH, Kondo T. (1998). Expression of a gene cluster *kaiABC* as a circadian feedback process in cyanobacteria. *Science* **281**: 1519–1523.
- Islam MS, Drasar BS, Bradley DJ. (1990). Long-term persistence of toxigenic *Vibrio cholerae* 01 in the mucilaginous sheath of a blue-green alga, *Anabaena variabilis*. *J Trop Med Hyg* **93**: 133–139.
- Ito H, Mutsuda M, Murayama Y, Tomita J, Hosokawa N, Terauchi K, Sugita C, Sugita M, Kondo T, Iwasaki H. (2009). Cyanobacterial daily life with Kai-based circadian and diurnal genome-wide transcriptional control in *Synechococcus elongatus*. *Proc Natl Acad Sci USA* **106**: 14168–14173.
- Iwasaki H, Nishiwaki T, Kitayama Y, Nakajima M, Kondo T. (2002). KaiA-stimulated KaiC phosphorylation in circadian timing loops in cyanobacteria. *Proc Natl Acad Sci USA* **99**: 15788–15793.
- Izard J, Limberger RJ. (2003). Rapid screening method for quantitation of bacterial cell lipids from whole cells. *J Microbiol Meth* **55**: 411–418.

- Jacquet S, Briand JF, Leboulanger C, Avois-Jacquet C, Oberhaus L, Tassin B, Vinçon-Leite B, Paolini G, Druart JC, Anneville O, Humbert JF. (2005). The proliferation of the toxic cyanobacterium *Planktothrix rubescens* following restoration of the largest natural French lake (Lac du Bourget). *Harmful algae* **4**: 651–672.
- Jančula D, Maršálek B. (2011). Critical review of actually available chemical compounds for prevention and management of cyanobacterial blooms. *Chemosphere* **85**: 1415–1422.
- Janse I, Kardinaal WEA, Meima M, Fastner J, Visser PM, Zwart G. (2004). Toxic and nontoxic *Microcystis* colonies in natural populations can be differentiated on the basis of rRNA gene internal transcribed spacer diversity. *Appl Environ Microbiol* **70**: 3979–3987.
- Jansson M, Karlsson J, Jonsson, A. (2012). Carbon dioxide supersaturation promotes primary production in lakes. *Ecol Lett* **15**: 527–532.
- Jensen SI, Steunou AS, Bhaya D, Kühl M, Grossman AR. (2011). In situ dynamics of O₂, pH and cyanobacterial transcripts associated with CCM, photosynthesis and detoxification of ROS. *ISME J* **5**: 317–328.
- Jeppesen E, Kronvang B, Meerhoff M, Søndergaard M, Hansen KM, Andersen HE, Lauridsen TL, Liboriussen L, Beklioglu M, Ozen A, Olesen JE. (2009). Climate change effects on runoff, catchment phosphorus loading and lake ecological state, and potential adaptations. *J Environ Qual* **38**: 1930–1941.
- Jeppesen E, Kronvang B, Olesen JE, Audet J, Søndergaard M, Hoffmann CC, Andersen HE, Lauridsen TL, Liboriussen L, Larsen SE, Beklioglu M, Meerhoff M, Özen A, Özkan K. (2011). Climate change effects on nitrogen loading from cultivated catchments in Europe: implications for nitrogen retention, ecological state of lakes and adaptation. *Hydrobiologia* **663**: 1–21.
- Jeppesen E, Søndergaard M, Sortkjær O, Mortensen E, Kristensen P. (1990). Interactions between phytoplankton, zooplankton and fish in a shallow, hypertrophic lake: a study of phytoplankton collapses in Lake Søbygård, Denmark. *Hydrobiologia* **191**: 149–164.
- Jochimsen EM, Carmichael WW, An J, Cardo DM, Cookson ST, Holmes CEM, Antunes MB, de Melo Filho DA, Lyra TM, Barreto VS, Azevedo SM, Jarvis WR. (1998). Liver failure and death following exposure to microcystin toxins at a hemodialysis center in Brazil. *N Engl J Med* **36**: 373–378.
- Jöhnk KD, Huisman J, Sharples J, Sommeijer B, Visser PM, Stroom JM. (2008). Summer heatwaves promote blooms of harmful cyanobacteria. *Glob Change Biol* **14**: 495–512.
- Kanehisa M, Goto S. (2000). KEGG: Kyoto encyclopedia of genes and genomes. *Nucleic Acids Res* **28**: 27–30.
- Kaneko T, Nakajima N, Okamoto S, Suzuki I, Tanabe Y, Tamaoki M, Nakamura Y, Kasai F, Watanabe A, Kawashima K, Kishida Y, Ono A, Shimizu Y, Takahashi C, Minami C, Fujishiro T, Kohara M, Katoh M, Nakazaki N, Nakayama S, Yamada M, Tabata S, Watanabe MM. (2007). Complete genomic structure of the bloom-forming toxic cyanobacterium *Microcystis aeruginosa* NIES-843. *DNA Res* **14**: 247–256.
- Kaplan A, Harel M, Kaplan-Levy, RN, Hadas O, Sukenik A, Dittmann E. (2012). The languages spoken in the water body (or the biological role of cyanobacterial toxins). *Front Microbiol* **3**: 138.
- Kaplan A, Reinhold L. (1999). CO₂ concentrating mechanisms in photosynthetic microorganisms. *Annu Rev Plant Physiol Plant Mol Biol* **50**: 539–570.
- Kardinaal WEA, Tonk L, Janse I, Hol S, Slot P, Huisman J, Visser PM. (2007). Competition for light between toxic and nontoxic strains of the harmful cyanobacterium *Microcystis*. *Appl Environ Microb* **73**: 2939–2946.
- Kardinaal WEA, Visser PM. (2005). Dynamics of cyanobacterial toxins. In: *Harmful Cyanobacteria*, Aquatic Ecology Series. Eds: Huisman J, Matthijs, HCP, Visser PM. pp. 41–64. Dordrecht, the Netherlands: Springer.
- Karlberg M, Wulff A. (2013). Impact of temperature and species interaction on filamentous cyanobacteria may be more important than salinity and increased pCO₂ levels. *Mar Biol* **160**: 2063–2072.
- Klindworth A, Pruesse E, Schweer T, Peplies J, Quast C, Horn M, Glöckner FO. (2012). Evaluation of general 16S ribosomal RNA gene PCR primers for classical and next-generation sequencing-based diversity studies. *Nucleic Acids Res* **41**: e1.

- Kolmakov VI. (2006). Methods for prevention of mass development of the cyanobacteria *Microcystis aeruginosa* Kutz. emend. Elenk. in aquatic systems. *Microbiology* **75**: 149–153.
- Kolman MA, Nishi CN, Perez-Cenci M, Salerno GL. (2015). Sucrose in cyanobacteria: from a salt-response molecule to play a key role in nitrogen fixation. *Life* **5**: 102–126.
- Kolman MA, Torres LL, Martin ML, Salerno GL. (2012). Sucrose synthase in unicellular cyanobacteria and its relationship with salt and hypoxic stress. *Planta* **235**: 955–964.
- Komárek J, Komárková J. (2002). Review of the European *Microcystis*-morphospecies (Cyanoprokaryotes) from nature. *Czech Phycol* **2**: 1–24.
- Koropatkin NM, Koppenaal DW, Pakrasi HB, Smith TJ. (2007). The structure of a cyanobacterial bicarbonate transport protein, CmpA. *J Biol Chem* **282**: 2606–2614.
- Kosten S, Huszar VLM, Bécares E, Costa LS, van Donk E, Hansson LA, Jeppesen E, Kruk C, Lacerot G, Mazzeo N, De Meester L, Moss B, Lürling M, Nöges T, Romo S, Scheffer M. (2012). Warmer climates boost cyanobacterial dominance in shallow lakes. *Glob Change Biol* **18**: 118–126.
- Kranz SA, Dieter S, Richter KU, Rost B. (2009). Carbon acquisition by *Trichodesmium*: the effect of pCO₂ and diurnal changes. *Limnol Oceanogr* **54**: 548–559.
- Kranz SA, Eichner M, Rost B. (2011). Interactions between CCM and N₂ fixation in *Trichodesmium*. *Photosynth Res* **109**: 73–84.
- Kranz SA, Levitan O, Richter KU, Prášil O, Berman-Frank I, Rost B. (2010). Combined effects of CO₂ and light on the N₂-fixing cyanobacterium *Trichodesmium* IMS101: physiological responses. *Plant Physiol* **154**: 334–345.
- Krasikov V. (2012). Dynamic changes in gene expression of the cyanobacterium *Synechocystis* sp. PCC 6803 in response to nitrogen starvation. PhD thesis, University of Amsterdam, the Netherlands.
- Kucho KI, Okamoto K, Tsuchiya Y, Nomura S, Nango M, Kanehisa M, Ishiura M. (2005). Global analysis of circadian expression in the cyanobacterium *Synechocystis* sp. strain PCC 6803. *J Bacteriol* **187**: 2190–2199.
- Kurisu G, Zhang H, Smith JL, Cramer WA. (2003). Structure of the cytochrome b6f complex of oxygenic photosynthesis: tuning the cavity. *Science* **302**: 1009–1014.
- Kurmayer R, Dittmann E, Fastner J, Chorus I. (2002). Diversity of microcystin genes within a population of the toxic cyanobacterium *Microcystis* spp. in lake Wannsee (Berlin, Germany). *Microbial Ecol* **43**: 107–118.
- Latifi A, Ruiz M, Zhang CC. (2009). Oxidative stress in cyanobacteria. *FEMS Microbiol Rev* **33**: 258–278.
- Lazzarino JK, Bachmann RW, Hoyer MV, Canfield DE. (2009). Carbon dioxide supersaturation in Florida lakes. *Hydrobiologia* **627**: 169–180.
- Leão PN, Engene N, Antunes A, Gerwick WH, Vasconcelos V. (2012). The chemical ecology of cyanobacteria. *Nat Prod Rep* **29**: 372–391.
- Lehman PW, Boyer G, Hall C, Waller S, Gehrts K. (2005). Distribution and toxicity of a new colonial *Microcystis aeruginosa* bloom in the San Francisco Bay Estuary, California. *Hydrobiologia* **541**: 87–99.
- Levitan O, Rosenberg G, Setlik I, Setlikova E, Grigel J, Klepetar J, Prasil O, Berman-Frank I. (2007). Elevated CO₂ enhances nitrogen fixation and growth in the marine cyanobacterium *Trichodesmium*. *Glob Change Biol* **13**: 531–538.
- Lewandowska AM, Boyce DG, Hofmann M, Matthiessen B, Sommer U, Worm B. (2014). Effects of sea surface warming on marine plankton. *Ecol Lett* **17**: 614–623.
- Lin S, Haas S, Zemojtel T, Xiao P, Vingron M, Li R. (2011). Genome-wide comparison of cyanobacterial transposable elements, potential genetic diversity indicators. *Gene* **473**: 139–149.
- Lin MT, Occhialini A, Andralojc PJ, Parry MAJ, Hanson MR. (2014). A faster Rubisco with potential to increase photosynthesis in crops. *Nature* **513**: 547–550.

