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Published in:
The journal of clinical investigation

DOI:
10.1172/JCI8267

Link to publication

Citation for published version (APA):

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Multidrug resistance protein 1 protects the choroid plexus epithelium and contributes to the blood-cerebrospinal fluid barrier

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Received for publication August 26, 1999, and accepted in revised form December 21, 1999.

Multidrug resistance protein 1 (MRP1) is a transporter protein that helps to protect normal cells and tumor cells against the influx of certain xenobiotics. We previously showed that Mrp1 protects against cytotoxic drugs at the testis-blood barrier, the oral epithelium, and the kidney urinary collecting duct tubules. Here, we generated Mrp1/Mdr1a/Mdr1b triple-knockout (TKO) mice, and used them together with Mdr1a/Mdr1b double-knockout (DKO) mice to study the contribution of Mrp1 to the tissue distribution and pharmacokinetics of etoposide. We observed increased toxicity in the TKO mice, which accumulated etoposide in brown adipose tissue, colon, salivary gland, heart, and the female urogenital system. Immunohistochemical staining revealed the presence of Mrp1 in the oviduct, uterus, salivary gland, and choroid plexus (CP) epithelium. To explore the transport function of Mrp1 in the CP epithelium, we used TKO and DKO mice cannulated for cerebrospinal fluid (CSF). We show here that the lack of Mrp1 protein causes etoposide levels to increase about 10-fold in the CSF after intravenous administration of the drug. Our results indicate that Mrp1 helps to limit tissue distribution of certain drugs and contributes to the blood-CSF drug-permeability barrier.


Introduction
Cancer cells overproducing cellular drug extruders may become resistant to a wide spectrum of drugs with different structures or cellular targets. This phenomenon is called multidrug resistance (MDR) (1). Three ATP-dependent transmembrane drug transporters — the multidrug resistance gene product MDR1 P-glycoprotein (Pgp) (1), the breast cancer resistance protein (2–4), and multidrug resistance protein 1 (MRP1) (5–9) — can render cells multidrug resistant in vitro. MRP1 is a so-called glutathione-conjugate pump, or multispecific organic anion transporter (10), and is a versatile pump. It can mediate the extrusion of glutathione-S-conjugates, glucuronide conjugates, and sulfate conjugates of drugs (10–13); it is the major transporter for endogenous LTC4, an important mediator of the inflammatory response (11, 14); and it can even extrude neutral and basic organic compounds if the cell contains normal levels of glutathione (15–18).

To elucidate the physiological functions of the MDR-conferring transporters, mutant mice have been generated lacking Mdr1a Pgp and Mdr1b Pgp (19, 20), or lacking Mrp1 (14, 21). Mdr1a Pgp plays an important role in the intestinal epithelium, where it actively excretes xenobiotics absorbed from the intestinal lumen back into the lumen. It thereby limits the entry of Pgp substrates into the body. In addition, Mdr1a Pgp plays an important role in the blood-brain barrier (19, 20, 22–24). This barrier is primarily formed by the endothelium of the blood capillaries in the brain. The presence of Mdr1a Pgp in the apical (luminal) plasma membranes of these capillary endothelial cells (25) is crucial for the protection of the brain against several xenobiotics entering from the blood (19).

Mrp1-deficient mice lack the main high-affinity glutathione conjugate pump activity in erythrocytes, and cells derived from these mice show increased sensitivity to anti-cancer drugs (14). Mrp1 has important protective roles in the epithelium of the tongue and cheek, the urinary collecting duct epithelium, and the epithelial Sertoli cells in the testicular tubules; we have shown that Mrp1 prevents drug-induced oral mucositis, dia-
Monoclonal antibodies against MRP1 (27, 28) detect this transport protein in several tissues. Its presence has been demonstrated in the basal plasma membrane of the Sertoli cells (26), in the basolateral membranes of the lung epithelium (26, 29), in the liver (30), in the rat choroid plexus (CP) epithelium (31), and in transfected kidney cell lines overproducing human MRP1 (32). Here, we show that Mrp1 is also present in the mouse CP epithelium.

