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Identification of Three Mannoproteins in the Cell Wall of *Saccharomyces cerevisiae*

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Three glucanase-extractable cell wall proteins from *Saccharomyces cerevisiae* were purified, and their N-terminal amino acid sequences were determined. With this information, we were able to assign gene products to three known open reading frames (ORFs). The N-terminal sequence of a 55-kDa mannoprotein corresponded with the product of ORF YKL096w, which we named *CWP1* (cell wall protein 1). A 80-kDa mannoprotein was identified as the product of the *TIP1* gene, and a 180-kDa mannoprotein corresponded to the product of the ORF YKL444, which we named *CWP2*. *CWP1*, *TIP1*, and *CWP2* encode proteins of 239, 210, and 92 amino acids, respectively. The C-terminal regions of these proteins all consist for more than 40% of serine/threonine and contain putative glycosylphosphatidylinositol attachment signals. Furthermore, *Cwp1p* and *Tip1p* were shown to carry a β1,6-glucose-containing side chain. The *cwp2* deletion mutant displayed an increased sensitivity to Congo red, calcifluor white, and Zymolyase. Electron microscopic analysis of the *cwp2* deletion mutant showed a strongly reduced electron-dense layer on the outside of the cell wall. These results indicate that *Cwp2p* is a major constituent of the cell wall and plays an important role in stabilizing the cell wall. Depletion of *Cwp1p* or *Tip1p* also caused increased sensitivities to Congo red and calcifluor white, but the effects were less pronounced than for *cwp2Δ*. All three cell wall proteins show a substantial homology with *Srplp*, which also appears to be localized in the cell wall. We conclude that these four proteins are small structurally related cell wall proteins.

The two major components of the cell wall of the yeast *Saccharomyces cerevisiae* are glucan, which constitutes the inner layer of the cell wall, and mannoproteins, which are embedded in and cover this glucan layer. Chitin is a minor component of the cell wall (13, 18). The mannoproteins can be divided into two groups, the sodium dodecyl sulfate (SDS)-extractable mannoproteins and the glucan-extractable mannoproteins, which are solubilized by glucanase digestion of the glucan layer. Several glucanase-extractable mannoproteins have been identified. These proteins have two characteristics in common: their C-terminal regions are rich in serine and threonine, and they all contain putative glycosylphosphatidylinositol (GPI) attachment signals. Two of these proteins, α-agglutinin (22) and the core subunit of α-agglutinin (33), are involved in mating. The third, which is involved in flocculation, is the product of the *FLO1* gene (42). Because of the high serine and threonine content of their C-terminal regions, these proteins are probably heavily O glycosylated, which could give the protein a rod-like structure (15). The presence of a GPI anchor has been demonstrated only for the intracellular precursor form of α-agglutinin (47). The glucanase-extractable mannoproteins are proposed to be covalently linked to glucan (29, 38, 45). Several groups have investigated which part of the protein is responsible for anchoring the protein to the cell wall. Schreuder et al. (36) showed that a fusion protein consisting of the invertase signal sequence, the gua α-galactosidase coding sequence, and the C-terminal half of α-agglutinin was incorporated into the cell wall and was released only by glucanase extraction of isolated cell walls. In contrast, a control protein lacking the α-agglutinin part was released into the medium. Recently, Van Berkel et al. (44) showed that in a similar construct, addition of only the C-terminal 30 amino acids of α-agglutinin is sufficient for incorporation of the chimeric protein into the cell wall. Thus, the C-terminal stretch of 30 amino acids of α-agglutinin contains all information necessary to incorporate a protein into the cell wall. In a third experiment, Wojciechowicz et al. (47) showed that removal of the C-terminal hydrophobic sequence allowed secretion of active α-agglutinin in the medium. This finding indicates that a GPI attachment signal is essential for incorporation of a protein into the cell wall. However, the proteins encoded by *KRE1* (4) and *GPP1* (30) are both GPI anchored but are not known to be associated with the cell wall; thus, it seems that a GPI anchor alone is not sufficient to locate a protein in the cell wall. It is not clear in which way the proteins are covalently linked to the cell wall. Recently, a carbohydrate side chain containing β1,6-glucose residues has been shown to be attached to the glucanase-extractable proteins (24, 45), and it has been suggested that this chain is involved in cell wall anchorage (8, 44).

