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Background: Crohn’s disease is characterised by a chronic relapsing inflammation of the bowel in which proinflammatory cytokines play an important perpetuating role. Mitogen activated protein kinase p38 (p38 MAPK) has been established as a major regulator of the inflammatory response, especially with regard to production of proinflammatory cytokines, but its role in inflammatory bowel disease is unexplored. In this paper we describe the effects of a specific p38 MAPK inhibitor, SB 203580, in trinitrobenzene sulphonic acid (TNBS) induced colitis in mice.

Results: SB 203580 had a dichotomalous effect in TNBS mice. Weight loss of TNBS mice treated with SB 203580 was significantly worse and colon weight on sacrifice was significantly increased in MAPK inhibitor treated TNBS mice (229.2 mg and 289.1 mg, respectively). However, the total number of cells in the caudal lymph node decreased to 188.8×10^6 cells in SB 203580 treated TNBS mice compared with 334×10^6 cells in vehicle treated mice. CD3/CD28 double stimulated caudal lymph node cells of SB 203580 treated mice showed decreased interferon γ production but increased tumour necrosis factor α production. The concentration of interleukin 12p70 in colon homogenates was significantly decreased in SB 203580 treated mice whereas concentrations of interleukin 12p40, tumour necrosis factor α, and interleukin 10 were in similar vehicle and SB 203580 treated TNBS mice.

Conclusion: Our results reveal a dichotomy in p38 MAPK action during experimental colitis.

MATERIALS AND METHODS
Mice and induction of colitis
All experiments were approved by the Animal Studies Ethics Committee of the University of Amsterdam, the Netherlands. BALB/c mice were obtained from Harlan Sprague Dawley Inc (Horst, the Netherlands). Mice were housed under standard conditions, and supplied with drinking water and food (AM-II 10mm; Hope Farms, Woerden, the Netherlands). Experiments

Abbreviations: MAPK, mitogen activated protein kinase; MAPKAP, MAP kinase activated protein kinase; IL, interleukin; TNF-α, tumour necrosis factor α; IFN-γ, interferon γ; TNBS, trinitrobenzene sulphonic acid; SDS, sodium dodecyl sulphate; TBS, Tris buffered saline; CIN, caudal lymph nodes; ATF-2, activation of transcription factor 2.
were conducted in eight and 10 week old female BALB/c mice. Colitis was induced by rectal administration of two doses (separated by a seven day interval) of 2 mg of 2,4,6-trinitrobenzene sulfonic acid (TNBS; Sigma Chemical Co., St Louis, Missouri, USA) in 40% ethanol (Merck, Darmstadt, Germany) using a vinyl catheter positioned 3 cm from the anus (10 mice per group). During administration, mice were anaesthetised using isoflurane (1-chloro-2,2,2-trifluoroethylisoflurane-difluoromethyl-ether; Abbott Laboratories Ltd, Queenborough, Kent, UK), and after administration were kept vertical for 60 seconds. Control mice underwent identical procedures but were given physiological salt. All mice were sacrificed nine days following the first TNBS administration (that is, 48 hours following the second TNBS challenge). Mice were treated daily by intraperitoneal injection of 1 ml SB 203580/kg body weight in 1 ml of sterile saline or 0.01% DMSO in 1 ml of sterile saline as a vehicle control.

To study the kinetics of p38 MAPK activity in TNBS colitis, mice administered TNBS were sacrificed, 0, 1, 3, 5, 8, and 9 days after induction of colitis. Mice were treated with either vehicle or SB 203580.

Assessment of inflammation
Body weights were recorded daily. Spleens, caudal lymph nodes (CLN), and colons were harvested on sacrifice. The colons were removed through a midline incision and opened longitudinally. After removal of faecal material the wet weight of the distal 6 cm was recorded and used as an index of disease related intestinal wall thickening. Subsequently, the colons were longitudinally divided into two parts, one of which was used for histological assessment.

Histological analysis
The longitudinally divided colons were rolled up, fixed in 4% formalin, and embedded in paraffin for routine histology. Two investigators who were blinded to the treatment allocation of the mice scored the following parameters: (1) percentage of area involved, (2) number of follicle aggregates, (3) oedema, (4) erosion/ulceration, (5) crypt loss, and (6) infiltration of mononuclear and polymorphonecuclear cells. The percentage of area involved and crypt loss was scored on a scale ranging from 0 to 4 as follows: 0, normal; 1, <10%; 2, 10%; 3, 10–50%; and 4, >50%. Erosions were defined as 0 if the epithelium was intact, 1 for involvement of the lamina propria, 2 for ulcerations involving the submucosa, and 3 when ulcerations were transmural. The severity of the other parameters was scored on a scale from 0 to 3 as follows: 0, absent; 1, weak; 2, moderate; and 3, severe. Hence the score ranged from 0 to a maximum of 26 points.