The brain is protected against drugs by 2 drug-permeability barriers: the blood-brain barrier and the blood-cerebrospinal fluid (CSF) barrier. The blood-brain barrier is formed by the endothelium of the blood capillaries; diffusion of some hydrophobic drugs through this barrier is counteracted by transporters such as Mdr1a Pgp (19, 33). The blood-CSF barrier (34) is formed by the epithelium of the CP (35). Recently, some drug transporters (e.g., organic anion transporter protein 1, MDR1 Pgp, and MRP1) have been postulated to play a drug-transporting role in the CP (31, 36, 37). The polarized choroidal epithelium uses active transport mechanisms to extract micronutrients from the blood, and produces and secretes CSF (38). The CP also has an important function in the clearance of metabolic waste products and substrates such as LTC4 (25–27 g) were anesthetized by intraperitoneal injection of 6 μL Hypnorm/Dormicum/water (1:1:2, vol/vol/vol) per gram body weight (Hypnorm, Janssen Pharmaceutica, Beerse, Belgium; Dormicum; Roche Pharmaceuticals, Basel, Switzerland). A tail cannula consisting of 15 cm of polyethylene tubing (outer diameter 0.61 cm, inner diameter 0.28 cm) and a 7-cm piece of Zeus tubing (outer diameter 0.48 mm, inner diameter 0.24 mm; Zeus Industrial Products, Orangeburg, South Carolina, USA) was inserted into the tail artery to reach the aorta. The

**Methods**

**Animals.** The animals used were Mdr1a- and Mdr1b-deficient (Mdr1a<sup>−/−</sup>/Mdr1b<sup>−/−</sup>) DKO mice and Mrp1-deficient (Mrp1<sup>−/−</sup>) mice, generated by gene targeting in embryonic stem cells as described by Schinkel et al. (19) and Wijnholds et al. (14). Using these mice, Mdr1a<sup>−/−</sup>/Mdr1b<sup>−/−</sup>/Mrp1<sup>−/−</sup> triple-knockout (TKO) mice were obtained by crossing. All mice used were on a 50% 129/Ola, 50% FVB genetic background. All animals were housed in constant-temperature rooms with a 12-hour-light/12-hour-dark cycle, and had access to a pelleted chow diet (Hope Farms BV, Woerden, The Netherlands) and acidified water ad libitum. Mouse handling and experimental procedures were conducted in accordance with the Netherlands Cancer Institute guidelines for animal care and use.

**Drug sensitivity and [3H]etoposide distribution in female mice.** Etoposide phosphate (Etopophos, 100 mg effective etoposide; Bristol-Myers Squibb Pharmaceutical, Princeton, New Jersey, USA) was dissolved in sterile 0.9% NaCl to obtain a stock solution of 4–20 mg/mL. Concentrations were adjusted to inject 5–6.5 μL/g body weight (20–130 mg/kg) intravenously in the tail vein of female or male mice (11–16 weeks old) that were lightly anesthetized with diethyl ether. Mice were weighed and were observed daily for a period of 2 weeks. Lethal toxicity occurred between days 2 and 13 after injection.

For tissue distribution experiments, tracer [3H]etoposide (475 Ci/mol) was diluted with carrier etoposide (Vepesid; Bristol-Myers Squibb Pharmaceutical) in 5% (wt/vol) glucose, and was injected into the tail vein at a dose of 1 mg/kg. Tissues from female mice were collected at 4 hours and were processed as described (19).

**Histological and immunohistochemical analyses.** Histological and immunohistochemical analyses were performed as described (26, 27).