- Livak KJ, Schmittgen TD. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2^{- $\Delta\Delta C_T$} method. *Methods* **25**: 402–408.
- Liu X, Lu X, Chen Y. (2011). The effects of temperature and nutrient ratios on *Microcystis* blooms in Lake Taihu, China: an 11-year investigation. *Harmful Algae* **10**: 337–343.
- Lohbeck KT, Riebesell U, Reusch TBH. (2012). Adaptive evolution of a key phytoplankton species to ocean acidification. *Nature Geosci.* **5**: 346–351.
- Long BM. (2010). Evidence that sulfur metabolism plays a role in microcystin production by *Microcystis aeruginosa*. *Harmful Algae* **9**: 74–81.
- Long BM, Badger MR, Whitney SM, Price GD. (2007). Analysis of carboxysomes from *Synechococcus* PCC7942 reveals multiple Rubisco complexes with carboxysomal proteins CcmM and CcaA. *J Biol Chem* **282**: 29323–29335.
- Long BM, Jones GJ, Orr PT. (2001). Cellular microcystin content in N-limited *Microcystis aeruginosa* can be predicted from growth rate. *Appl Environ Microbiol* **67**: 278–283.
- López-Archilla A, Moreira D, López-García P, Guerrero C. (2004). Phytoplankton diversity and cyanobacterial dominance in a hypereutrophic shallow lake with biologically produced alkaline pH. *Extremophiles* **8**: 109–115.
- Low-Décarie E, Bell G, Fussmann GF. (2015). CO₂ alters community composition and response to nutrient enrichment of freshwater phytoplankton. *Oecologia* **177**: 875–883.
- Low-Décarie E, Jewell MD, Fussmann GF, Bell G. (2013). Long-term culture at elevated atmospheric CO₂ fails to evoke specific adaptation in seven freshwater phytoplankton species. *Proc Royal Soc B: Biol Sci* **280**: 20122598.
- Low-Décarie E, Fussmann GF, Bell G. (2011). The effect of elevated CO₂ on growth and competition in experimental phytoplankton communities. *Glob Change Biol* **17**: 2525–2535.
- Lürling M, Eshetu F, Faassen EJ, Kosten S, Huszar VLM. (2013). Comparison of cyanobacterial and green algal growth rates at different temperatures. *Freshw Biol* **58**: 552–559.
- Lürling M, Faassen EJ. (2012). Controlling toxic cyanobacteria: effects of dredging and phosphorus-binding clay on cyanobacteria and microcystins. *Water Res* **46**: 1447–1459.
- Maberly SC. (1996). Diel, episodic and seasonal changes in pH and concentrations of inorganic carbon in a productive lake. *Freshwater Biol* **35**: 579–598.
- Maberly SC, Ball LA, Raven JA, Sültemeyer D. (2009). Inorganic carbon acquisition by chrysophytes. *J Phycol* **45**: 1052–1061.
- Maberly SC, Barker PA, Stott AW, De Ville MM. (2013). Catchment productivity controls CO₂ emissions from lakes. *Nat Clim Change* **3**: 391–394.
- MacKintosh C, Beattie KA, Klumpp S, Cohen P, Codd GA. (1990). Cyanobacterial microcystin-LR is a potent and specific inhibitor of protein phosphatases 1 and 2A from both mammals and higher plants. *FEBS Lett* **264**: 187–192.
- Maeda S, Badger MR, Price GD. (2002). Novel gene products associated with NdhD3/D4-containing NDH-I complexes are involved in photosynthetic CO₂ hydration in the cyanobacterium, *Synechococcus* sp. PCC7942. *Mol Microbiol* **43**: 425–435.
- Maeda S, Price GD, Badger MR, Enomoto C, Omata T. (2000). Bicarbonate binding activity of the CmpA protein of the cyanobacterium *Synechococcus* sp. strain PCC 7942 involved in active transport of bicarbonate. *J Biol Chem* **275**: 20551–20555.
- Makower K, Schuurmans JM, Groth D, Zilliges Y, Matthijs HCP, Dittmann E. (2015). Transcriptomics-aided dissection of the intracellular and extracellular roles of microcystin in *Microcystis aeruginosa* PCC 7806. *Appl Environ Microbiol* **81**: 544–554.

- Matthijs HCP, Visser PM, Reeze B, Meeuse J, Slot PC, Wijn G, Talens R, Huisman J. (2012). Selective suppression of harmful cyanobacteria in an entire lake with hydrogen peroxide. *Water Res* **46**: 1460–1472.
- McFadden JI. (2001). Chloroplast origin and integration. *Plant Physiol* **125**: 50–53.
- McGinn PJ, Price GD, Maleszka R, Badger MR. (2003). Inorganic carbon limitation and light control the expression of transcripts related to the CO₂-concentrating mechanism in the cyanobacterium *Synechocystis* sp. strain PCC6803. *Plant Physiol* **132**: 218–229.
- McGinn PJ, Price GD, Badger MR. (2004). High light enhances the expression of low-CO₂-inducible transcripts involved in the CO₂-concentrating mechanism in *Synechocystis* sp. PCC6803. *Plant Cell Environ* **27**: 615–626.
- McGrath JM, Long SP. (2014). Can the cyanobacterial carbon-concentrating mechanism increase photosynthesis in crop species? A theoretical analysis. *Plant Physiol* **164**: 2247–2261.
- Meissner S, Fastner J, Dittmann E. (2013). Microcystin production revisited: conjugate formation makes a major contribution. *Environ Microbiol* **15**: 1810–1820.
- Meissner S, Steinhäuser D, Dittmann E. (2015). Metabolomic analysis indicates a pivotal role of the hepatotoxin microcystin in high light adaptation of *Microcystis*. *Environ Microbiol* **17**: 1497–1509
- Merel S, Walker D, Chicana R, Snyder S, Baurès E, Thomas O. (2013). State of knowledge and concerns on cyanobacterial blooms and cyanotoxins. *Environ Int* **59**: 303–327.
- Metcalfe JS, Codd GA. (2012). Cyanotoxins. In: *Ecology of cyanobacteria II. Their diversity in time and space*. Ed: Whitton BA. Dordrecht, the Netherlands: Springer.
- Metcalfe JS, Reilly M, Young FM, Codd GA. (2009). Localization of microcystin synthetase genes in colonies of the cyanobacterium *Microcystis* using fluorescence in situ hybridization. *J Phycol* **45**: 1400–1404.
- Meyer M, Stenzel U, Myles S, Prüfer K, Hofreiter M. (2007). Targeted high-throughput sequencing of tagged nucleic acid samples. *Nucleic Acids Res* **35**: e97
- Mi H, Endo T, Ogawa T, Asada K. (1995). Thylakoid membrane-bound, NADPH-specific pyridine nucleotide dehydrogenase complex mediated cyclic electron transport in the cyanobacterium *Synechocystis* sp. PCC 6803. *Plant Cell Physiol* **36**: 661–668.
- Michalak AM, Anderson EJ, Beletsky D, Boland S, Bosch NS, Bridgeman TB, Chaffin JD, Cho K, Confesor R, Daloglu I, Depinto JV, Evans MA, Fahnenstiel GL, He L, Ho JC, Jenkins L, Johengen TH, Kuo KC, Laporte E, Liu X, McWilliams MR, Moore MR, Posselt DJ, Richards RP, Scavia D, Steiner AL, Verhamme E, Wright DM, Zagorski MA. (2013). Record-setting algal bloom in Lake Erie caused by agricultural and meteorological trends consistent with expected future conditions. *Proc Natl Acad. Sci USA* **110**: 6448–6452.
- Mikkat S, Milkowski C, Hagemann M. (2000). The gene *slr0273* of the cyanobacterium *Synechocystis* sp. strain PCC6803 encodes a protein essential for growth at low Na⁺/K⁺ ratios. *Plant Cell Environ* **23**: 549–559.
- Miller AG, Espie GS, Canvin DT. (1988). Active transport of inorganic carbon increases the rate of O₂ photoreduction by the cyanobacterium *Synechococcus* UTEX 625. *Plant Physiol* **88**: 6–9.
- Miller AG, Turpin DH, Canvin DT. (1984). Growth and photosynthesis of the cyanobacterium *Synechococcus leopoliensis* in HCO₃⁻-limited chemostats. *Plant Physiol* **75**: 1064–1070.
- Millie DF, Fahnenstiel GL, Bressie JD, Pigg R J, Rediske RR, Klarer DM, Tester PA, Litaker RW. (2009). Late-summer phytoplankton in western Lake Erie (Laurentian Great Lakes): bloom distributions, toxicity, and environmental influences. *Aquat Ecol* **43**: 915–934.
- Mitrovic SM, Oliver RL, Rees C, Bowling LC, Buckney RT. (2003). Critical flow velocities for the growth and dominance of *Anabaena circinalis* in some turbid freshwater rivers. *Freshw Biol* **48**: 164–174.

- Mitschke J, Georg J, Scholz I, Sharma CM, Dienst D, Bantscheff J, Voss B, Steglich C, Wilde A, Vogel J, Hess WR. (2011). An experimentally anchored map of transcriptional start sites in the model cyanobacterium *Synechocystis* sp. PCC6803. *Proc Natl Acad Sci USA* **108**: 2124–2129.
- Moisander PH, Lehman PW, Ochiai M, Corum S. (2009). Diversity of *Microcystis aeruginosa* in the Klamath River and San Francisco Bay delta, California, USA. *Aquat Microb Ecol* **57**: 19–31.
- Moran AM. (2009). Metatranscriptomics: eavesdropping on complex microbial communities. *Microbe* **4**: 329–335.
- Moroney JV, Ma Y, Frey WD, Fusilier KA, Pham TT, Simms TA, DiMario RJ, Yang J, Mukherjee B. (2011). The carbonic anhydrase isoforms of *Chlamydomonas reinhardtii*: Intracellular location, expression, and physiological roles. *Photosynth Res* **109**: 133–149.
- Moroney JV, Ynalvez RA. (2007). Proposed carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*. *Eukaryot Cell* **6**: 1251–1259.
- Murata N, Takahashi S, Nishiyama Y, Allakhverdiev SI. (2007). Photoinhibition of photosystem II under environmental stress. *BBA-Bioenergetics* **1767**: 414–421.
- Muro-Pastor AM, Hess WR. (2012). Heterocyst differentiation: from single mutants to global approaches. *Trends Microbiol* **20**: 548–557.
- Murphy J, Riley JP. (1962). A modified single solution for the determination of phosphate in natural waters. *Anal Chim Acta* **27**: 31–36.
- Muyzer G, De Waal EC, Uitterlinden AG. (1993). Profiling of complex microbial populations by denaturing gradient gel electrophoresis analysis of polymerase chain reaction-amplified genes coding for 16S rRNA. *Appl Environ Microbiol* **59**: 695–700.
- Nakamura Y, Kaneko T, Hirosawa M, Miyajima N, Tabata S. (1998). CyanoBase, a www database containing the complete nucleotide sequence of the genome of *Synechocystis* sp. strain PCC6803. *Nucleic Acids Res* **26**: 63–67.
- Nakasugi KR, Alexova CJ, Svenson, Neilan BA. (2007). Functional analysis of PilT from the toxic cyanobacterium *Microcystis aeruginosa* PCC 7806. *J Bacteriol* **189**: 1689–1697.
- Nakasugi K, Neilan BA. (2005). Identification of pilus-like structures and genes in *Microcystis aeruginosa* PCC7806. *Appl Environ Microbiol* **71**: 7621–7625.
- Neilan BA, Pearson LA, Muenchhoff J, Moffitt MC, Dittmann E. (2013). Environmental conditions that influence toxin biosynthesis in cyanobacteria. *Environ Microbiol* **15**: 1239–1253.
- Nishimura T, Takahashi Y, Yamaguchi O, Suzuki H, Maeda S, Omata T. (2008). Mechanism of low CO₂-induced activation of the *cmp* bicarbonate transporter operon by a LysR family protein in the cyanobacterium *Synechococcus elongatus* strain PCC 7942. *Mol Microbiol* **68**: 98–109.
- Nishiwaki-Matsushima R, Ohta T, Nishiwaki S, Sukanuma M, Kohyama K, Ishikawa T, Carmichael WW, Fujiki H. (1992). Liver tumor promotion by the cyanobacterial cyclic peptide toxin microcystin-LR. *J Canc Res Clin Oncol* **118**: 420–424.
- Nodop A, Pietsch D, Höcker R, Becker A, Pistorius EK, Forchhammer K, Michel KP. (2008). Transcript profiling reveals new insights into the acclimation of the mesophilic fresh-water cyanobacterium *Synechococcus elongatus* PCC 7942 to iron starvation. *Plant Physiol* **147**: 747–763.
- Oberholster PJ, Myburgh JG, Ashton PJ, Botha AM. (2010). Responses of phytoplankton upon exposure to a mixture of acid mine drainage and high levels of nutrient pollution in Lake Loskop, South Africa. *Ecotox Environ Safety* **73**: 326–335.
- Ogden DE, Sleep NH. (2012). Explosive eruption of coal and basalt and the end-Permian mass extinction. *Proc Natl Acad Sci USA* **109**: 59–62.
- Ohkawa H, Price GD, Badger MR, Ogawa T. (2000). Mutation of *ndh* genes leads to inhibition of CO₂ uptake rather than HCO₃⁻ uptake in *Synechocystis* sp. strain PCC 6803. *J Bacteriol* **182**: 2591–2596.