Cell culture and ELISA for cytokines
CLN cell suspensions were prepared using filter cell strainers (Becton/Dickinson Labware, New Jersey, USA). Cells were suspended in RPMI 1640 medium (BioWhittaker-Boehringer, Verviers, Belgium) containing 10% fetal calf serum and the antibiotics ciprofloxin (10 µg/ml) (Sigma Chemical Co., St Louis, Missouri, USA). Cell suspensions were counted and 1×10⁵ cells were incubated in 200 µl RPMI containing antibiotics and 10% fetal calf serum in triplicate wells. Cells were stimulated by pre-coating with anti-CD3 antibody (1:30 concentration; 145.2C11 clone) and soluble anti-CD28 antibody (1:1000 concentration; Pharmingen, San Diego, California, USA). Supernatants were removed after 48 hours and IFN-γ and TNF-α (R&D systems, Abingdon, UK) concentrations measured by ELISA.

Colon homogenates
The colon was harvested and homogenates were made with a tissue homogeniser in nine volumes of Greenburger lysis buffer (300 mM NaCl, 15 mM Tris HCl, 2 mM MgCl₂, 2 mM Triton (X-100), pepstatin A, leupeptin, aprotinin (all 20 ng/ml), pH 7.4). Tissue was lysed for 30 minutes on ice followed by two centrifugations (10 minutes, 14 000 g, 4°C). Homogenates were stored at −20°C until use. TNF-α and IL-10 (both R&D), and IL-12p70 and IL-12p40 (both Pharmingen) concentrations were measured by ELISA.

p38 MAPK activity assay and western blotting
Colon homogenates were made using a tissue homogeniser in nine volumes of ice cold cell lysis buffer (20 mM Tris (pH 7.5), 150 mM NaCl, 1 mM EDTA, 1 mM EGTA, 1% Triton, 2.5 mM sodium pyrophosphate, 1 mM β-glycerophosphate, 1 mM Na₃VO₄, 1 µg/ml leupeptin, and 1 mM pefabloc (Merck, Darmstadt, Germany)). Samples were sonicated 4×5 seconds on ice and spun at 7000 g for 10 minutes at 4°C. Protein content in the clear supernatant was determined using a bicinchoninic acid protein assay kit (Pierce, Rockford, Illinois, USA), using bovine serum albumin as the standard, and the supernatant was stored at −80°C.

Approximately 250 µg of protein were used to measure p38 MAPK enzymatic activity with the p38 MAPK assay kit
purchased from Cell Signalling (Beverly, Massachusetts, USA). A once diluted slurry of agarose hydrazide bound antibodies to phosphorylated (Thr\(^{180}\)/Yhr\(^{182}\)) p38 MAPK (40 µl) was used to selectively immunoprecipitate active p38 MAPK from the colon cell lysate (in 200 µl of cell lysis buffer) by gently shaking overnight at 4°C. The immunoprecipitate was washed twice with 500 µl of ice cold cell lysis buffer and twice with 500 µl of Ice cold kinase buffer (25 mM Tris, pH 7.5, 5 mM β-glycerolphosphate, 2 mM DTT, 0.1 mM Na\(_3\)VO\(_4\), and 10 mM MgCl\(_2\)) at 4°C. The kinase reactions were carried out in the presence of 200 µM ATP and 2 µg of ATF-2 fusion protein at 30°C for 30 minutes. After the reaction had been terminated by addition of 3x sodium dodecyl sulphate (SDS) sample buffer, the mixture was boiled for five minutes followed by brief centrifugation. ATF-2 phosphorylation was selectively measured by western immunoblotting. Samples were loaded on 10% SDS-polyacrylamide gels and transferred to polyvinylidene difluoride membranes (Millipore, Bedford, Massachusetts, USA). Subsequently, membranes were blocked in 5% non-fat dry milk in Tris buffered saline (TBS) supplemented with 0.1% Tween-20 and washed in TBS supplemented with 0.1% Tween-20. Membranes were incubated overnight using specific antibodies against phosphorylated (Thr\(^{180}\)) ATF-2 in 5% bovine serum albumin in TBS supplemented with 0.1% Tween-20. After three washes for 10 minutes, secondary antibody incubation was performed for one hour with GAR-PO in a 1:2000 dilution. After enhanced chemiluminescence using Lumilight® substrate (Boehringer Manheim, Germany), antibody binding was visualised using a Lumino imager (Boehringer Manheim).