**Cannulation of lateral brain ventricles for CSF sampling.** Female DKO mice and TKO mice 11–16 weeks of age (25–27 g) were anesthetized by intraperitoneal injection of 6 μL Hypnorm/Dormicum/water (1:1:2, vol/vol/vol) per gram body weight (Hypnorm, Janssen Pharmaceutica, Beerse, Belgium; Dormicum; Roche Pharmaceuticals, Basel, Switzerland). A tail cannula consisting of 15 cm of polyethylene tubing (outer diameter 0.61 cm, inner diameter 0.28 cm) and a 7-cm piece of Zeus tubing (outer diameter 0.48 mm, inner diameter 0.24 mm; Zeus Industrial Products, Orangeburg, South Carolina, USA) was inserted into the tail artery to reach the aorta.

**Figure 1**

Loss of body weight in mice lacking Mrp1 on a genetic background lacking Mdr1a Pgp and Mdr1b Pgp (TKO mice). DKO mice, TKO mice, and Mrp1<sup>−/−</sup> mice received a single intravenous dose of 60 mg/kg Etopophos<sup>®</sup>, and the loss of body weight (%) was measured over 7 days. *P < 0.001.
Table 1
Etophos® toxicity in male wild-type, Mrp1–/–, DKO, and TKO mice

<table>
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<th>Etopophos® dose (mg/kg)</th>
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The number of surviving animals per group at each intravenous Etopophos® dose is shown. The data are from 2 independent experiments (A and B), using mice 11–16 weeks old. For additional toxicity experiments using wild-type and Mrp1–/– mice, see ref. 26. TKO mice show increased sensitivity to Etopophos® (P < 0.01).

Altered drug sensitivity in mice lacking Mrp1 on an Mdr1a/Mdr1b–null genetic background. We have previously shown that Mrp1 deficiency renders mice hypersensitive to etoposide or its water-soluble derivative, etoposide phosphate (Etopophos®) (14, 26). Because etoposide is also transported by Mdr1-type Pgps, we crossed Mrp1–/– mice with DKO Mdr1a–/–/Mdr1b–/– mice to obtain TKO Mrp1–/–/Mdr1a–/–/Mdr1b–/– mice. No differences in health, life span, or fertility were observed among TKO, DKO, Mrp1–/–, or wild-type mice in our animal facility. The TKO mice are more sensitive to etoposide than are DKO and Mrp1–/– mice; all 3 of those groups are more sensitive than are wild-type mice (Tables 1 and 2). Seven days after a single intravenous 60 mg/kg dose of Etopophos®, female Mrp1–/– mice (n = 5) retained 92.3 ± 3.8% of their body weight (24.7 ± 1.1 g at day 0) and DKO mice (n = 5) retained 99.5 ± 1.2% of their body weight (27.8 ± 0.9 g at day 0), whereas female...
TKO mice (n = 5) retained 74.1 ± 2.6% (P < 0.01) of their body weight (26.5 ± 0.7 g at day 0) (Figure 1).

Except for their weight loss, the drug-treated TKO mice showed no clear gross abnormalities. Microscopic analysis at day 7 of TKO, DKO, and Mrp1–/– tissues from mice treated with 60 mg/kg Etopophos showed a disruption of the epithelial layer of the tongue and cheek only in the TKO mouse tissue (all TKO mice were thus affected). Inflammation of the tongue epithelium was detected in 4 out of 5 TKO animals (results not shown). The defects were similar to those observed in Mrp1-deficient mice after higher drug doses (26). Analysis of the colon of mice treated with 50–60 mg/kg Etopophos showed crypt degeneration (9 of 12 TKO mice; 3 of 11 DKO mice) and colitis (5 of 12 TKO mice; 1 of 11 DKO mice) (data not shown). The increase in gut damage to TKO mice suggests that Mrp1 contributes to the protection of the colon. Except for sporadic signs of inflammation in the stomach (in 3 of 5 mice), and a degeneration of the oocytes in the ovary (in 2 of 5 mice), an extensive histopathological examination revealed no further abnormalities in other tissues (data not shown) of the TKO mice.