- Okano K, Miyata N, Ozaki Y. (2015). Whole genome sequence of the non-microcystin-producing *Microcystis aeruginosa* strain NIES-44. *Genome Announc* **3**: e00135–15.
- Omata T, Gohta S, Takahashi Y, Harano Y, Maeda, S. (2001). Involvement of a CbbR homolog in low CO₂-induced activation of the bicarbonate transporter operon in cyanobacteria. *J Bacteriol* **183**: 1891–1898.
- Omata T, Price GD, Badger MR, Okamura M, Gohta S, Ogawa T. (1999). Identification of an ATP-binding cassette transporter involved in bicarbonate uptake in the cyanobacterium *Synechococcus* sp. strain PCC 7942. *Proc Natl Acad Sci USA* **96**: 13571–13576.
- Omata T, Takahashi Y, Yamaguchi O, Nishimura T. (2002). Structure, function and regulation of the cyanobacterial high-affinity bicarbonate transporter, BCT1. *Funct Plant Biol* **29**: 151–159.
- O’Neil JM, Davis TW, Burford MA, Gobler CJ. (2012). The rise of harmful cyanobacteria blooms: the potential roles of eutrophication and climate change. *Harmful Algae* **14**: 313–334.
- Orcutt KM, Ren S, Gundersen K. (2009). Detecting proteins in highly autofluorescent cells using quantum dot antibody conjugates. *Sensors* **9**: 7540–7549.
- Orr JC, Fabry VJ, Aumont O, Bopp L, Doney SC, Feely RA, Gnanadesikan A, Gruber N, Ishida A, Joos F, Key RM, Lindsay K, Maier-Reimer E, Matear R, Monfray P, Mouchet A, Najjar RG, Plattner GK, Rodgers KB, Sabine CL, Sarmiento JL, Schlitzer R, Slater RD, Totterdell IJ, Weirig MF, Yamanaka Y, Yool A. (2005). Anthropogenic ocean acidification over the twenty-first century and its impact on calcifying organisms. *Nature* **437**: 681–686.
- Orr PT, Jones GJ, Douglas GB. (2004). Response of cultured *Microcystis aeruginosa* from the Swan River, Australia, to elevated salt concentration and consequences for bloom and toxin management in estuaries. *Mar Freshw Res* **55**: 277–283.
- Otsuka S, Suda S, Shibata S, Oyaizu H, Matsumoto S, Watanabe MM. (2001). A proposal for the unification of five species of the cyanobacterial genus *Microcystis* Kützing ex Lemmermann 1907 under the Rules of the Bacteriological Code. *Int J Syst Evol Microbiol* **51**: 873–879.
- Pacala SW, Hurtt GC, Baker D, Peylin P, Houghton RA, Birdsey RA, Heath L, Sundquist ET, Stallard RF, Ciais P, Moorcroft P, Caspersen JP, Shevliakova E, Moore B, Kohlmaier G, Holland E, Gloor M, Harmon ME, Fan SM, Sarmiento JL, Goodale CL, Schimel D, Field CB. (2001). Consistent land-and atmosphere-based US carbon sink estimates. *Science* **292**: 2316–2320.
- Paerl HW. (2008). Nutrient and other environmental controls of harmful cyanobacterial blooms along the freshwater-marine continuum. *Adv Exp Med Biol* **619**: 217–237.
- Paerl HW, Fulton RS 3rd, Moisaner PH, Dyle J. (2001). Harmful freshwater algal blooms, with an emphasis on cyanobacteria. *ScientificWorld* **1**: 76e113.
- Paerl HW, Huisman J. (2008). Blooms like it hot. *Science* **320**: 57–58.
- Paerl, HW, Huisman J. (2009). Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. *Environ Microbiol Rep* **1**: 27–37.
- Paerl HW, Paul VJ. (2012). Climate change: links to global expansion of harmful cyanobacteria. *Water Res* **46**: 1349–1363.
- Paerl HW, Ustach J. (1982). Blue-green algal scums: an explanation for their occurrence during freshwater blooms. *Limnol Oceanogr* **27**: 212–217.
- Parker DL, Kumar HD, Rai LC, Singh JB. (1997). Potassium salts inhibit the growth of the cyanobacteria *Microcystis* spp. in pond water and defined media: implications for control of microcystin-producing aquatic blooms. *Appl Environ Microbiol* **63**: 2324–2329.
- Partensky F, Garczarek L. (2010). *Prochlorococcus*: advantages and limits of minimalism. *Ann Rev Mar Sci* **2**: 305–331.

- Paul AJ, Achterberg EP, Bach LT, Boxhammer T, Czerny J, Haunost M, Schulz KG, Stuhler A, Riebesell U. (2015). No observed effect of ocean acidification on nitrogen biogeochemistry in a summer Baltic Sea plankton community. *Biogeosci Discuss* **12**: 17507–17541.
- Pearson PN, Palmer MR. (2000). Estimating Paleogene atmospheric pCO₂ using boron isotope analysis of foraminifera. *GFF* **122**: 127–128.
- Peeters F, Straile D, Lorke A, Livingstone DM. (2007). Earlier onset of the spring phytoplankton bloom in lakes of the temperate zone in a warmer climate. *Glob Chang Biol* **13**: 1898–1909.
- Pengelly JLL, Förster B, Von Caemmerer S, Badger MR, Price GD, Whitney SM. (2014). Transplastomic integration of a cyanobacterial bicarbonate transporter into tobacco chloroplasts. *J Exp Bot* **65**: 3071–3080.
- Penn K, Wang J, Fernando S, Thompson J. (2014). Secondary metabolite gene expression and interplay of bacterial functions in a tropical freshwater cyanobacterial bloom. *ISME J* **8**: 1866–1878.
- Poretsky R, Rodriguez-R LM, Luo C, Tsementzi D, Konstantinidis KT. (2014). Strengths and limitations of 16S rRNA gene amplicon sequencing in revealing temporal microbial community dynamics. *PLoS One* **9**: e93827.
- Price GD. (2011). Inorganic carbon transporters of the cyanobacterial CO₂ concentrating mechanism. *Photosynth Res* **109**: 47–57.
- Price GD, Badger MR, von Caemmerer S. (2011a). The prospect of using cyanobacterial bicarbonate transporters to improve leaf photosynthesis in C₃ crop plants. *Plant Physiol* **155**: 20–26.
- Price GD, Badger MR, Woodger FJ, Long BM. (2008). Advances in understanding the cyanobacterial CO₂-concentrating-mechanism (CCM): functional components, Ci transporters, diversity, genetic regulation and prospects for engineering into plants. *J Exp Bot* **59**: 1441–1461.
- Price GD, Howitt SM. (2011). The cyanobacterial bicarbonate transporter BicA: its physiological role and the implications of structural similarities with human SLC26 transporters. *Biochem Cell Biol* **89**: 178–188.
- Price GD, Howitt SM. (2014). Plant science: towards turbocharged photosynthesis. *Nature* **513**: 497–498.
- Price GD, Maeda S, Omata T, Badger MR. (2002). Modes of active inorganic carbon uptake in the cyanobacterium, *Synechococcus* sp. PCC7942. *Funct Plant Biol* **29**: 131–149.
- Price GD, Pengelly JJ, Forster B, Du J, Whitney SM, von Caemmerer S, Badger MR, Howitt SM, Evans JR. (2013). The cyanobacterial CCM as a source of genes for improving photosynthetic CO₂ fixation in crop species. *J Exp Bot* **64**: 753–768.
- Price GD, Sheldon MC, Howitt SM. (2011b). Membrane topology of the cyanobacterial bicarbonate transporter, SbtA, and identification of potential regulatory loops. *Mol Membr Biol* **28**: 265–275.
- Price GD, Woodger FJ, Badger MR, Howitt SM, Tucker L. (2004). Identification of a SulP-type bicarbonate transporter in marine cyanobacteria. *Proc Natl Acad Sci USA* **101**: 18228–18233.
- Qin B, Zhu G, Gao G, Zhang Y, Li W, Paerl HW, Carmichael WW. (2010). A drinking water crisis in Lake Taihu, China: linkage to climatic variability and lake management. *Environ. Management* **45**: 105–112.
- Rae BD, Forster B, Badger MR, Price GD. (2011). The CO₂-concentrating mechanism of *Synechococcus* WH5701 is composed of native and horizontally-acquired components. *Photosynth Res* **109**: 59–72.
- Ramakers C, Ruijter JM, Deprez RH, Moorman AF. (2003). Assumption-free analysis of quantitative real-time polymerase chain reaction (PCR) data. *Neurosci Lett* **339**: 62–66.
- Raven JA. (2010). Inorganic carbon acquisition by eukaryotic algae: four current questions. *Photosynth Res* **106**: 123–134.
- Raven JA, Cockell CS, De La Rocha CL. (2008). The evolution of inorganic carbon concentrating mechanisms in photosynthesis. *Phil Trans R Soc Lond B Biol Sci* **363**: 2641–2650.
- Raven JA, Giordano M, Beardall J, Maberly SC. (2012). Algal evolution in relation to atmospheric CO₂: carboxylases, carbon-concentrating mechanisms and carbon oxidation cycles. *Phil Trans R Soc B* **367**: 493–507.

- Reyes JC, Muro-Pastor MI, Florencio FJ. (1997). Transcription of glutamine synthetase genes (*glnA* and *glnN*) from the cyanobacterium *Synechocystis* sp. strain PCC 6803 is differently regulated in response to nitrogen availability. *J Bacteriol* **179**: 2678–2689.
- Reynolds CS, Walsby AE. (1975). Water-blooms. *Biol Rev* **50**: 437–481.
- Richey JE, Melack JM, Aufdenkampe AK, Ballester VM, Hess LL. (2002). Outgassing from Amazonian rivers and wetlands as a large tropical source of atmospheric CO₂. *Nature* **416**: 617–620.
- Rigosi A, Carey CC, Ibelings BW, Brookes JD. (2014). The interaction between climate warming and eutrophication to promote cyanobacteria is dependent on trophic state and varies among taxa. *Limnol Oceanogr* **59**: 99–114.
- Rippka R, Deruelles J, Waterbury JB, Herdman M, Stanier RY. (1979). Generic assignments, strain histories and properties of pure cultures of cyanobacteria. *Microbiology* **111**: 1–61.
- Roberts RD, Zohary T. (1987). Temperature effects on photosynthetic capacity, respiration, and growth rates of bloom-forming cyanobacteria. *N Z J Mar Freshw Res* **21**: 391–399.
- Roberts EW, Cai F, Kerfeld CA, Cannon GC, Heinhorst S. (2012). Isolation and characterization of the *Prochlorococcus* carboxysome reveal the presence of the novel shell protein CsoS1D. *J Bacteriol* **194**: 787–795.
- Robinson NJ, Robinson PJ, Gupta A, Bleasby AJ, Whitton BA, Morby AP. (1995). Singular over-representation of an octameric palindrome, HIP1, in DNA from many cyanobacteria. *Nucleic Acids Res* **23**: 729–735.
- Robinson PJ, Cranenburgh RM, Head IM, Robinson NJ. (1997). HIP1 propagates in cyanobacterial DNA via nucleotide substitutions but promotes excision at similar frequencies in *Escherichia coli* and *Synechococcus* PCC 7942. *Mol Microbiol* **24**: 181–189.
- Robson BJ, Hamilton DP. (2003). Summer flow event induces a cyanobacterial bloom in a seasonal Western Australian estuary. *Mar Freshwater Res* **54**: 139–151.
- Rost B, Riebesell U, Burkhardt S, Sültemeyer D. (2003). Carbon acquisition of bloom-forming marine phytoplankton. *Limnol Oceanogr* **48**: 55–67.
- Rost B, Zondervan I, Wolf-Gladrow D. (2008). Sensitivity of phytoplankton to future changes in ocean carbonate chemistry: current knowledge, contradictions and research directions. *Mar Ecol Prog Ser* **373**: 227–237.
- Ruijter JM, Ramakers C, Hoogaars WM, Karlen Y, Bakker O, Van den Hoff MJ, Moorman AF. (2009). Amplification efficiency: linking baseline and bias in the analysis of quantitative PCR data. *Nucleic Acids Res* **37**: e45.
- Runnegar MT, Falconer LR, Silver J. (1981). Deformation of isolated rat hepatocytes by a peptide from the blue-green alga *Microcystis aeruginosa*. *Arch Pharmacol* **317**: 268–272.
- Runnegar M, Berndt N, Kong SM, Lee EY, Zhang L. (1995). *In vivo* and *in vitro* binding of microcystin to protein phosphatases 1 and 2A. *Biochem Biophys Res Commun* **216**: 162–169.
- Saker ML, Fastner J, Dittmann E, Christiansen G, Vasconcelos VM. (2005). Variation between strains of the cyanobacterium *Microcystis aeruginosa* isolated from a Portuguese river. *J Appl Microbiol* **99**: 749–757.
- Sandrini G, Cunsolo S, Schuurmans JM, Matthijs HCP, Huisman J. (2015a). Changes in gene expression, cell physiology and toxicity of the harmful cyanobacterium *Microcystis aeruginosa* at elevated CO₂. *Front Microbiol* **6**: 401.
- Sandrini G, Huisman J, Matthijs HCP. (2015b). Strain-specific potassium ion sensitivity of the harmful cyanobacterium *Microcystis* correlates with the prevalence of specific salt tolerance genes. *FEMS Microbiol Lett* **362**: fmv121.
- Sandrini G, Jakupovic D, Matthijs HCP, Huisman J. (2015c). Strains of the harmful cyanobacterium *Microcystis aeruginosa* differ in gene expression and activity of inorganic carbon uptake systems at elevated CO₂ levels. *Appl Environ Microbiol* **81**: 7730–7739.