To gain further insight into the yeast cell wall structure, we aimed to isolate the major glucanase-extractable cell wall proteins of *S. cerevisiae*. Here we report the identification of three open reading frames (ORFs) encoding glucanase-extractable cell wall mannoproteins. They all have a serine- and/or threonine-rich C-terminal domain and contain putative GPI attachment signals. The effect of depletion of these cell wall protein genes was investigated.
TABLE 1. Oligonucleotides used for the construction of deletion fragments

<table>
<thead>
<tr>
<th>Gene</th>
<th>Oligonucleotide</th>
</tr>
</thead>
<tbody>
<tr>
<td>CWPI</td>
<td>1. GCGGAAATGTTAACCAGCAGCTCTATACCTACCC</td>
</tr>
<tr>
<td></td>
<td>2. CGGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>3. CGACATGGCAAGCCTGCGTTCGACATCTTCCCTAC</td>
</tr>
<tr>
<td></td>
<td>4. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
<tr>
<td></td>
<td>5. GCGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>6. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
<tr>
<td>CWP2</td>
<td>1. GCGGAAATGTTAACCAGCAGCTCTATACCTACCC</td>
</tr>
<tr>
<td></td>
<td>2. CGGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>3. CGACATGGCAAGCCTGCGTTCGACATCTTCCCTAC</td>
</tr>
<tr>
<td></td>
<td>4. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
<tr>
<td></td>
<td>5. GCGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>6. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
<tr>
<td>TIP1</td>
<td>1. GCGGAAATGTTAACCAGCAGCTCTATACCTACCC</td>
</tr>
<tr>
<td></td>
<td>2. CGGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>3. CGACATGGCAAGCCTGCGTTCGACATCTTCCCTAC</td>
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<td></td>
<td>4. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
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<td></td>
<td>5. GCGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
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<tr>
<td></td>
<td>6. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
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<tr>
<td>SRP1</td>
<td>1. GCGGAAATGTTAACCAGCAGCTCTATACCTACCC</td>
</tr>
<tr>
<td></td>
<td>2. CGGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
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<tr>
<td></td>
<td>3. CGACATGGCAAGCCTGCGTTCGACATCTTCCCTAC</td>
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<tr>
<td></td>
<td>4. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
<tr>
<td></td>
<td>5. GCGGATCCGTCGCGAGGGATCCATGGTCCAAAGGGCG</td>
</tr>
<tr>
<td></td>
<td>6. CGGAACTGGTAAACGATTGTAATCAGACGCAGAC</td>
</tr>
</tbody>
</table>

* Orientation is 5' to 3'. 1, 5' oligonucleotide of the left-hand fragment; 2, 3' oligonucleotide of the left-hand fragment; 3, 5' oligonucleotide of the right-hand fragment; 4, 3' oligonucleotide of the right-hand fragment.

MATERIALS AND METHODS

Strains and media. The Escherichia coli strain used in this study was JM109 (endA1 recA1 gyrA96 thi hsdR17 rK2 F′ traD36 proA6 lacIqZ M15) (46) and was grown in Luria broth (35) with 100 μg of ampicillin per ml when appropriate. The S. cerevisiae yeast strains used were TM2683-20 (MATα mel9 les2 his4) and SU30 (YEp6–2 LMA) (47). Yeast strains were grown in YPD (1% yeast extract, 2% Bacto Peptone, 2% glucose) or synthetic minimal medium (MM) consisting of 0.7% yeast nitrogen base, 2% glucose, and amino acids as necessary (37).