A 250 µg sample of the colon cell lysate was suspended in a final volume of 80 µl SDS sample buffer; 25 µl were loaded on a SDS-polyacrylamide gel to measure p38 MAPK phosphorylation using western blotting, as described previously. p38 MAPK phosphorylation was detected using antibodies against phosphorylated p38 MAPK (Cell Signaling).

Antibody binding was quantified using image analysis software (Boehringer Manheim) and samples were compared with a control sample set on 100 arbitrary units.

**Statistical analysis**

Values are presented as mean (SEM) per treatment group. Differences between groups were analysed using the non-parametric Mann-Whitney U test. Body weight changes with time were analysed by one way analysis of variance: p<0.05 was considered significant. SPSS statistical software (SPSS inc., Chicago, USA) was used for analyses.

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**RESULTS**

**p38 MAPK is activated during TNBS colitis and inhibited by SB 203580**

To investigate the kinetics of p38 MAPK phosphorylation and enzymatic activity, colons from mice subjected to TNBS colitis were harvested over a nine day period. In mice subjected to TNBS and treated with vehicle, enhanced p38 MAPK enzymatic activity was observed, most evident five days after induction of colitis compared with basal levels (fig 1A). Strikingly, after this clear activation on day 5, a reduction in p38 MAPK enzymatic activity was observed. These data were confirmed by direct analysis of p38 MAPK phosphorylation status in colon homogenates which paralleled the kinetics of the kinase assay (fig 1B). The function of p38 MAPK was further explored by analysing the effects of SB 203580, a selective p38 inhibitor, in TNBS colitis. Treatment with SB 203580 almost completely prevented p38 MAPK enzymatic activity in colonic cell lysates induced by TNBS administration (fig 1A). Thus SB 203580 effectively inhibited p38 MAPK at the concentrations used (1 µM/kg body weight/day) in these mice. Analysis of p38 MAPK phosphorylation in SB 203580 treated mice showed no inhibition of phosphorylation status (fig 1B). These data confirm previous reports that SB 203580 binds to the ATP binding site thereby preventing enzymatic activity although not preventing phosphorylation of p38 MAPK by its upstream activators.

**Mice treated with SB 203580 showed significantly more wasting after induction of colitis**

Mice were intrarectally instilled with TNBS or saline on day 0 and day 7 and subsequently sacrificed on day 9. All 10 TNBS mice who received vehicle survived but in the TNBS treated mice who received SB 203580 three mice died during the course of the experiment. All mice intrarectally instilled with 0.9% NaCl, treated with either vehicle or SB 203580, survived. Body weights of all mice were recorded daily. Induction of colitis was paralleled by significant weight loss in both groups who received TNBS (fig 2). Mice treated with SB 203580 lost more weight compared with vehicle treated mice with colitis (p<0.05). Vehicle treated mice with colitis started recovering from their initial 15% weight loss on day 3 but in TNBS mice treated with SB 203580 body weight declined further to less than 80% of baseline weight. Mean body weight of TNBS colitis mice treated with SB 203580 remained lower then vehicle treated mice during the course of the experiment. Daily injection of SB 203580 in mice instilled with 0.9% NaCl did not induce weight loss, and body weight changes in these mice were similar to mice treated with vehicle. Hence the effects of SB 203580 on body weight changes during colitis were disease related and did not reflect a generalised effect on p38 MAPK inhibition on murine physiology.
Colon weights were significantly increased in MAPK inhibitor treated TNBS mice

Colon weight of saline instilled mice treated with vehicle or SB 203580 was similar (81.7 (5.3) mg and 81.3 (6.5) mg respectively) (fig 3). After induction of colitis, colon weight increased to 229.2 (22.2) mg in TNBS colitis, and treatment with SB 203580 caused a further significant increase in colon weight (289.1 (29.1) mg in SB 203580 treated mice) (p<0.05). TNBS administration caused a significant increase in baseline colitis score but on histopathological examination no differences were observed between vehicle and SB 203580 treated TNBS mice. The colitis score of TNBS mice was comparable between vehicle and SB 203580 treated mice (mean total score 13.9 (1.1) and 13.3 (0.4) respectively) (fig 4).