- Sandrini G, Matthijs HCP, Verspagen JMH, Muyzer G, Huisman J. (2014). Genetic diversity of inorganic carbon uptake systems causes variation in CO₂ response of the cyanobacterium *Microcystis*. *ISME J* **8**: 589–600.
- Scheffer M. (1998). *Ecology of Shallow Lakes*. Chapman and Hall: London.
- Scheinin M, Riebesell U, Rynearson TA, Lohbeck KT, Collins S. (2015). Experimental evolution gone wild. *J R Soc Interface* **12**: 20150056.
- Schippers P, Lürling M, Scheffer M. (2004). Increase of atmospheric CO₂ promotes phytoplankton productivity. *Ecol Lett* **7**: 446–451.
- Schirrmeister BE, de Vos JM, Antonelli A, Bagheri HC. (2013). Evolution of multicellularity coincided with increased diversification of cyanobacteria and the Great Oxidation Event. *Proc Natl Acad Sci USA* **110**: 1791–1796.
- Schmider E, Ziegler M, Danay E, Beyer L, Bühner M. (2010). Is it really robust? Reinvestigating the robustness of ANOVA against violations of the normal distribution assumption. *Methodology (Gott)* **6**: 147–151.
- Schoener, TW. (2011). The newest synthesis: understanding the interplay of evolutionary and ecological dynamics. *Science* **331**: 426–429.
- Schopf JW. (1993) Microfossils of the Early Archean Apex chert: new evidence of the antiquity of life. *Science* **260**: 640–646.
- Schwabe W, Weihe A, Börner T, Henning M, Kohl JG. (1988). Plasmids in toxic and nontoxic strains of the cyanobacterium *Microcystis aeruginosa*. *Curr Microbiol* **17**: 133–137.
- Schwarz D, Nodop A, Hüge J, Purfürst S, Forchhammer K, Michel KP, Bauwe H, Kopka J, Hagemann M. (2011). Metabolic and transcriptomic phenotyping of inorganic carbon acclimation in the cyanobacterium *Synechococcus elongatus* PCC 7942. *Plant Physiol* **155**: 1640–1655.
- Sevilla E, Martín-Luna B, Vela L, Bes MT, Fillat MF, Peleato ML. (2008). Iron availability affects *mcyD* expression and microcystin-LR synthesis in *Microcystis aeruginosa* PCC7806. *Environ Microbiol* **10**: 2476–2483.
- Shapiguzov A, Lyukevich AA, Allakhverdiev SI, Sergeyenko TV, Suzuki I, Murata N, Los DA. (2005). Osmotic shrinkage of cells of *Synechocystis* sp. PCC 6803 bywater efflux via aquaporins regulates osmotic stress-inducible gene expression. *Microbiology* **151**: 447–455.
- Shapiro J. (1990). Current beliefs regarding dominance of bluegreens: the case for the importance of CO₂ and pH. *Verh Int Ver Theor Angew Limnol* **24**: 38–54.
- Shapiro J. (1997). The role of carbon dioxide in the initiation and maintenance of blue-green dominance in lakes. *Freshw Biol* **37**: 307–323.
- Shibata M, Katoh H, Sonoda M, Ohkawa H, Shimoyama M, Fukuzawa H, Kaplan A, Ogawa T. (2002). Genes essential to sodium-dependent bicarbonate transport in cyanobacteria. *J Biol Chem* **277**: 18658–18664.
- Shibata M, Ohkawa H, Kaneko T, Fukuzawa H, Tabata S, Kaplan A, Ogawa T. (2001). Distinct constitutive and low-CO₂-induced CO₂ uptake systems in cyanobacteria: genes involved and their phylogenetic relationship with homologous genes in other organisms. *Proc Natl Acad Sci USA* **98**: 11789–11794.
- Shih PM, Wu D, Latifi A, Axen SD, Fewer DP, Talla E, Calteau A, Cai F, Tandeau de Marsac N, Rippka R, Herdman M, Sivonen K, Coursin T, Laurent T, Goodwin L, Nolan M, Davenport KW, Han CS, Rubin EM, Eisen JA, Woyke T, Guggen M, Kerfeld CA. (2013). Improving the coverage of the cyanobacterial phylum using diversity-driven genome sequencing. *Proc Natl Acad Sci USA* **110**: 1053–1058.
- Shively JM, van Keulen G, Meijer WG. (1998). Something from almost nothing: carbon dioxide fixation in chemoautotrophs. *Annu Rev Microbiol* **52**: 191–230.
- Shokralla S, Spall JL, Gibson JF, Hajibabaei M. (2012). Next-generation sequencing technologies for environmental DNA research. *Mol Ecol* **21**: 1794–1805.
- Shukla B, Rai LC. (2007). Potassium-induced inhibition of nitrogen and phosphorus metabolism as a strategy for controlling *Microcystis* blooms. *World J Microb Biot* **23**: 317–322.
- Siegenthaler U, Sarmiento JL. (1993). Atmospheric carbon dioxide and the ocean. *Nature* **365**: 119–125.

- Sivonen K, Jones G. (1999). Cyanobacterial toxins. In: *Toxic Cyanobacteria in water: A Guide to their Public Health-Consequences, Monitoring and Management*. Eds: Chorus I, Bartram J. London, UK: E & FN Spon.
- Smith AM, Nie SM. (2004). Chemical analysis and cellular imaging with quantum dots. *Analyst* **129**: 672–677.
- Smith KS, Ferry JG. (2000). Prokaryotic carbonic anhydrases. *FEMS Microbiol Rev* **24**: 335–366.
- Smith VH. (1998). Cultural eutrophication of inland, estuarine, and coastal waters. In: *Successes, limitations, and frontiers in ecosystem science* pp. 7–49. New York, NY, USA: Springer.
- Smyth GK. (2005). Limma: linear models for microarray data. In: *Bioinformatics and Computational Biology Solutions Using R and Bioconductor*. Eds: Gentleman R, Carey V, Dudoit S, Irizarry R, Huber W. pp. 397–420. New York, NY, USA: Springer.
- So AKC, Van Spall HGC, Coleman JR, Espie GS. (1998). Catalytic exchange of ^{18}O from $^{13}\text{C}^{18}\text{O}$ -labelled CO_2 by wild-type cells and *ecaA*, *ecaB*, and *ccaA* mutants of the cyanobacteria *Synechococcus* PCC7942 and *Synechocystis* PCC6803. *Can J Botany* **76**: 1153–1160.
- Sobek S, Tranvik LJ, Cole JJ. (2005). Temperature independence of carbon dioxide supersaturation in global lakes. *Global Biogeochem Cycles* **19**: GB2003. doi:10.1029/2004GB002264.
- Solomon S, Qin D, Manning M, Marquis M, Averyt K, Tignor MMB Miller HL Jr, Chen Z. (2007). Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge, UK: Cambridge University Press.
- Soltes-Rak E, Mulligan ME, Coleman JR. (1997). Identification and characterization of a gene encoding a vertebrate-type carbonic anhydrase in cyanobacteria. *J Bacteriol* **179**: 769–774.
- Song YF, Qiu BS. (2007). The CO_2 concentrating mechanism in the bloom-forming cyanobacterium *Microcystis aeruginosa* (Cyanophyceae) and effects of UVB radiation on its operation. *J Phycol* **43**: 957–964.
- Spalding MH. (2008). Microalgal carbon-dioxide-concentrating mechanisms: *Chlamydomonas* inorganic carbon transporters. *J Exp Bot* **59**: 1463–1473.
- Spijkerman E, De Castro F, Gaedke U. (2011). Independent colimitation for carbon dioxide and inorganic phosphorus. *PLoS One* **6**: e28219.
- Steffen MM, Belisle BS, Watson SB, Boyer GL, Wilhelm SW. (2014). Status, causes and controls of cyanobacterial blooms in Lake Erie. *J Great Lakes Res* **40**: 215–225.
- Steffen MM, Belisle BS, Watson SB, Boyer GL, Bourbonniere RA, Wilhelm SW. (2015). Metatranscriptomic evidence for co-occurring top-down and bottom-up controls on toxic cyanobacterial communities. *Appl Environ Microbiol* **81**: 3268–3276.
- Storey JD, Tibshirani R. (2003). Statistical significance for genomewide studies. *Proc Natl Acad Sci USA* **100**: 9440–9445.
- Straub C, Quillardet P, Vergalli J, De Marsac NT, Humbert JF. (2011). A day in the life of *Microcystis aeruginosa* strain PCC 7806 as revealed by a transcriptomic analysis. *PLoS One* **6**: e16208.
- Stumm W, Morgan JJ. (1996). *Aquatic Chemistry: Chemical Equilibria and Rates in Natural Waters*. New York, NY, USA: Wiley-Interscience.
- Suikkanen S, Laamanen M, Huttunen M. (2007). Long-term changes in summer phytoplankton communities of the open northern Baltic Sea. *Estuar Coast S Sci* **71**: 580–592.
- Sültemeyer D, Price GD, Yu JW, Badger MR. (1995). Characterisation of carbon dioxide and bicarbonate transport during steady-state photosynthesis in the marine cyanobacterium *Synechococcus* strain PCC7002. *Planta* **197**: 597–607.
- Talling JF. (1976). The depletion of carbon dioxide from lake water by phytoplankton. *J Ecol* **64**: 79–121.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* **28**: 2731–2739.

- Tans PP, Fung IY, Takahashi T. (1990). Observational constraints on the global atmospheric CO₂ budget. *Science* **247**: 1431–1439.
- Taranu ZE, Zurawell RW, Pick F, Gregory-Eaves I. (2012). Predicting cyanobacterial dynamics in the face of global change: the importance of scale and environmental context. *Glob Change Biol* **18**: 3477–3490.
- Taş S, Okuş E, Aslan-Yılmaz A. (2006). The blooms of a cyanobacterium, *Microcystis* cf. *aeruginosa* in a severely polluted estuary, the Golden Horn, Turkey. *Estuar Coast Shelf S* **68**: 593–599.
- Tchernov D, Helman Y, Keren N, Luz B, Ohad I, Reinhold L, Ogawa T, Kaplan A. (2001). Passive entry of CO₂ and its intracellular conversion to HCO₃⁻ in cyanobacteria are driven by a photosystem I-generated ΔμH⁺. *J Biol Chem* **276**: 23450–23455.
- Thiel T, Pratte BS, Zhong J, Goodwin L, Copeland A, Lucas S, Han C, Pitluck S, Land ML, Kyrpidis NC, Woyke T. (2014). Complete genome sequence of *Anabaena variabilis* ATCC 29413. *Stand Genomic Sci* **9**: 562.
- Thomas RH, Walsby AE. (1985). Buoyancy regulation in a strain of *Microcystis*. *J Gen Microbiol* **131**: 799–809.
- Thompson JD, Higgins DG, Gibson TJ. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res* **22**: 4673–4680.
- Tillett D, Dittmann E, Erhard M, von Dohren H, Borner T, Neilan BA. (2000). Structural organization of microcystin biosynthesis in *Microcystis aeruginosa* PCC7806: an integrated peptide-polyketide synthetase system. *Chem Biol* **7**: 753–764.
- Tonk L, Bosch K, Visser PM, Huisman J. (2007). Salt tolerance of the harmful cyanobacterium *Microcystis aeruginosa*. *Aquat Microb Ecol* **46**: 117–123.
- Tonk L, Van de Waal DB, Slot P, Huisman J, Matthijs HCP, Visser PM. (2008). Amino acid availability determines the ratio of microcystin variants in the cyanobacterium *Planktothrix agardhii*. *FEMS Microbiol. Ecol.* **65**: 383–390.
- Tonk L, Visser PM, Christiansen G, Dittmann E, Snelder EO, Wiedner C, Mur LR, Huisman J. (2005). The microcystin composition of the cyanobacterium *Planktothrix agardhii* changes toward a more toxic variant with increasing light intensity. *Appl Environ Microbiol* **71**: 5177–5181.
- Tonk L, Welker M, Huisman J, Visser PM. (2009). Production of cyanopeptolins, anabaenopeptins, and microcystins by the harmful cyanobacteria *Anabaena* 90 and *Microcystis* PCC 7806. *Harmful Algae* **8**: 219–224.
- Tooming-Klunderud A, Sogge H, Rounge TB, Nederbragt AJ, Lagesen K, Glöckner G, Hayes PK, Rohrlack T, Jakobsen KS. (2013). From green to red: horizontal gene transfer of the phycoerythrin gene cluster between *Planktothrix* strains. *Appl Environ Microbiol* **79**: 6803–6812.
- Tortell PD, DiTullio GR, Sigman DM, Morel FMM. (2002). CO₂ effects on taxonomic composition and nutrient utilization in an Equatorial Pacific phytoplankton assemblage. *Mar Ecol Prog Ser* **236**: 37–43.
- Trenberth KE (2005). The impact of climate change and variability on heavy precipitation, floods, and droughts. In: *Encyclopedia of Hydrological Sciences*. ED: Anderson MG. Chichester, UK: John Wiley & Sons.
- Trogen GB, Annala A, Eriksson J, Kontteli M, Meriluoto J, Sethson I, Zdunek J, Edlund U. (1996). Conformational studies of microcystin-LR using NMR spectroscopy and molecular dynamics calculations. *Biochemistry* **35**: 3197–3205.
- US DOE. (2008). Carbon cycling and biosequestration: integrating biology and climate through systems science. Report from the March 2008 Workshop, DOE/SC-108, U.S. Department of Energy Office of Science (genomicscience.energy.gov/carboncycle/).
- Van de Waal, DB, Ferreruella G, Tonk L, Van Donk E, Huisman J, Visser, PM, Matthijs HCP. (2010a). Pulsed nitrogen supply induces dynamic changes in the amino acid composition and microcystin production of the harmful cyanobacterium *Planktothrix agardhii*. *FEMS Microbiol Ecol* **74**: 430–438.