Reagents. Yeast nitrogen base, Bacto Peptone, Bacto Yeast Extract, and Bacto Agar were from Difco Laboratories (Detroit, Mich.). DNA restriction and modification enzymes were from New England Biolabs, Inc. (Beverly, Mass.) and Boehringer Mannheim Biochemicals (Indianapolis, Ind.). Phenylmethylsulfonyl fluoride was purchased from Boehringer Mannheim; laminarinase from mollusk (catalog no. L5144) and from Trehodroma species (catalog no. L5272) were obtained from Sigma Chemical Co. (St. Louis, Mo.). Polyvinylidene difluoride membranes (Immobilon P) were from Millipore Corp. (Bedford, Mass.). The goat anti-rabbit antibody was purchased from Bio-Rad Laboratories (Hercules, Calif.), and the ECL chemiluminescence kit was obtained from Amersham (Arlington Heights, Ill.). Zymolyase 20T and 100T were purchased from Seikagaku Kogyo Co. (Tokyo, Japan).

Isolation of cell wall mannoproteins. Cell walls from TM2683-20 were isolated and extracted with SDS and glucanase as described by Schreuder et al. (36). In addition, SDS-extracted cell walls were digested with both molusk and Trehodroma laminarinase. The glucanase-extracted cell wall mannoproteins were purified further by concanavalin A affinity chromatography as described by Van Rijnsum et al. (45). The eluted mannoproteins were precipitated with trichloroacetic acid (15%, final concentration), and the pellet was resuspended in 1/10 volume in SDS sample buffer (50 mM Tris-HCl [pH 6.8], 2% [wt/vol] SDS, 0.1% [wt/vol] bromphenol blue, 10% [wt/vol] glycerol).

Silver staining of SDS-polyacrylamide gels and Western blot (immunoblot) analysis. Glucanase-extracted mannoproteins were separated by polyacrylamide gel electrophoresis (PAGE) as described by Laemmli (21) and transferred to a 20% gradient gel. Silver staining of gels was performed as described by De Nobè et al. (9). For Western analysis, the proteins were subsequently transferred onto Immobilon polyvinylidene difluoride membranes as described by Towbin et al. (43). The blot was blocked in phosphate-buffered saline (PBS) containing 3% (wt/vol) bovine serum albumin for 1 h. The blot was subsequently washed three times in PBS and incubated with a 1:25,000 dilution of the anti-β,1,6-glucan antiserum for 14 h. The antiserum binding was visualized by use of the ECL fluorescent labelling kit and then chemiluminescence exposure of X-ray film.

Protein sequencing. Protein bands corresponding to those recognized by the anti-β,1,6-glucan antiserum were excised from a Western blot, and N-terminal sequences were determined by the Edman degradation method (1). The material obtained by extraction with trichodermin was further purified by removal of smaller molecules by filtration over a 100-kDa-cutoff filter (Amicon Corp., Danvers, Mass.). The 180-kDa protein that was predominantly present after filtration was N-terminally sequenced, as a soluble sample, by the Edman degradation method.

Transformation of strains and DNA isolation. E. coli JM109 was used for propagation of plasmids, and competent cells were prepared by the method of Nishimura et al. (26). Plasmid DNA was isolated by the alkaline lysis method described by Sambrook et al. (35). S. cerevisiae SU50 was transformed by electro-
cells) of Zymolyase 20T per ml. Subsequently, the decrease of the OD$_{600}$ was monitored for several hours.

**Electron microscopy.** Cells were grown in MM (OD$_{600}$ = 0.5), harvested, washed two times with distilled H$_2$O, fixed in 1.5% K$_2$CrO$_7$, and incubated at room temperature for 20 min. Fixed cells were embedded in low-melting-point agar and subsequently immersed in 30% aqueous N,N-dimethylformamide for 30 min and cryofixed in liquid propane. The frozen cells were freeze-substituted for 2 days at −90°C in anhydrous methanol (CS auto; Reichert-Jung). After the temperature was gradually increased (5°C/h) to room temperature, the methanol was replaced by anhydrous acetone and infiltrated with Spurr’s epoxy resin (40). Tissue was embedded in the resin at room temperature for 20 min. Fixed cell sections were sectioned with a diamond knife on an ultramicrotome (Reichert-Jung). Sections were viewed on a Philips EM420 electron microscope at an operating voltage of 80 kV. Fifty independent measurements of the thickness of the electron-dense outer layer of the cell wall were carried out on median sections.