We previously found that Mrp1-deficient mice, when treated with a dose of 100 mg Etopophos per kg body weight, developed diabetes insipidus, demonstrated by the overproduction of dilute urine (26). At close to the maximal drug dose tolerated by the TKO mice in metabolic cages (50 mg/kg), water intake, urine production (DKO, 1.7 ± 0.5 mL per 48 hours; TKO, 1.3 ± 0.3 mL per 48 hours), food consumption, and feces production did not differ between DKO and TKO mice. No differences were detected 7 days after drug administration in plasma sodium, urea, creatinine, or protein levels. The number of leukocytes was reduced in TKO mice (DKO, [13.6 ± 1.5] × 10⁹/L; TKO, [3.0 ± 1.2] × 10⁹/L; P < 0.01), whereas hemoglobin, hematocrit, and the number of thrombocytes did not differ between DKO mice and TKO mice (data not shown).

Altered drug tissue distribution in mice lacking Mrp1. Although Mrp1 is a ubiquitous protein present in several epithelia (26, 27, 29, 31), our previous etoposide tissue distribution experiments did not show a significant increase in the accumulation of etoposide in Mrp1-deficient tissues (14). This negative result could be explained by redundancy of etoposide transporters; obvious candidates for the redundant transporters are the Mdr1-type Pgps. To test whether Mrp1 can contribute to etoposide cellular clearing in vivo, we administered 1 mg/kg of radiolabeled etoposide intravenously to DKO mice and TKO mice, and determined the tissue distribution of total radioactivity 4 hours after injection. Table 3 shows that the etoposide plasma levels of DKO and TKO mice were comparable, indicating that Mrp1 does not significantly contribute to the renal, liver, or intestinal clearance of etoposide as was observed for Mdr1-type Pgps (19, 20). In the TKO mice, however, we observed increased etoposide accumulation in brown adipose (2.7-fold), colon (2.3-fold), urogenital tract (1.5-fold), salivary gland (1.4-fold), and heart (1.3-fold) tissues (P < 0.05); no significant accumulation was detected in brain, liver, kidney, or
Figure 3
Increased accumulation of etoposide into the CSF of mice lacking Mrp1 (TKO mice) compared with mice containing Mrp1 (DKO mice). Brain vessels were cannulated and then perfused with artificial CSF. Etopophos® (60 mg/kg) was administered via a tail vein; CSF was collected for 1 hour after drug administration, and the etoposide concentration was determined. (a) The TKO mice (n = 5) accumulated 10-fold more etoposide in the CSF than did the DKO mice (n = 4) (8.1 ± 0.9 μM and 0.8 ± 0.3 μM, respectively; P < 0.001), whereas levels in total brain (3.9 ± 0.7 μmol/kg [n = 5] and 2.9 ± 0.7 μmol/kg [n = 4], TKO and DKO mice, respectively) did not differ significantly 1 hour after drug administration. (b) Etoposide levels in plasma (98 ± 12 μM [n = 5] and 114 ± 24 μM [n = 4], TKO and DKO mice, respectively) did not differ significantly 1 hour after drug administration.

Discussion
Our results indicate that Mrp1 helps to prevent the accumulation of etoposide in vivo, especially in epithelial tissues lining lumina. For some tissues, this protection by Mrp1 is considerable — for example in the CP epithelium (31). If Mrp1 contributes to the drug-permeability barrier between blood and CSF, we would anticipate that Mrp1 transports drugs specifically from the CP toward the blood. To test this hypothesis, we compared the accumulation of etoposide in the CSF of DKO mice and TKO mice. This allowed us to avoid the complicating contribution of Mdr1a Pgp, which is present in the brain capillaries (20) and is reported to be present in the apical membrane of the CP epithelium (31). We chose etoposide as a model substrate because it is clinically relevant and is the best (anti-cancer) drug substrate reported for Mrp1. We preferred to use etoposide as the substrate because the best substrates reported — e.g., the physiological substrate LTC4 and the test substance S-(2,4-dinitrophenyl)-glutathione — are hydrophilic and are unlikely to enter the CP epithelium. Cannulas were inserted into the left and right lateral brain ventricles adjacent to the CP, and artificial CSF was slowly perfused at a rate of 0.1 μL/min through the brain lateral ventricles. Etopophos® (60 mg/kg) was administered into a tail artery, and CSF was collected for a period of 1 hour. We found that the TKO mice accumulated 10-fold more etoposide in the CSF than did the DKO mice (P < 0.001), whereas levels in total brain and plasma did not differ 1 hour after drug administration (Figure 3, a and b). These results show the potential importance of Mrp1 at the blood-CSF barrier in determining drug levels in the CSF.
salivary gland, heart, and urogenital tract, as demonstrated by the lack of an observed altered drug distribution in the presence of Mdr1-type Pgps (14).