- Van de Waal DB, Verschoor AM, Verspagen JMH, Van Donk E, Huisman J. (2010b). Climate-driven changes in the ecological stoichiometry of aquatic ecosystems. *Front Ecol Environ* **8**: 145–152.
- Van de Waal DB, Verspagen JMH, Finke JF, Vournazou V, Immers AK, Kardinaal WEA, Tonk L, Becker S, Van Donk E, Visser PM, Huisman J. (2011). Reversal in competitive dominance of a toxic versus non-toxic cyanobacterium in response to rising CO₂. *ISME J* **5**: 1438–1450.
- Van de Waal DB, Verspagen JMH, Lüring M, Van Donk E, Visser PM, Huisman J. (2009). The ecological stoichiometry of toxins produced by harmful cyanobacteria: an experimental test of the carbon-nutrient balance hypothesis. *Ecol Lett* **12**: 1326–1335.
- Van Dijk EL, Auger H, Jaszczyszyn Y, Thermes C. (2014). Ten years of next-generation sequencing technology. *Trends Genet* **30**: 418–426.
- Van Gremberghe I, Leliaert F, Mergeay J, Vanormelingen P, Van der Gucht K, Debeer AE, Lacerot G, De Meester L, Vyverman W. (2011). Lack of phylogeographic structure in the freshwater cyanobacterium *Microcystis aeruginosa* suggests global dispersal. *PLoS One* **6**: e19561.
- Verschoor AM, van Dijk MA, Huisman J, van Donk E. (2013). Elevated CO₂ concentrations affect the elemental stoichiometry and species composition of an experimental phytoplankton community. *Freshw Biol* **58**: 597–611.
- Verspagen JMH, Passarge J, Jöhnk KD, Visser PM, Peperzak L, Boers P, Laanbroek HJ, Huisman J. (2006). Water management strategies against toxic *Microcystis* blooms in the Dutch delta. *Ecol Appl* **16**: 313–327.
- Verspagen JMH, Van de Waal DB, Finke JF, Visser PM., Huisman J. (2014a). Contrasting effects of rising CO₂ on primary production and ecological stoichiometry at different nutrient levels. *Ecol Lett* **17**: 951–960.
- Verspagen JMH, Van de Waal DB, Finke JF, Visser PM, Van Donk E, Huisman J. (2014b). Rising CO₂ levels will intensify phytoplankton blooms in eutrophic and hypertrophic lakes. *PLoS One* **9**: e104325.
- Větrovský T, Baldrian P. (2013). The variability of the 16S rRNA gene in bacterial genomes and its consequences for bacterial community analyses. *PLoS One* **8**: e57923.
- Vézie C, Rapala J, Vaitomaa J, Seitsonen J, Sivonen K. (2002). Effect of nitrogen and phosphorus on growth of toxic and nontoxic *Microcystis* strains and on intracellular microcystin concentrations. *Microbial Ecol* **43**: 443–454.
- Visser PM, Ibelings BW, Mur LR. (1995). Autumnal sedimentation of *Microcystis* spp. as result of an increase in carbohydrate ballast at reduced temperature. *J Plankt Res* **17**: 919–933.
- Visser PM, Ibelings BW, Van der Veer B, Koedood J, Mur LR. (1996). Artificial mixing prevents nuisance blooms of the cyanobacterium *Microcystis* in Lake Nieuwe Meer, The Netherlands. *Freshwat Biol* **36**: 435–450.
- Visser PM, Ibelings BW, Bormans M, Huisman J. (2015). Artificial mixing to control cyanobacterial blooms: a review. *Aquat Ecol* (in press). doi:10.1007/s10452-015-9537-0
- Visser PM, Passarge J, Mur LR. (1997). Modelling vertical migration of the cyanobacterium *Microcystis*. *Hydrobiol* **349**: 99–109.
- Von Wobeser EA, Ibelings BW, Bok J, Krasikov V, Huisman J, Matthijs HCP. (2011). Concerted changes in gene expression and cell physiology of the cyanobacterium *Synechocystis* sp. strain PCC 6803 during transitions between nitrogen and light-limited growth. *Plant Physiol* **155**: 1445–1457.
- Wacklin P, Hoffmann L, Komárek J. (2009). Nomenclatural validation of the genetically revised cyanobacterial genus *Dolichospermum* (Ralfs ex Bornet et Flahault) comb. nova. *Fottea* **9**: 59–64.
- Wagner C, Adrian R. (2009). Cyanobacteria dominance: quantifying the effects of climate change. *Limnol Oceanogr* **54**: 2460–2468.
- Wallace BB, Bailey MC, Hamilton DP. (2000). Simulation of vertical position of buoyancy regulating *Microcystis aeruginosa* in a shallow eutrophic lake. *Aquat Sci* **62**: 320–333.
- Walsby AE. (1994). Gas vesicles. *Microbiol Rev* **58**: 94–144.

- Walsby AE, Hayes PK, Boje R, Stal LJ. (1997). The selective advantage of buoyancy provided by gas vesicles for planktonic cyanobacteria in the Baltic Sea. *New Phytol* **136**: 407–417.
- Wang HL, Postier BL, Burnap RL. (2004). Alterations in global patterns of gene expression in *Synechocystis* sp. PCC 6803 in response to inorganic carbon limitation and the inactivation of *ndhR*, a LysR family regulator. *J Biol Chem* **279**: 5739–5751.
- Wang H, Sivonen K, Rouhiainen L, Fewer DP, Lyra C, Rantala-Ylinen A, Vestola J, Jokela J, Rantasärkkä K, Li Z, Liu B. (2012). Genome-derived insights into the biology of the hepatotoxic bloom-forming cyanobacterium *Anabaena* sp. strain 90. *BMC Genomics* **13**: 613.
- Wang X, Hao C, Zhang F, Feng C, Yang Y. (2011a). Inhibition of the growth of two-blue-green algae species (*Microcystis aruginosa* and *Anabaena spiroides*) by acidification treatments using carbon dioxide. *Bioresour Technol* **102**: 5742–5748.
- Wang Y, Duanmu D, Spalding MH. (2011b). Carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*: inorganic carbon transport and CO₂ recapture. *Photosynth Res* **109**: 115–122.
- Wannicke N, Endres S, Engel A, Grossart HP, Nausch M, Unger J, Voss M. (2012). Response of *Nodularia spumigena* to pCO₂. Part 1: Growth, production and nitrogen cycling. *Biogeosciences* **9**: 2973–2988.
- Waterhouse AM, Procter JB, Martin DM, Clamp M, Barton GJ. (2009). Jalview Version 2: a multiple sequence alignment editor and analysis workbench. *Bioinformatics* **25**: 1189–1191.
- Watson SB, Ridal J, Boyer GL. (2008). Taste and odour and cyanobacterial toxins: impairment, prediction, and management in the Great Lakes. *Can J Fish Aquat Sci* **65**: 1779–1796.
- Weiss R. (1974). Carbon dioxide in water and seawater: the solubility of a non-ideal gas. *Marine Chem* **2**: 203–215.
- Welker M, Von Döhren H. (2006). Cyanobacterial peptides – Nature's own combinatorial biosynthesis. *FEMS Microbiol Rev* **30**: 530–563.
- Wetzel RG, Likens GE. (2000). *Limnological Analyses, 3rd Ed.* New York: Springer-Verlag.
- Whitehead L, Long BM, Price GD, Badger MR. (2014). Comparing the *in vivo* function of α -carboxysomes and β -carboxysomes in two model cyanobacteria. *Plant Physiol* **165**: 398–411.
- Wiedner C, Rucker J, Brüggemann R, Nixdorf B. (2007). Climate change affects timing and size of populations of an invasive cyanobacterium in temperate regions. *Oecologia* **152**: 473–484.
- Wiedner C, Visser PM, Fastner J, Metcalf JS, Codd GA, Mur LR. (2003). Effects of light on the microcystin content of *Microcystis* strain PCC 7806. *Appl Environ Microbiol* **69**: 1475–1481.
- Wilhelm SW, Boyer GL. (2011). Healthy competition. *Nature Clim Change* **1**: 300–301.
- Wilson AE, Sarnelle O, Tillmanns AR. (2006). Effects of cyanobacterial toxicity and morphology on the population growth of freshwater zooplankton: Meta-analyses of laboratory experiments. *Limnol Oceanogr* **51**: 1915–1924.
- Wood JM. (1999). Osmosensing by bacteria: signals and membranebased sensors. *Microbiol Mol Biol R* **63**: 230–262.
- Woodger FJ, Badger MR, Price GD. (2003). Inorganic carbon limitation induces transcripts encoding components of the CO₂-concentrating mechanism in *Synechococcus* sp. PCC7942 through a redox-independent pathway. *Plant Physiol* **133**: 2069–2080.
- Woodger FJ, Badger MR, Price GD. (2005). Sensing of inorganic carbon limitation in *Synechococcus* PCC7942 is correlated with the size of the internal inorganic carbon pool and involves oxygen. *Plant Physiol* **139**: 1959–1969.
- Woodger FJ, Bryant DA, Price GD. (2007). Transcriptional regulation of the CO₂-concentrating mechanism in a euryhaline, coastal marine cyanobacterium, *Synechococcus* sp. Strain PCC 7002: role of NdhR/CcmR. *J Bacteriol* **189**: 3335–3347.
- Wu X, Kong F, Zhang M. (2011a). Photoinhibition of colonial and unicellular *Microcystis* cells in a summer bloom in Lake Taihu. *Limnology* **12**: 55–61.