**RESULTS**

**Isolation and N-terminal sequencing of glucanase-extractable mannoproteins.** It has been demonstrated that the glucanase-extractable mannoproteins often contain a carbohydrate side chain containing β1,6-glucose residues (24, 45). A polyclonal antiserum raised against β1,6-glucan (provided by Roy Montijn, University of Amsterdam) recognized four glucanmannoproteins released from the cell walls by the mollusk laminarinase extraction procedure. These four protein bands were excised from the Western blot and N-terminally sequenced.

The N-terminal sequence of the protein band with an estimated molecular mass of 55 kDa (Table 2) proved to be 100% homologous with the YKL096w gene product present in the yeast DNA sequencing data bank. This ORF, previously sequenced by Pallier et al. (28), is located on chromosome XI and represents a transcript capable of encoding 239 amino acids. It contains a putative GPI attachment signal at the C terminus, and the C-terminal part of the protein is very rich in serine (35%) and threonine (11%). A signal sequence is present at the N terminus, as the determined N-terminal sequence starts exactly after the predicted signal peptide cleavage site (46). We named this gene CWP1 (cell wall protein 1).

The N-terminal sequence of the protein band of approximately 80 kDa (Table 2) corresponded with the TIP1 gene product, identified by Kondo and Inouye (19) as a temperature-inducible protein by differential gene expression analysis. This protein consists of 210 amino acids and contains a putative GPI attachment signal, and the C-terminal part of the protein is rich in serine (37%). The presence of an N-terminal signal sequence was demonstrated as for Cwp1p.

No sequence data were obtained from the two upper bands of 145 and 205 kDa as estimated from SDS-gel electrophoresis. This was possibly caused by an N-terminal blockage of these proteins.

When the SDS-extracted cell walls were digested with laminarinase isolated from Trichoderma species, one predominant band of 180 kDa, which did not react with the anti β1,6-glucan antiserum, was detectable by silver staining (data not shown). This protein band was also N-terminally sequenced (Table 2).

No striking homology was found between this sequence and proteins in the *S. cerevisiae* gene bank. On the basis of this sequence, degenerate probes were designed for PCR. The sense primer 5’ CAA AAG CCT GCT GCT/CATT GCT/C CA 3’ was based on amino acids 4 to 8 and contained an additional HindIII restriction site. The antisense primer 5’ TC GAA TTC AC TGG A/GGC A/GGC 3’ was based on amino acids 28 to 31 and contained an additional EcoRI restriction site. The PCR gave, among others, a product of the predicted size which hybridized with an internal oligonucleotide (5’ GAT GGT CAA ATT/C INN GCT GT 3’) based on amino acids 11 to 17. The PCR product was isolated and used as a template for a second round of PCR. The products of this second amplification were cloned in pUC18, and the inserts of several plasmids were shown to hybridize with the internal oligonucleotide. One such clone had the capacity to code for an amino acid sequence identical to the N terminus of the protein encoded by a small ORF, YKL444 (CWP2), sequenced by Pallier et al. (28). In many cases, threonine residues deduced from the DNA sequence had been misidentified during the amino acid analysis, but closer inspection of the amino acid sequence data showed small threonine peaks in these places. Misidentification of these amino acids is possibly caused by glycosylation of the threonine residues. The ORF CWP2 is located on chromosome XI, codes for a protein consisting of 92 amino acids, has a putative GPI attachment signal, and is rich in serine (15%) and threonine (26%) in the C-terminal part. The determined N-terminal sequence starts at the predicted site for signal sequence cleavage; thus, a signal sequence is also present on this protein. This ORF was not included in the analysis of chromosome XI, as it encodes a protein of less than 100 amino acids.

