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Case report

Multicentric Castleman's disease and Kaposi's sarcoma in a cyclosporin treated, HIV-1 negative patient: case report

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Abstract

**Background:** Multicentric Castleman's disease (MCD) is a rare disease, but is more frequent in AIDS patients. MCD has only been reported twice before in patients receiving immunosuppressive therapy after renal transplantation, and never in patients receiving immunosuppressive therapy without transplantation. About half of the cases of MCD are human herpesvirus 8 (HHV8) – related, in contrast to Kaposi's sarcoma, a more common complication arising after immunosuppression, where the virus is found in virtually all cases.

**Case presentation:** We report a HIV-1 negative, non-transplant patient who developed HHV8-associated multicentric Castleman's disease and Kaposi's sarcoma after 17 years of immunosuppressive treatment with cyclosporin A for a minimal change nephropathy. Chemotherapy with liposomal doxorubicin resolved both symptoms of multicentric Castleman's disease and Kaposi's sarcoma in this patient. A concomitant decline in the HHV8 viral load in serum/plasma, as determined by a quantitative real-time PCR assay, was observed.

**Conclusions:** Multicentric Castleman's disease can be a complication of cyclosporin A treatment. Both multicentric Castleman's disease and Kaposi's sarcoma in this patient were responsive to liposomal doxorubicin, the treatment of choice for Kaposi's sarcoma at the moment, again suggesting a common mechanism linking both disorders, at least for HHV8-positive multicentric Castleman's disease and Kaposi's sarcoma.

HHV8 viral load measurements can be used to monitor effectiveness of therapy.

**Background**

Castleman’s disease, also called angiofollicular or giant lymph node hyperplasia, is a clinically heterogeneous entity that can be either localized (unicentric), or multicentric. The unicentric form, histopathologically described as the hyaline-vascular subtype, is more common, is not associated with HHV8-infection, and can mostly be treated effectively with radiotherapy or surgery.
as it presents as a solitary mass [1]. The multicentric form is less well understood: it is an atypical lymphoproliferative disorder of a plasma cell type, and is related to immune dysfunction. A mixture of both hyaline-vascular and plasma-cell variants can also be found [2]. Patients with MCD often develop malignancies like Kaposi’s sarcoma and non-Hodgkin’s lymphoma [2].

Human herpesvirus 8 (HHV8, or Kaposi’s sarcoma-associated herpesvirus) infection is present in nearly 100% of multicentric Castleman’s disease (MCD) cases associated with HIV-1 infection, and in about 50% of cases that are HIV-1 negative [3,4]. Recently, it has been proposed that HHV8-associated MCD is histologically distinct from HHV8-negative MCD [5,6]. Besides MCD and KS, HHV8 is also involved in a third disorder called primary effusion lymphoma (PEL). Interestingly, the HHV8-infected cells in MCD and PEL are of the B-cell lineage [7], while HHV8-infected cells in KS are presumably endothelial cells with macrophage characteristics [8].

Clinically, HHV8-positive MCD is more aggressive, and has a poor prognosis with a median survival of less than 30 months [2,4,9,10]. MCD, whether or not HHV8-associated, is generally treated with combination chemotherapy (CHOP) and steroids [11].

Case presentation
In August 2002, a 38-year old homosexual, HIV-negative Dutch male was admitted to our hospital with high fever, night sweats, dyspnoea, coughing, weakness, hepatosplenomegaly, peripheral lymphadenopathy and oedema after returning from a holiday in Spain. Medical history revealed a minimal change nephropathy, for which he had been taking cyclosporin A (Neoral) for the past 17 years (5 mg/kg per day). Laboratory analysis revealed anaemia (haemoglobin = 4.6 mmol/l), thrombocytopenia (101 × 109/L), normal WBC count and differential, renal failure (creatinine = 218 µmol/l), hypoalbuminemia (24 g/l), and elevated CRP (133 mg/l). Diagnostic assays for HIV-1 were performed and repeatedly negative. No acute CMV or EBV infection was detected (by serology and PCR analysis). Chest X-ray showed interstitial infiltrates, however, bronchoalveolar lavage did not reveal an infectious agent. CT-scans revealed hepatosplenomegaly and enlarged abdominal lymph nodes. A bone marrow biopsy showed an active haematopoiesis in all cell lines without any abnormalities.

Histological examination of an axillary lymph node biopsy (Fig. 1A) showed a multifocal proliferation of spindle cells. The diagnosis of Kaposi’s sarcoma was confirmed by positive staining of these spindle cells with antibodies to factor VIII-related antigen (not shown) and HHV8 (Fig. 1B). The remainder of the lymph node showed partly involuted germinal centres, surrounded by small lymphocytes. A number of these lymphocytes also stained positive for HHV8, as seen in multicentric Castleman’s disease (Fig. 1C). The clinical syndrome and the histological findings were compatible with KS and MCD caused by a HHV8 infection in a long-term immunocompromised patient. HHV8 DNA load in serum or plasma was determined by real-time quantitative PCR [12], and was found to increase from 9.0 × 10^5 copies/ml at the time of admission to 6.3 × 10^6 copies/ml before treatment start (Fig. 2).