- Wu X, Wu Z, Song L. (2011b). Phenotype and temperature affect the affinity for dissolved inorganic carbon in a cyanobacterium *Microcystis*. *Hydrobiologia* **675**: 175–186.
- Xu M, Bernát G, Singh A, Mi H, Rögner M, Pakrasi HB, Ogawa T. (2008). Properties of mutants of *Synechocystis* sp. strain PCC 6803 lacking inorganic carbon sequestration systems. *Plant Cell Physiol* **49**: 1672–1677.
- Xu H, Paerl HW, Qin B, Zhu G, Gao G. (2010). Nitrogen and phosphorus inputs control phytoplankton growth in eutrophic Lake Taihu, China. *Limnol Oceanogr* **55**: 420–432.
- Yamaguchi H, Suzuki S, Tanabe Y, Osana Y, Shimura Y, Ishida KI, Kawachi M. (2015). Complete genome sequence of *Microcystis aeruginosa* NIES-2549, a bloom-forming cyanobacterium from Lake Kasumigaura, Japan. *Genome Announc* **3**: e00551-15.
- Yamamoto Y, Tsukada H. (2009). Measurement of in situ specific growth rates of *Microcystis* (cyanobacteria) from the frequency of dividing cells. *J Phycol* **45**: 1003–1009.
- Yang C, Zhang W, Ren M, Song L, Li T, Zhao J. (2013). Whole-genome sequence of *Microcystis aeruginosa* TAIHU98, a nontoxic bloom-forming strain isolated from Taihu Lake, China. *Genome Announc* **1**: e00333–13.
- Ye J, Coulouris G, Zaretskaya I, Cutcutache I, Rozen S, Madden TL. (2012). Primer-BLAST: a tool to design target-specific primers for polymerase chain reaction. *BMC Bioinformatics* **13**: 134.
- Yeremenko N. (2004) Functional flexibility of photosystem I in cyanobacteria. PhD thesis, University of Amsterdam, the Netherlands.
- Yeremenko N, Kouril R, Ihalainen JA, D'Haene S, Van Oosterwijk N, Andrizhievskaya EG, Keestra W, Dekker HL, Hagemann M, Boekema EJ, Matthijs HCP, Dekker JP. (2004). Supramolecular organization and dual function of the IsiA chlorophyll-binding protein in cyanobacteria. *Biochemistry* **43**: 10308–10313.
- Yu J, Price GD, Badger MR. (1994a). A Mutant Isolated from the Cyanobacterium *Synechococcus* PCC7942 is unable to adapt to low inorganic carbon conditions. *Plant Physiol* **104**: 605–611.
- Yu J, Price GD, Badger MR. (1994b). Characterisation of CO₂ and HCO₃⁻ uptake during steady-state photosynthesis in the cyanobacterium *Synechococcus* PCC7942. *Funct Plant Biol* **21**: 185–195.
- Zehr JP, Carpenter EJ, Villareal TA. (2000). New perspectives on nitrogen-fixing microorganisms in tropical and subtropical oceans. *Trends Microbiol* **8**: 68–73.
- Zhang P, Allahverdiyeva Y, Eisenhut M, Aro EM. (2009). Flavodiiron proteins in oxygenic photosynthetic organisms: photoprotection of photosystem II by Flv2 and Flv4 in *Synechocystis* sp. PCC 6803. *PLoS One* **4**: e5331.
- Zhang P, Eisenhut M, Brandt AM, Carmel D, Silen HM, Vass I, Allahverdiyeva Y, Salminen TA, Aro EM. (2012). Operon *flv4-flv2* provides cyanobacterial photosystem II with flexibility of electron transfer. *Plant Cell* **24**: 1952–1971.
- Zilliges Y, Kehr JC, Meissner S, Ishida K, Mikkat S, Hagemann M, Kaplan A, Börner T, Dittmann E. (2011). The cyanobacterial hepatotoxin microcystin binds to proteins and increases the fitness of *Microcystis* under oxidative stress conditions. *PLoS One* **6**: e17615.
- Zurawell RW, Chen H, Burke JM, Prepas EE. (2005). Hepatotoxic cyanobacteria: A review of the biological importance of microcystins in freshwater environments. *J Toxicol Environ Health B* **8**: 1–37.



**List of frequently used abbreviations,
symbols and genes**

| | |
|---|---|
| 2PG | 2-phosphoglycerate |
| 3PG | 3-phosphoglycerate |
| ATP | adenosine triphosphate |
| BCT1 | ATP-dependent bicarbonate uptake system (high affinity, low flux) |
| <i>bicA</i> | gene encoding sodium-dependent bicarbonate uptake system BicA |
| BicA | sodium-dependent bicarbonate uptake system (low affinity, high flux) |
| CA | carbonic anhydrase |
| CCM | CO ₂ -concentrating mechanism |
| <i>ccmR</i> | gene encoding transcriptional regulator CcmR |
| <i>ccmR2</i> | gene encoding transcriptional regulator CcmR2 |
| cDNA | complementary DNA, synthesized from messenger RNA |
| C _i | inorganic carbon |
| Chl <i>a</i> | chlorophyll <i>a</i> (pigment) |
| <i>chpY</i> (= <i>cupA</i>) | gene encoding CO ₂ hydration subunit of CO ₂ uptake system NDH-I ₃ |
| <i>chpX</i> (= <i>cupB</i>) | gene encoding CO ₂ hydration subunit of CO ₂ uptake system NDH-I ₄ |
| <i>cmpA</i> | gene encoding bicarbonate-binding subunit of uptake system BCT1 |
| CO ₂ (aq) | carbon dioxide dissolved in water |
| CO ₃ ²⁻ | carbonate |
| DIC | dissolved inorganic carbon |
| DOC | dissolved organic carbon |
| HCO ₃ ⁻ | bicarbonate |
| <i>g</i> _{CO2} | CO ₂ gas influx |
| gDNA | genomic DNA |
| <i>flv1-4</i> | genes encoding flavodiiron proteins, involved in oxidative stress protection |
| <i>I</i> / <i>I</i> _{in} / <i>I</i> _{out} | irradiance / incident irradiance / irradiance penetrating through vessel |
| <i>isiA</i> | gene encoding iron stress-induced protein |
| <i>K</i> _{0.5} | half-saturation constant |
| <i>K</i> _H | solubility constant of CO ₂ gas in water |
| MC | microcystin (hepatotoxin) |
| <i>mcyB</i> | microcystin synthetase gene |
| NADPH | nicotinamide adenine dinucleotide phosphate |
| NDH-I ₃ | CO ₂ uptake system (high affinity, low flux) |
| NDH-I ₄ | CO ₂ uptake system (low affinity, high flux) |
| <i>nhaS1-6</i> | genes encoding sodium/proton antiporters |
| PAR | photosynthetically active radiation, spectral range used for photosynthesis |
| pCO ₂ | partial pressure of CO ₂ |

| | |
|-------------|--|
| PKS | polyketide synthetase |
| ppm | parts per million |
| PSI | photosystem I |
| PSII | photosystem II |
| <i>rbcX</i> | gene encoding RuBisCO chaperone |
| ROS | reactive oxygen species, chemically reactive molecules containing oxygen |
| RT-qPCR | reverse transcription quantitative polymerase chain reaction |
| RuBisCO | ribulose-1,5-bisphosphate carboxylase/oxygenase |
| <i>sbtA</i> | gene encoding bicarbonate uptake system SbtA (high affinity, low flux) |
| SbtA | sodium-dependent bicarbonate uptake system (high affinity, low flux) |
| <i>sbtB</i> | gene found downstream of <i>sbtA</i> , associated with <i>sbtA</i> |
| <i>v</i> | gas transfer velocity (piston velocity) across the air-water interface |



Summary

Effects of rising CO₂ on the harmful cyanobacterium *Microcystis*

Harmful cyanobacteria ('blue-green algae') are notorious for causing worldwide ecological and economical problems in eutrophic lakes and reservoirs, where they can produce dense and often toxic blooms. Climate change is foreseen to have large effects on these photosynthetic microorganisms. Yet, while several studies have investigated effects of global warming on harmful cyanobacteria, the implications of rising CO₂ have received relatively little attention. Cyanobacteria are often assumed to be favored at low inorganic carbon conditions, because of the presence of an effective CO₂-concentrating mechanism (CCM) to fix CO₂. But how will they perform at elevated CO₂ levels? This thesis investigates the impact of elevated CO₂ on various strains of the ubiquitous harmful cyanobacterium *Microcystis aeruginosa*.

The following questions are addressed:

- 1) How variable are the CCMs within the genus *Microcystis*? (**Chapter 2**)
- 2) What are the adaptations of *Microcystis* to elevated CO₂? (**Chapter 3**)
- 3) What are the similarities and differences in CCM gene expression (1) among *Microcystis* strains, and (2) between *Microcystis* and other cyanobacteria? (**Chapters 2 and 4**)
- 4) How are the *Microcystis* CCM genes regulated *in situ*? (**Chapter 6**)
- 5) Can genetic variability of the *Microcystis* CCM cause strain-specific differences in growth rate at elevated CO₂ concentrations? (**Chapters 2 and 4**)
- 6) Does rising CO₂ affect the competition between different *Microcystis* strains, and if so, which strains will benefit most? (**Chapter 5**)
- 7) What are the effects of rising CO₂ on other harmful cyanobacteria? (**Chapter 7**)
- 8) Will rising CO₂ concentrations stimulate cyanobacterial blooms and make them more toxic? (**Chapters 2, 3, 5 and 7**)
- 9) Is potassium ion addition an effective method to combat harmful cyanobacterial blooms? (**Chapter 8**)

To investigate the first question, 20 *Microcystis* strains from different continents were studied at the gene level. Cyanobacteria often use a combination of CO₂ and bicarbonate uptake systems to import inorganic carbon (C_i). Genes encoding the two CO₂ uptake systems, the ATP-dependent bicarbonate transporter BCT1, the CO₂-fixing enzyme RuBisCO and carboxysomes (compartments containing RuBisCO) were detected in all 20 *Microcystis* strains. Eight of the analyzed strains also contain the genes *bicA* and *sbtA*, encoding the sodium-dependent bicarbonate uptake systems BicA and SbtA, respectively. BicA has a low affinity for bicarbonate

and high flux rate, whereas SbtA has a high affinity and low flux rate. Affinity refers to the effectiveness of bicarbonate uptake at low bicarbonate concentrations, whereas the flux rate refers to the bicarbonate uptake rate at high bicarbonate concentrations. A unique feature of these *Microcystis* strains is that the genes *bicA* and *sbtA* are present in one operon and are co-transcribed. In contrast to these C_i uptake generalists, 12 of the 20 analyzed *Microcystis* strains lack either the *bicA* or *sbtA* gene. The results show that *Microcystis* strains have adapted differently to the wide natural variation in CO_2 concentrations.

The second question was answered in chemostat experiments with *Microcystis* PCC 7806. Changes in the transcriptome (expression of all genes in the genome) were monitored from 45 minutes up to 2 weeks after increasing the CO_2 concentration. Surprisingly, elevated CO_2 affected the expression of only a small number of genes. The bicarbonate uptake genes were downregulated at elevated CO_2 . Other regulated genes were involved in the stress response of the cells, control of the cellular C/N ratio, and the production of two weakly characterized polyketides. Expression of genes encoding the CO_2 uptake systems, carboxysome, RuBisCO, photosystems, C metabolism and microcystin synthetases did not respond significantly to elevated CO_2 .

The third question was answered by exposing batch cultures of six different *Microcystis* strains to elevated CO_2 . The high-affinity gene *cmpA* (encoding a subunit of the bicarbonate uptake system BCT1) was downregulated at elevated CO_2 in all strains. Most strains also downregulated *bicA* and *sbtA* at elevated CO_2 , but two strains showed constitutive expression of these bicarbonate uptake genes. The high-flux BicA uptake system remained active at high CO_2 levels in all strains containing the *bicA* gene. Interestingly, expression of the high- and low-affinity CO_2 uptake genes of *Microcystis* was not affected by elevated CO_2 , which deviates from most other cyanobacterial species that downregulate the high-affinity CO_2 uptake genes. The carboxysome and RuBisCO genes were also constitutively expressed in all *Microcystis* strains. We discovered a new CCM transcriptional regulator gene (*ccmR2*), located upstream of the *bicA-sbtA* operon. Both *ccmR2* and the *bicA-sbtA* operon are so far unique for *Microcystis*.

The fourth question was answered with an *in situ* study at Lake Kennemermeer (the Netherlands) of a cyanobacterial bloom that contained *Microcystis*. The lake showed large diel fluctuations in bicarbonate, pH and dissolved oxygen as a consequence of the photosynthetic activity of the bloom. Expression of the bicarbonate uptake genes of *Microcystis* was tuned to the diel variation in bicarbonate concentration. In contrast, expression of the CO_2 uptake genes was constitutive, and expression of the RuBisCO and carboxysomal genes was slightly increased during nighttime.

To address the fifth question, a series of laboratory experiments were carried out in batch culture at different CO₂ concentrations. The results showed that strains with the high-affinity gene *sbtA* perform better at low C_i concentrations, strains with the high-flux gene *bicA* perform better at high C_i concentrations, and *bicA+sbtA* strains containing both genes perform well across the entire range of C_i conditions investigated.