**Structural analysis of the deduced protein sequences encoded by CWP1, TIP1, and CWP2.** Translation of the three identified ORFs into their amino acid sequences reveals that they have several characteristics in common. First, they all contain an N-terminal signal sequence for import into the endoplasmic reticulum (46). Second, their C-terminal halves are all rich in serine and/or threonine, which implies that these domains are highly O glycosylated. For the cell wall protein α-agglutinin, Wojciechowicz et al. (47) postulated that this structure facilitates the exposure of the N-terminal binding domain on the surface of the cell wall. Third, the deduced C-terminal amino acid sequences of these ORFs contain putative GPI attachment signals. Nuoffer et al. (27) analyzed the sequence requirements for peptide cleavage and glycolipid addition for Gas1p of *S. cerevisiae* and showed that asparagine is the most efficient anchor attachment site and that the two adjacent amino acids to the carbohydrate-terminal should have relative short side chains. The deduced protein sequences of Cwp1p, Tip1p, and Cwp2p all contain an asparagine followed by glycine and alanine near the carboxyl termini (Fig. 1); the necessary polar and hydrophobic regions (6, 7) are also present, and although no specific sequence is required (5), we

<table>
<thead>
<tr>
<th>Protein mol wt (10^3)</th>
<th>N-terminal amino acid sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>55...................</td>
<td>D/G-S-E-E-F-G-L-V-S-P^</td>
</tr>
</tbody>
</table>

^ Determined by SDS-PAGE.

| Contaminating peaks derived from the concanavalin A column material were subtracted from the determined sequence.
underlined. in boldface, and an identical sequence found in Srp1p, Cwp1p, and Cwp2p is dues, which are proposed to act as the attachment sites of the GPI anchors, are and homologous carboxyl-terminal sequences are boxed. The asparagine residues, which such as to act as the attachment sites of the GPI anchors, are in boldface, and an identical sequence found in Srp1p, Cwp1p, and Cwp2p is underlined.

find a high degree of homology between the hydrophobic regions (Fig. 1).

Disruption of the CWP1, TIP1, CWP2, and SRP1 genes. Disruption mutations in the newly identified ORFs were constructed as described in Materials and Methods. To confirm that each identified ORF encodes the mannoprotein that was N-terminally sequenced, glucanase-extractable mannoproteins were isolated from cell walls of each deletion mutant. Comparison of the protein patterns of the parent strain and the deletion mutants confirmed that the identified ORFs code for the expected glucanase-extractable mannoproteins. In a Western analysis with the anti-β1,6-glucan antiserum of the Tip1p copy analysis: no difference between the electron-dense outer layer of the parent strain and any of the deletion mutants. The lack of an increased sensitivity of the Cwp2 mutant in this phase was in agreement with electron microscopy analysis. Electron microscopy of the cwp2 mutant showed a sharp reduction of the electron-dense outer layer of the cell wall (Fig. 4). The thickness of the electron-dense outer layer of the cell wall of the parental strain was 119 ± 19 nm, whereas in the cwp2 mutant, the thickness was 36 ± 7 nm. The other deletion mutants displayed no increased Zyomyces sensitivity compared with the wild type, and in stationary phase, no effect was found for any of the deletion mutants. The lack of an increased sensitivity of the cwp2 mutant in this phase was in agreement with electron microscopy analysis: no difference between the electron-dense outer

TABLE 3. Phenotypic characteristics of exponentially growing cells of the deletion mutants

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Sensitivity to:</th>
<th>Calcofluor white</th>
<th>Congo red</th>
<th>Zyomyces</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Wild type</td>
<td>+</td>
<td>+</td>
<td>wt</td>
</tr>
<tr>
<td>cwp1Δ</td>
<td>cwp2Δ</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>tip1Δ</td>
<td>cwp1Δ cwp2Δ</td>
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<td>++</td>
<td>++</td>
</tr>
<tr>
<td>srp1Δ</td>
<td>+</td>
<td>++</td>
<td>++</td>
<td>++</td>
</tr>
</tbody>
</table>

a +, inhibitory concentration = 2.0 mg/ml; ++, inhibitory concentration = 1.8 mg/ml; ++++, inhibitory concentration = 1.4 mg/ml.