Cellularity of caudal lymph nodes in SB 203580 treated TNBS mice

The total number of cells in the CLN that drain the inflamed colon increased in TNBS colitis from 65.7 (17.1) \times 10^4 cells (saline) to 334 (36.7) \times 10^4 cells (TNBS) (fig 5). Significantly less CLN cells were found in SB 203580 treated mice with colitis compared with vehicle treated mice with colitis (188.6 (44.9) \times 10^4 cells in SB 203580 treated TNBS mice; p<0.05). This effect of SB 203580 in control mice was not observed in saline instilled mice (52.1 (11.2) \times 10^4 cells). Apparently, p38 MAPK is an important regulator of CLN cell number.

Cytokine production

Cytokine production on activation of T cells in the CLN was determined by CD3/CD28 double stimulation (fig 6). In TNBS...
Inhibition of p38 MAPK with SB 203580 in experimental colitis

Several reports describe the inhibitory activity of SB 203580, a specific p38 MAPK inhibitor. SB 203580 was reported to specifically inhibit induction of T cell apoptosis. Therefore, it seems that the inhibition in cell number could be better explained by diminished proliferation.

In line with previous reports, stimulated CLN cells produced significantly less IFN-γ.

There were several explanations for this unexpected observation. We have recently reported that IFN-γ does not inhibit disease mediator in TNBS colitis because mice that lack IFN-γ or its receptor are susceptible to TNBS colitis and, in fact, develop more severe disease. It was reported that IL-12 synthesis and specifically IL-12p40 synthesis are dependent on p38 MAPK activation.

We found a specific reduction in IL-12p70 and no changes in IL-12p40 concentrations after SB 203580 treatment. The pathogenic importance of IL-12 in TNBS colitis is well known and IL-12 neutralizing antibody treatment is highly protective in this model.

It should be noted that the two IL-12 subunits, p35 and p40, have different biological functions. IL-12p35 deficient mice are protected against the development of TNBS induced colitis, and in this situation neutralisation of p40 restores the normal sensitivity to TNBS. Hence IL-12p40 is protective, possibly through the activity of IL-12 homodimers or dimerisation with another ligand. Against this background, the combination of a reduction in IL-12p70 without an effect on IL-12 p40 concentration would be expected to be protective but this was not substantiated in our study. The observed changes may have been too small to result in a therapeutic effect or IL-12p35 may have formed heterodimers with a ligand other than p40. It is known that heterodimers of IL-12p35 and Epstein-Barr virus induced gene 3 have proinflammatory effects.

The enhanced wasting observed during colitis in p38 MAPK inhibited animals coincided with enhanced TNF-α production. In accordance with this finding, TNF-α production of T lymphocytes was reported to be only partially dependent on p38 MAPK activation.

Furthermore, we have recently shown that SB 203580 increases TNF-α production by macrophages and increased TNF-α production in mast cells has been reported. TNF-α plays an important role in intestinal inflammation, as confirmed in Crohn's disease and experimental colitis.

Alternatively, it is possible that activation of p38 MAPK occurs downstream of the disease perpetuating signal transduction elements and that inhibition at this level does not affect disease severity. We consider it more likely that cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly. T regulatory cells that play a protective role in intestinal inflammation also need p38 MAPK to function properly.

Another explanation for our findings is that inhibition of p38 MAPK results in accumulation of upstream activators of p38 MAPK. SB 203580 binds to the ATP binding site thus preventing phosphorylation of downstream targets, including MAPKAP 2 and ATF-2, although not preventing phosphorylation of p38 MAPK by its upstream activators MKK3 and MKK6.

MKK3 and MKK6 can phosphorylate downstream targets other than p38 MAPK, such as the JNK1 and JNK2 pathway. Because JNK2 is involved in Th1 differentiation, p38 MAPK inhibition may paradoxically increase the severity...
of Th1-mediated diseases. It is however also possible that the observed effects are specific for the inhibitor or species used in these experiments and studies with other p38 MAPK inhibitors are needed to elucidate this.

In summary, treatment with the p38 MAPK inhibitor SB 203580 does not ameliorate TNBS colitis although it does prevent IFN-γ and IL-12p70 production. This indicates that p38 MAPK may have a broader role in the mucosal immune response and is not only responsible for the production of proinflammatory cytokines but may also be involved in counterregulatory responses.

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