Despite discontinuation of cyclosporin A and supportive care his clinical condition deteriorated with fever, dyspnoea, oedema and exhaustion, together with the development of oral and facial KS lesions. Liposomal doxorubicin (Caelyx 20 mg/m^2) was initiated at three-week intervals, ten days after discontinuation of Neoral. Although Caelyx is nowadays the drug of choice to treat KS [13], it has not been reported as a treatment for MCD, however, it has been shown to lower the HHV8 load [14], suggesting it may also have an effect on other HHV8-related diseases. HHV8 viral load has earlier been associated with clinical outcome in both KS and MCD, whereby a decrease in viral load measured in blood was linked to a favourable outcome [15-18], and an increase in HHV8 viral load with exacerbations in clinical symptoms [19-21].

Within two weeks after treatment start he clinically improved with disappearance of hepatomegaly, lymphadenopathy, and pulmonary infiltrates, and all laboratory parameters became normal. Simultaneously, the HHV8 load dropped to 7.4 × 10^4 copies/ml, but started to rise again after two weeks to 1.7 × 10^5 copies/ml, with recurrence of his clinical symptoms. After the second course of Caelyx, the HHV8 load decreased to 1.1 × 10^3 copies/ml. The treatment response was again transient and three weeks later his HHV8 load had risen to 2.4 × 10^4 copies/ml. After the third Caelyx treatment the viral load was permanently reduced to around the detection limit of the assay (1.66 × 10^2 copies/ml). Another three courses of Caelyx were given. Fourteen months later he is still disease free without fever, lymphadenopathy or KS lesions. Immunosuppressive therapy with cyclosporin A was discontinued, which is expected to contribute to the ongoing remission.

KS specifically occurs under cyclosporin A therapy in about 0.87% of renal transplant patients [22]. MCD however, is rarely seen under immunosuppressive treatment after transplantation [23,24], and has never been reported before in non-transplant patients treated with cyclosporin A [25].
Retrospective analysis of the stored serum samples with the Diavir HHV8 peptide mix ELISA (Biotrin International, Dublin, Ireland) showed that this patient was already IgG antibody-positive with very high titres at presentation (result not shown), making an acute HHV8 infection unlikely. As expected, all subsequent serum samples were also HHV8 antibody-positive (results not shown). So, the relatively late development of MCD in this homosexual patient (arising only after 17 years of cyclosporin treatment) is most probable due to a reactivation of an existing HHV8 infection. However, the HHV8 infection might be younger than the start of cyclosporin treatment 17 years ago, and as no earlier serum samples were available, it is difficult to estimate the time period elapsed between HHV8 infection and development of MCD. In The Netherlands, HHV8 infections are more common among men having sex with men than in the general population or any other risk group [26], making homosexuality the main risk factor for HHV8 acquisition, with iatrogenic immunosuppression being the risk factor for subsequent MCD development in this patient.

In general MCD has a poor prognosis with a rapidly fatal clinical course [4,9,10], therefore it is remarkable that in this case the patient responded well to Caelyx therapy, showing a concomitant decline of the HHV8 load in blood. Interestingly, a case of EBV-negative post-transplantation lymphoproliferative disorder (PTLD), attributed to a transplantation-associated HHV8-infection in a 17 year-old boy, responded only to a combination of...
liposomal daunorubicin and alpha-interferon [27]. EBV-positive PTLD’s, but not EBV-negative cases, have been found to respond to withdrawal of immunosuppression alone [27]. Possibly, HHV8-associated lymphoproliferative disorders share common features, and require similar treatment.

Conclusions
MCD can arise as a complication of cyclosporin treatment. HHV8 viral load measurements are indicative of effectiveness of therapy in HHV8-associated MCD.

MCD, as well as KS, in this patient was responsive to liposomal doxorubicin, the treatment of choice for KS at the moment, again suggesting a common mechanism linking both disorders.

List of abbreviations used
MCD = multicentric Castleman’s disease, KS = Kaposi’s sarcoma, PEL = primary effusion lymphoma, HHV8 = Human herpesvirus 8, HIV-1 = human immunodeficiency virus type 1, WBC = white blood cell count, CRP = C-reactive protein, CMV = Cytomegalovirus, EBV = Epstein-Barr virus, PCR = polymerase chain reaction, PTLD = post-transplantation lymphoproliferative disorder.

Competing interests
None declared.

Authors’ contributions
JMB and MHJvO treated the patient and organised clinical and laboratory investigations, AMP and MC performed the real-time HHV8 PCR assays, JFW did the other bacterial and viral diagnostic assays, LAN did histopathological

Figure 2
HHV8 DNA load of consecutive serum/plasma samples from the case measured with real-time PCR. Arrows indicate the chemotherapy courses. All samples were HHV8 IgG antibody-positive.
analysis and immunohistochemistry, and ACvKD drafted the manuscript. All authors read and approved the final manuscript.

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