Zyomyces concentration was 30 μg/ml; incubation lasted for 2.5 h. wt, wild-type sensitivity; sensitive, increased sensitivity relative to the wild type.

FIG. 2. Comparison of the cell wall proteins of the parent strain and the deletion mutants. (A) Western blot of glucanoproteins separated on an SDS–2% polyacrylamide gel and incubated with the anti-β1,6-glucan antiserum. The mannoproteins visualized in the right-hand panel were isolated and silver-stained to reveal the proteins. whereas the wild-type strain was able to grow at a Congo red concentration of 2 mg/ml (Table 3). The cwp1Δ, tip1Δ, and srp1Δ mutants also displayed an increased sensitivity to these compounds, but for these mutants, the effect was less pronounced: their growth was inhibited on plates containing more than approximately 1.8 mg of Congo red per ml.

The deletion mutants were also tested for an increased sensitivity to Zyomyces. The sensitivity of the cwp2Δ mutant, in the exponential phase of growth, increased (Fig. 3), and this was also reflected in an increase in sensitivity found for the cwp1Δ cwp2Δ double mutant (data not shown). The depletion of Cwp2p allows a faster start to the digestion of the glucan layer by Zyomyces. This finding implies a thinner mannoprotein layer which is likely to be caused by the lack of Cwp2p in the outer part of the cell wall. Electron microscopy of the cwp2 mutant showed a sharp reduction of the electron-dense outer layer of the cell wall (Fig. 4). The thickness of the electron-dense outer layer of the cell wall of the parental strain was 119 ± 19 nm, whereas in the cwp2 Δ mutant, the thickness was 36 ± 7 nm. The other deletion mutants displayed no increased Zyomyces sensitivity compared with the wild type, and in stationary phase, no effect was found for any of the deletion mutants. The lack of an increased sensitivity of the cwp2Δ mutant in this phase was in agreement with electron microscopy analysis: no difference between the electron-dense outer

FIG. 1. Comparison of the deduced amino acid sequences of CWP1, CWP2, TIP1, and SRP1. Putative amino-terminal signal sequences in Cwp1p and Cwp2p and homologous carboxyl-terminal sequences are boxed. The asparagine residues, which are proposed to act as the attachment sites of the GPI anchors, are in boldface, and an identical sequence found in Srp1p, Cwp1p, and Cwp2p is underlined.
layer of cells of the cwp2Δ mutant strain and cells of the parental strain was observed (data not shown).

The deletion mutants did not show increased sensitivity to low pH (pH 2 to 4) or high salt concentrations (NaCl, 5 to 20%) (data not shown).

DISCUSSION

Structural analysis of CWP1, TIP1, CWP2, and SRP1 and their deduced protein sequences. In this report, three ORFs encoding related glucanase-extractable mannoproteins have been identified. Translation of these ORFs into their deduced amino acid sequences revealed that they share several characteristics which are also present in the deduced amino acid sequence of SRP1, suggesting that Srp1p is a potential cell wall protein. For this reason, Srp1p was included in our analysis. The similarities between Tip1p and Srp1p have already been described by Kondo and Inouye (19). In the N-terminal signal sequences of Cwp1p and Cwp2p, there is a very homologous stretch of 15 amino acids. Furthermore, the element ISQIT DGQIQA is found in Cwp1p (amino acids 197 to 207), Cwp2p (amino acids 26 to 36), and Srp1p (amino acids 213 to 223). This element displays a high degree of similarity with the 11-amino-acid tandem repeats found in the secretory glycoprotein encoded by the HSP150 gene (34). The C-terminal 25 amino acids of Cwp1p, Cwp2p, Srp1p, and the C-terminal 12 amino acids of Tip1p containing (part of) the GPI attachment signal are also very homologous. The four most C-terminal amino acids, predicted to remain on Cwp1p, Cwp2p, and Srp1p (QTEN) after cleavage of the GPI attachment signal, might act as an element directing proteins to their correct place in the cell wall. Munoz-Dorado et al. (25) identified two elements (ATTGG) in the promoter region of TIP1 which are located in the region responsible for induction of the gene after a cold shock. In E. coli, this DNA sequence is recognized by a cold shock protein (CspA [17]). The promoter region of the CWP1 gene also contains two of these elements (positions −334 and −669 relative to ATG).