The sixth question was addressed by investigating mixtures with multiple *Microcystis* strains in laboratory competition experiments and a lake study. The competition experiments and lake study both showed that strains with the high-flux gene *bicA* have a selective advantage at elevated C_i levels. These results provide laboratory and field evidence that changes in C_i availability induce rapid adaptive changes in the genotype composition of harmful cyanobacterial blooms. Hence, future cyanobacterial blooms may have a genetic composition that differs from contemporary blooms.

The seventh question was answered by analyzing recently sequenced genomes of other harmful cyanobacteria, including *Anabaena*, *Aphanizomenon* and *Planktothrix* strains. These cyanobacteria also showed intraspecific variation in the presence of the *bicA* and *sbtA* genes, similar to *Microcystis*, suggesting that they are well adapted to a wide range of CO₂ conditions. However, in *Anabaena*, *Aphanizomenon* and *Planktothrix*, *bicA* and *sbtA* are not organized in one operon, and in some strains both *bicA* and *sbtA* are absent. Presumably, these harmful cyanobacteria display a similar phenotypic variation as *Microcystis*, with a selective advantage for strains with the high-affinity uptake systems SbtA and/or BCT1 at low C_i conditions and a selective advantage for strains with the high-flux uptake system BicA at high C_i conditions.

To address the eighth question, it was shown that there was no direct connection between the presence of the C_i uptake genes *bicA* or *sbtA* and the microcystin synthetase genes. In chemostat experiments with *Microcystis* PCC 7806, elevated CO₂ levels led to a shift from carbon- to light-limited conditions. The strain contained ~2.5 times more unbound microcystins per cell at elevated CO₂, indicating that the cells can become more toxic at elevated CO₂ levels. Biomass of this strain increased strongly at elevated CO₂, suggesting that cyanobacterial blooms will intensify in eutrophic lakes. Furthermore, the dry weight of the cells was reduced twofold, indicating that elevated CO₂ can promote buoyancy of the cells, and thus scum layer formation in lakes. Hence, elevated CO₂ is foreseen to worsen the problems with *Microcystis* blooms in eutrophic lakes.

The ninth question was answered by investigating the potassium ion sensitivity of selected laboratory *Microcystis* strains. Strain PCC 7806, originating from brackish water, was not affected by the increased potassium ion concentration, while the growth of two freshwater *Microcystis* strains was strongly reduced. The potassium ion sensitivity of the freshwater strains was linked to the absence of specific salt tolerance genes. These results show that the salt

tolerance and potassium sensitivity of *Microcystis* differ between strains. Hence, on the short run, potassium ion addition might be a successful remediation strategy to combat *Microcystis* blooms in freshwater lakes, but over time more tolerant *Microcystis* strains or other cyanobacteria are likely to become dominant.

This thesis contributes to a better understanding of how harmful cyanobacteria respond to climate change. The results show how cyanobacteria subtly adjust their cells at the molecular and physiological level to changes in C_i availability. Furthermore, the results demonstrate how genetic diversity in the C_i uptake systems provide cyanobacteria with the potential for rapid microevolutionary adaptation to changes in CO_2 conditions, with a selective advantage for strains with the high-flux bicarbonate uptake gene *bicA* at elevated CO_2 levels. Hence, one of the key lessons of this work is that future studies of climate change effects should keep in mind the large genetic and physiological variation within species. In total, the results indicate that a further rise of the atmospheric CO_2 concentration is likely to increase the frequency and intensity of cyanobacterial blooms in eutrophic waters, and possibly may also increase their toxicity. The predicted intensification of cyanobacterial blooms should be countered by the reduction of CO_2 emissions and the development of effective methods to combat and prevent harmful cyanobacterial blooms.



Samenvatting

Effecten van stijgend CO₂ op de schadelijke cyanobacterie *Microcystis*

Schadelijke cyanobacteriën ('blauwalgen') zijn bekende veroorzakers van wereldwijde ecologische en economische problemen in oppervlaktewateren zoals voedselrijke meren, waar ze dichte en vaak toxische bloeien kunnen vormen. Het is voorzien dat klimaatverandering grote effecten zal hebben op deze fotosynthetische microorganismen. Terwijl verschillende studies de effecten van stijgende temperaturen hebben onderzocht, hebben de mogelijke gevolgen van stijgende CO₂-concentraties tot nog toe weinig aandacht gekregen. Vaak wordt aangenomen dat cyanobacteriën voordeel hebben bij lage CO₂-condities, door de aanwezigheid van een effectief CO₂-concentreermechanisme (CCM) om CO₂ te fixeren. Maar hoe presteren ze bij verhoogde CO₂-niveaus? Dit proefschrift onderzoekt de effecten van stijgende CO₂-concentraties op verschillende stammen van de wijdverspreide schadelijke cyanobacterie *Microcystis aeruginosa*.

De volgende vragen worden behandeld:

- 1) Hoe variabel zijn de CCM's binnen het genus *Microcystis*? **(Hoofdstuk 2)**
- 2) Wat zijn de aanpassingen van *Microcystis* bij verhoogde CO₂-concentraties? **(Hoofdstuk 3)**
- 3) Wat zijn de overeenkomsten en verschillen in CCM-genexpressie (1) tussen *Microcystis* stammen onderling, en (2) tussen *Microcystis* en andere cyanobacteriën? **(Hoofdstukken 2 en 4)**
- 4) Hoe worden de CCM-genen van *Microcystis* gereguleerd? **(Hoofdstuk 6)**
- 5) Kan genetische variatie in de CCM's leiden tot verschillen in groeisnelheid van *Microcystis* stammen bij verhoogde CO₂-concentraties? **(Hoofdstukken 2 en 4)**
- 6) Heeft stijgende CO₂ invloed op de competitie tussen verschillende *Microcystis* stammen, en zo ja, welke stammen hebben het meeste voordeel? **(Hoofdstuk 5)**
- 7) Wat zijn de effecten van stijgend CO₂ op andere schadelijke cyanobacteriën? **(Hoofdstuk 7)**
- 8) Zullen stijgende CO₂-concentraties bloeien van schadelijke cyanobacteriën stimuleren en hun toxiciteit verhogen? **(Hoofdstukken 2, 3, 5 en 7)**
- 9) Is toevoeging van kaliumionen een effectieve methode om schadelijke cyanobacteriën te bestrijden? **(Hoofdstuk 8)**

Om de eerste vraag te onderzoeken zijn 20 *Microcystis* stammen uit verschillende delen van de wereld op genniveau onderzocht. Cyanobacteriën gebruiken vaak een combinatie van CO₂- en bicarbonaat-opnamesystemen om anorganische koolstof (C_i) te importeren in hun cellen. Alle

20 stammen bevatten genen die coderen voor de twee CO₂-opnamesystemen, de ATP-afhankelijke bicarbonaattransporter BCT1, het CO₂-fixatie-enzym RuBisCO en carboxyzomen (compartimenten die RuBisCO bevatten). Acht van de onderzochte stammen hadden ook de genen *bicA* en *sbtA*, die coderen voor de natrium-afhankelijke bicarbonaatopnamesystemen BicA en SbtA. BicA heeft een lage affiniteit voor bicarbonaat en een hoge opnamesnelheid, terwijl SbtA een hoge affiniteit heeft en een lage opnamesnelheid. Met affiniteit wordt hier bedoeld de effectiviteit van bicarbonaatopname bij lage bicarbonaatconcentraties, terwijl de opnamesnelheid refereert naar de bicarbonaatopname bij hoge bicarbonaatconcentraties. Een unieke eigenschap van deze *Microcystis* stammen is dat de genen *bicA* en *sbtA* zich bevinden op hetzelfde operon en dus gezamenlijk worden getranscribeerd (co-transcriptie). In tegenstelling tot deze C₁-opnamegeneralisten, misten 12 van de 20 *Microcystis* stammen het *bicA* of *sbtA* gen. De resultaten laten zien dat *Microcystis* stammen zich op verschillende wijze hebben aangepast aan de natuurlijke variatie in CO₂-concentraties.

De tweede vraag is beantwoord met behulp van chemostaatexperimenten met *Microcystis* PCC 7806. Veranderingen in het transcriptoom (expressie van alle genen in het genoom) van deze stam werden bekeken van 45 minuten tot 2 weken na het verhogen van de CO₂-concentratie. De verhoogde CO₂-concentratie had slechts effect op de expressie van een verrassend klein aantal genen. De bicarbonaatopnamegenen waren omlaag gereguleerd bij verhoogde CO₂. Andere gereguleerde genen waren betrokken bij de stress respons van cellen, controle van de cellulaire C/N ratio en de productie van twee nader te karakteriseren polyketides. De verhoogde CO₂-concentratie leidde niet tot significante veranderingen in genexpressie van de CO₂-opnamesystemen, het carboxysoom, RuBisCO, de fotosystemen, het C metabolisme en de microcystine synthetases.

De derde vraag is beantwoord door batch culturen met zes verschillende *Microcystis* stammen bloot te stellen aan verhoogde CO₂-concentraties. Het hoge-affiniteit-gen *cmpA* (coderend voor een subunit van het bicarbonaatopnamesysteem BCT1) werd in alle stammen omlaag gereguleerd. De meeste stammen verlaagden ook de genexpressie van *bicA* en *sbtA* bij verhoogde CO₂-concentraties, terwijl in twee stammen de expressie van deze bicarbonaatopnamegenen niet werd aangepast. Het hogesnelheidsopnamesysteem BicA bleef actief bij hoge CO₂-concentraties in alle stammen met het *bicA* gen. Opvallend genoeg werd de expressie van de hoge- en lage-affiniteit-CO₂-opnamegenen van *Microcystis* niet beïnvloed door verhoogde CO₂-concentraties, in tegenstelling tot de meeste andere cyanobacteriën die de expressie van hun hoge-affiniteit-CO₂-opnamegenen verlagen. Expressie van de carboxysoom en RuBisCO genen werd in geen van de *Microcystis* stammen aangepast. We ontdekten een nieuw CCM transcriptioneel regulatorgen (*ccmR2*), dat zich stroomopwaarts van het *bicA-sbtA* operon bevindt. Zowel *ccmR2* als het *bicA-sbtA* operon zijn tot nu toe uniek voor *Microcystis*.

De vierde vraag is beantwoord met een veldstudie in het Kennemermeer (Nederland) van een cyanobacteriële bloei met *Microcystis*. Het meer vertoonde grote dagelijkse schommelingen in bicarbonaat, pH en opgelost zuurstof als een gevolg van de fotosynthese-activiteit van de bloei. Expressie van de bicarbonaatopnamegenen van *Microcystis* was afgestemd op de dagelijkse variatie in de bicarbonaatconcentratie. Expressie van de CO₂-opnamegenen vertoonde echter geen dagelijkse variatie en expressie van de RuBisCO- en carboxysoomgenen was iets verhoogd tijdens de nacht.

Om de vijfde vraag te onderzoeken zijn een serie van laboratoriumexperimenten uitgevoerd in batch-culturen bij verschillende CO₂-concentraties. De resultaten laten zien dat stammen met het hoge-affiniteit-gen *sbtA* beter presteren bij lage C_i-concentraties, stammen met het hogesnelheids-gen *bicA* beter presteren bij hoge C_i-concentraties, terwijl stammen die beschikken over beide genen goed presteren over het hele bereik van onderzochte C_i-condities.

De zesde vraag is onderzocht door mengsels van verschillende *Microcystis* stammen te bestuderen in laboratorium competitie-experimenten en in een veldstudie van een cyanobacteriële bloei in een eutroof meer. De competitie-experimenten en de veldstudie laten beide zien dat stammen met het hogesnelheids-gen *bicA* een selectief voordeel hebben bij verhoogde C_i-concentraties. De resultaten leveren zowel experimenteel als observationeel bewijs dat veranderingen in de C_i beschikbaarheid leiden tot snelle adaptieve veranderingen in de genotypesamenstelling van schadelijke cyanobacteriën. Dit suggereert dat cyanobacteriële bloeien in de toekomst waarschijnlijk een genetische samenstelling zullen hebben die verschilt van de huidige bloeien.

De zevende vraag is beantwoord door recent gesequencede genomen van andere schadelijke cyanobacteriën te analyseren, waaronder *Anabaena*, *Aphanizomenon* en *Planktothrix* stammen. Deze cyanobacteriën lieten ook intraspecifieke variatie zien in de aanwezigheid van *bicA* en *sbtA*, net als bij *Microcystis*, wat suggereert dat ze goed zijn aangepast aan een wijde range van CO₂-condities. Echter, in *Anabaena*, *Aphanizomenon* en *Planktothrix* zijn *bicA* en *sbtA* niet ondergebracht in hetzelfde operon, en in sommige stammen zijn zowel *bicA* als *sbtA* afwezig. Waarschijnlijk laten deze schadelijke cyanobacteriën een vergelijkbare fenotypische variatie zien als *Microcystis*, met een selectief voordeel voor stammen met de hoge-affiniteit-opnamesystemen SbtA en/of BCT1 bij lage C_i-concentraties en een selectief voordeel voor stammen met het hogesnelheids opname systeem BicA bij hoge C_i-concentraties.