Most importantly, our experiments provide new information on the physiological function of Mrp1 in the CP. Rao et al. (31) recently showed that enriched membrane fractions of human, rat, and mouse CP contain Mrp1, and that the Mrp1 band on Western blots is missing in CP from Mrp1−/− mice. Staining with the anti-Mrp1 antibody was confined to epithelial cells, and mainly the basolateral part of the cells was stained. In cultured rat CP epithelial cells, most of the Mrp1 was present in a vesicular compartment (31). Inhibition of Mrp1 with the LTDy-receptor antagonist and the Mrp1 inhibitor MK-571 increased accumulation of Tc-sestamibi (an Mrp1 substrate) by 2-fold in these cells, but did not inhibit transport of Tc-sestamibi through the cell monolayer. Our experiments on the accumulation of etoposide in CSF of DKO mice and TKO mice show that mouse Mrp1 in the CP effectively prevents the entry of etoposide from the blood into the CSF.

The increased accumulation of etoposide in colon tissue of drug-treated TKO mice is interesting, because high-dose drug treatment may disrupt the normal function of the colonic epithelium (51). This epithelium has an important function in the reabsorption of water from the stool, and drug-induced damage may contribute to dehydration and body weight loss. It is possible that the observed increased frequency of colitis in drug-treated Mrp1-deficient mice is related to the genetic background of our DKO and TKO mice. It has been reported that some Mdr1a-deficient mice (on an FVB genetic background) are susceptible to development of a severe spontaneous intestinal inflammation with pathology similar to that of human inflammatory bowel disease (52), whereas this has not yet been observed in any of the deficient mice generated and bred in this institute (19, 20).

The increased accumulation of etoposide in brown adipose tissue of drug-treated TKO mice indicates a protective function for Mrp1. However, we could not detect Mrp1 protein in brown adipose tissue. The increased accumulation of etoposide after its intravenous administration may be caused by the drug-clearing activity of Mrp1 present at low levels. Tissue-level concentrations of drugs taken up by passive diffusion are determined by the partition ratio and the perfusion rate, however, and these may differ between TKO and DKO mice. Damage or disturbance of the normal function of brown adipose cells may disturb body temperature regulation and contribute to weight loss. The increased accumulation of etoposide in the female urogenital system may lead to a degeneration of oocytes, as was observed for some of the drug-treated TKO animals. The increased accumulation of etoposide in the salivary glands may result in abnormal function of this epithelium, such as an overproduction of mucus as has been reported (14).

In conclusion, these results indicate that at least 2 drug-transporter proteins (MDR1 Pgp and MRP1 in humans) may affect the tissue distribution and pharmacokinetics of an anti-cancer drug such as etoposide.

Acknowledgments

We thank our colleagues Zsolt Holló and Tohru Saeki for critical advice on the manuscript, and A.J. Schrauwers for biotechnical assistance. This work was supported in part by grants from the Dutch Cancer Society.

23. de Lange, E.C., de Bock, G., Schinkel, A.H., de Boer, A.G., and Breimer,
3. Allikmets, R., Schriml, L.M., Hutchinson, A., Romano-Spica, V., and
27. Kim, C.S., and Prichard, J.B. 1993. Transport of 2,4,5-trichloropheno-