FIG. 3. Zymolyase sensitivities of the deletion mutant SU50 cwp2Δ and SU50. Exponentially growing cells were incubated in 50 μg of Zymolyase 20T, and the decrease of the OD660 nm was monitored for 2.5 h. □, SU50; ◇, SU50 cwp2Δ.

FIG. 4. Electron microscopy of SU50 (A) and SU50 cwp2Δ (B) cells harvested from the exponential growth phase. Electron microscopy was carried out by W. Müller (Utrecht University) as described in Materials and Methods. Bars represent 150 nm.
Comparison of the molecular weight of Cwp2p determined by SDS-PAGE and its predicted molecular weight, calculated from the deduced amino acid sequence of the mature protein, reveals a substantial difference. This discrepancy cannot be accounted for by glycosylation, as Cwp2p contains no consensus sequence for N-glycosylation and even O-glycosylation of all serine and threonine residues would lead to a lower molecular weight than observed by SDS-PAGE. Furthermore, the Cwp2p mannoprotein was not recognized by the β,1,6-glucan antiserum, indicating that a β,1,6-glucose-containing side chain is not present on this protein. This implies that the β,1,6-glucose-containing side chain is not essential for cell wall anchorage of Cwp2p. The anomalous behavior of Cwp2p in SDS-PAGE may, for example, be caused by low binding of SDS to this protein, which would cause the slower migration on an SDS-polyacrylamide gel.

Functional analysis of deletion mutants. The cwp1Δ, tip1Δ, cwp2Δ, and srp1Δ deletion mutants grew as well as the parent strain in both minimal and rich media. Under these conditions, the products of the deleted genes are therefore not essential for growth. However, when the deletion mutants were grown in the presence of calcofluor white or Congo red, both of which cause a change in the cell wall structure and aggravate the effects of cell wall defects (11, 20, 31), increased sensitivities toward both compounds were found. The fact that the cwp2Δ mutant is more sensitive to these compounds than the other three mutants could be explained by a greater abundance of Cwp2p in the cell wall, causing a greater perturbation to the cell wall upon deletion.

Zlotnik et al. (49) showed that lysis of the cell wall of S. cerevisiae with Z-glucanase requires a previous incubation with Z-protease. This finding implies that the mannoproteins are involved in protecting the structural glucan from the action of the glucanase. Incubation of the deletion mutants with Zymolase, which contains Z-glucanase and Z-protease, showed an increased sensitivity of the cwp2Δ mutant and the cwp1Δ cwp2Δ double mutant in exponentially growing cells. Electron microscopic analysis showed that the depletion of Cwp2p corresponds with a considerable decrease in thickness of the electron-dense layer around the glucan layer in this mutant. In the other deletion mutants, no increased Zymolase sensitivity was found. On the basis of their sequence similarity, it is possible that these proteins can substitute for each other’s function. Another possibility is that these cell wall proteins are less important for the stability of the cell wall. Stationary-phase cells of the mutants showed no enhanced Zymolase sensitivity. This could imply that other cell wall proteins become more abundant in the stationary phase.

Through analysis of cell wall glucanmannoproteins, we have been able to assign gene products to two ORFs identified in the yeast genomic sequencing program. In addition, we have identified the TIP1 gene product as a cell wall component. These genes encode cell wall proteins in S. cerevisiae.

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