De achtste vraag is beantwoord door aan te tonen dat er geen direct verband is tussen de aanwezigheid van de C_i-opnamegenen *bicA* of *sbtA* en de microcystine-synthetase-genen. In chemostaatexperimenten met *Microcystis* PCC 7806 leidde verhoogde CO₂-concentraties tot een verschuiving van koolstof-gelimiteerde naar licht-gelimiteerde condities. De stam bevatte ~2.5 keer meer ongebonden microcystine per cel bij verhoogde CO₂-concentraties, wat

aangeeft dat de cellen toxischer kunnen worden bij stijging van het CO₂-niveau. De biomassa van de stam was sterk toegenomen bij verhoogde CO₂-concentraties, wat suggereert dat cyanobacteriële bloeien in eutrofe meren intenser worden. Daarnaast was het drooggewicht van de cellen gehalveerd, wat doet vermoeden dat verhoogde CO₂-concentraties het drijfvermogen van cellen kan bevorderen en daarmee de vorming van drijfslagen in meren kan stimuleren. Deze resultaten geven aan dat stijgende CO₂-concentraties de problemen met *Microcystis* in eutrofe meren waarschijnlijk zullen verergeren.

De negende vraag is beantwoord door onderzoek naar de kaliumgevoeligheid van geselecteerde *Microcystis* stammen. Stam PCC 7806, die oorspronkelijk is geïsoleerd uit brak water, werd niet beïnvloed door verhoogde kalium concentraties, terwijl de groei van twee zoetwater *Microcystis* stammen sterk gereduceerd werd. De kaliumgevoeligheid van de zoetwater stammen lijkt samen te hangen met de afwezigheid van specifieke genen betrokken bij zoutwatertolerantie. Dit laat zien dat *Microcystis* stammen verschillen in hun zoutwatertolerantie en kaliumgevoeligheid. Het gevolg is dat kaliumtoevoeging op korte termijn een succesvolle beheersmethode kan zijn om *Microcystis* bloeien in zoet water te bestrijden, maar dat op de langere termijn waarschijnlijk minder kaliumgevoelige *Microcystis* stammen of andere tolerante cyanobacteriën de dominantie zullen overnemen.

Dit proefschrift draagt bij aan een beter begrip van hoe schadelijke cyanobacteriën reageren op klimaatverandering. De resultaten laten zien hoe cyanobacteriën hun cel op moleculair en fysiologisch niveau op subtiele wijze kunnen aanpassen aan de C_i beschikbaarheid. Ook laten de resultaten zien dat de genetische diversiteit in C_i-opnamesystemen van cyanobacteriën kan leiden tot een snelle micro-evolutie, waarbij de genotype samenstelling van de bloei zich aanpast aan veranderingen in de CO₂-concentratie. Hierbij hebben stammen met het hogesnelheids-bicarbonaatopnamegen *bicA* een selectief voordeel bij hoge C_i-concentraties. Een van de belangrijkste lessen van dit werk is daarom dat toekomstige studies naar de effecten van klimaatsverandering attent moeten zijn op de genetische en fysiologische variatie binnen soorten. Alles samenvattend geven de resultaten aan dat verdere stijging van de CO₂-concentratie waarschijnlijk zal leiden tot een toename van de frequentie en intensiteit van cyanobacteriële bloeien in eutrofe wateren, en mogelijk ook de toxiciteit van deze bloeien zal verhogen. De voorspelde toename van cyanobacteriële bloeien zou kunnen worden beperkt door reductie van de CO₂-uitstoot en door de verdere ontwikkeling van effectieve methoden om schadelijke cyanobacteriën te bestrijden.



Acknowledgements

Luctor et Emergo

Ik worstel en kom boven

After several years of hard work it is finally done! Time went by very fast and so much changed since I started working here at the Science Park of Amsterdam in February 2011. I really enjoyed being part of the Aquatic Microbiology (AMB) group and I learned a lot. Yet, I could not have done the work presented in my thesis without the help of several other people. Also many people made my stay at the University of Amsterdam more pleasant, or supported me outside working hours.

First I would like to thank Jef. As my promoter, supervisor of the TOP project and group leader, you guided my research into the right direction and I really learned a lot from you. With my molecular background, ecology was a relatively new aspect for me, and I thank you for introducing me to this field. I also admire your aim for perfection, and your enthusiasm for and dedication to the research of this thesis.

Of course I also want to thank Hans. As my co-promoter and daily supervisor, we had a lot of meetings and discussions. I thank you for introducing me to the worlds of cyanobacteria and photosynthesis. I really like that you granted me a lot of freedom during the last years. Your biochemistry background was very useful for the fine-tuning of experiments and for bright interpretations of results, and your large network of people also proved to be very useful. You really were a source of inspiration during the project.

Next, I would like to thank Jolanda. In the beginning of the project I learned a lot from you about *Microcystis* and inorganic carbon chemistry. Although we were both involved in the TOP project, theoretical modeling and molecular work in the lab did not always overlap. However, during several occasions exchanging knowledge proved to be very useful.

I am also grateful to Pieter and Bas. Without you, the labs would turn into chaos. I know that sometimes I asked for seemingly impossible things, but in the end everything turned out fine. I really enjoyed the fieldwork in 2013 with you and other colleagues, especially the adventures with the lab boat (and in early stages the Intertoys boat).

I also want to thank Amanda. We started with our PhD at about the same time at the AMB group. Although we worked on very different projects, parts of our research sometimes overlapped and we could help each other out. Your presence in the 'PhD room' and in the canteen was really enjoyable. The after-work activities together with other people from the group were also really fun. I will definitely miss you and the Thanksgiving dinners on your house boat!

Next I would like to thank my former master students, Serena, Robert and Dennis. I thank Serena for all the hard work and many analyses during the chemostat experiment with *Microcystis* PCC 7806. Thanks to your accuracy and positive spirit, the experiment really was a success. I thank Robert for all the efforts during the fieldwork in 2013. We set out with good and bad weather, and we sometimes encountered lakes without blooms, and other times lakes with a stinky *Microcystis* scum layer, dead fish and occasionally even a dead rabbit. Especially the 24-hour experiment was hard work with hardly any sleep, but in the end it was also quite fun and the results turned out to be really nice. I thank Dennis for maintaining and analyzing numerous *Microcystis* cultures and generating a ton of data. Your organizational skills and persistence really made this part of the research a success. I wish you all success in your future lives.

I want to thank Merijn and Jason for the pleasant collaborations during the last years. I would also like to thank the other (former) members of the AMB group, Gerard, Petra, Maayke, Pascale, Corrien, Michael, Elisa, Quan-Xing, Fleur, Anouk, Verena, Joost, Suzanne, Dina, Ruben, Tom V, Tom B, Veerle, Emily, Lex, Muhe, Catarina, Anne-Catherine, Charlotte, Cherele, Tim B, Tim P, Erik, Estelle, Qian, and students that did an internship in the group, for support during the project and the pleasant time during and after work hours. The AMB group has really changed and expanded a lot since I started. I really enjoyed the visits to the Polder and Brouwerij 't IJ, other 'borrels', the soccer/volleyball during summer days after work, and the stories or discussions during lunch time. I am sure I will see several of you around in the future.

I would like to thank the Netherlands Organisation for Scientific Research (NWO) for funding this project. I would like to thank Leo Hoitinga for assistance with the analysis of the numerous DIC samples. I want to express my gratitude to Gertjan Bon of the UvA glass workshop for constructing several essential glass components for the experiments, including the large 1m columns for the NaOH and silica pellets, the chemostat vessels, and the glass O₂ optode vessels. It really takes great skill to create these glass items.

I would like to thank my previous classmates of Life Science & Technology, Geert, Martijn, Bas, Régis, Ruben and the mathematician Floris, for the amusing “Halve Liter Woensdag” gatherings, which kept me going. I would like to thank the people of the Slagwerkgroep de Vliegende Hollander Terneuzen for the fun times on Friday evenings in the early phase of my PhD. I would like to thank my housemates for making my stay in Leiden more pleasant. I would like to thank my family, Menno, Mieke, Nydia, Emanuel, as well as my grandparents, for their support. Finally, I would like to thank Deborah for support during the last phase of my PhD and making my life more meaningful.



Curriculum Vitae

Giovanni Sandrini was born on the 16th of June 1986, in Terneuzen, the Netherlands. After completing secondary school at the Zeldenrust-Steelant College in Terneuzen in 2004, he completed the bachelor and master program of Life Science & Technology organized by a collaboration between Leiden University and Delft University of Technology. During his bachelor internship he worked on the isolation and genetic analysis of novel streptomycetes from soil samples, in the Microbial Development group of prof. dr. Gilles van Wezel at Leiden University. During his masters he did his industrial internship at the Netherlands Organisation for Applied Scientific Research (TNO) in Delft, in the group of dr. Harald J. Ruijsenaars, where he used genetic engineering on the solvent tolerant soil bacterium *Pseudomonas putida* S12 to allow it to grow on lignocellulose hydrolysate. He did his master research internship at the nidrovirus group of prof. dr. Eric J. Snijder, at the Leiden University Medical Center (LUMC), under supervision of dr. Clara C. Poshuma and dr. Sjoerd van den Worm, where he investigated the presence of an RNA proofreading mechanism in coronaviruses. He graduated in 2010, with the major Functional Genomics. Early 2011, he started his PhD in the group of Aquatic Microbiology of prof. dr. Jef Huisman at the University of Amsterdam, under supervision of dr. Hans C.P. Matthijs. The work of his PhD is described in this thesis and resulted in several publications. He also presented the results of this thesis at national and international conferences.

Publications

- Sandrini G**, Matthijs HCP, Verspagen JMH, Muyzer G, Huisman J. (2014). Genetic diversity of inorganic carbon uptake systems causes variation in CO₂ response of the cyanobacterium *Microcystis*. *ISME Journal* **8**: 589–600.
- Sandrini G**, Cunsolo S, Schuurmans JM, Matthijs HCP, Huisman J. (2015). Changes in gene expression, cell physiology and toxicity of the harmful cyanobacterium *Microcystis aeruginosa* at elevated CO₂. *Frontiers in Microbiology* **6**: 401.
- Sandrini G**, Huisman J, Matthijs HCP. (2015). Strain-specific potassium ion sensitivity of the harmful cyanobacterium *Microcystis* correlates with the prevalence of specific salt tolerance genes. *FEMS Microbiology Letters* **362**: fmv121.
- Sandrini G**, Jakupovic D, Matthijs HCP, Huisman J. (2015). Strains of the harmful cyanobacterium *Microcystis aeruginosa* differ in gene expression and activity of inorganic carbon uptake systems at elevated CO₂ levels. *Applied and Environmental Microbiology* **81**: 7730–7739.
- Sandrini G**, Ji X, Verspagen JMH, Tann RP, Slot PC, Luimstra VM, Schuurmans JM, Matthijs HCP, Huisman J. Rapid microevolutionary adaptation of harmful cyanobacteria to changes in CO₂ availability. (submitted manuscript)
- Sandrini G**, Tann RP, Schuurmans JM, Van Beusekom SAM, Matthijs HCP, Huisman J. Diel variation of gene expression of the CO₂-concentrating mechanism during a harmful cyanobacterial bloom. (submitted manuscript)
- Visser PM, Verspagen JMH, **Sandrini G**, Stal LJ, Matthijs HCP, Davis TW, Paerl HW, Huisman J. (2016). How rising CO₂ and global warming may stimulate harmful cyanobacterial blooms. *Harmful Algae* (in press)

Attended conferences

- ESF-EMBO symposium on Molecular Bioenergetics of Cyanobacteria: From Cells to Community. Sant Feliu de Guixols, Spain, April 2011.
- 14th International Symposium on Phototrophic Prokaryotes. Porto, Portugal, August 2012.
- 15th International Symposium on Microbial Ecology. Seoul, South-Korea, August 2014.
- 9th European Workshop on the Molecular Biology of Cyanobacteria. Texel, the Netherlands, September 2014.
- 15th International Symposium on Phototrophic Prokaryotes. Tübingen, Germany, August 2015.