Mutation screening in Borjeson-Forssman-Lehmann syndrome: identification of a novel de novo PHF6 mutation in a female patient

Published in:
Journal of Medical Genetics

DOI:
10.1136/jmg.2005.033084

Citation for published version (APA):
Mutation screening in Börjeson-Forssman-Lehmann syndrome: identification of a novel de novo PHF6 mutation in a female patient

J Crawford, K M Lower, R C M Hennekam, H Van Esch, A Mégarbané, S A Lynch, G Turner and J Gécz

J. Med. Genet. 2006;43;238-243; originally published online 1 Jul 2005; doi:10.1136/jmg.2005.033084

Updated information and services can be found at:
http://jmg.bmj.com/cgi/content/full/43/3/238

These include:

References
This article cites 17 articles, 5 of which can be accessed free at:
http://jmg.bmj.com/cgi/content/full/43/3/238#BIBL
1 online articles that cite this article can be accessed at:
http://jmg.bmj.com/cgi/content/full/43/3/238#otherarticles

Rapid responses
You can respond to this article at:
http://jmg.bmj.com/cgi/eletter-submit/43/3/238

Email alerting service
Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article

Topic collections
Articles on similar topics can be found in the following collections

Other Neurology (3647 articles)
Genetics (3952 articles)

Notes

To order reprints of this article go to:
http://www.bmjjournals.com/cgi/reprintform

To subscribe to Journal of Medical Genetics go to:
http://www.bmjjournals.com/subscriptions/
SHORT REPORT

Mutation screening in Börjeson-Forssman-Lehmann syndrome: identification of a novel de novo PHF6 mutation in a female patient

J Crawford, K M Lower, R C M Hennekam, H Van Esch, A Mégarbané, S A Lynch, G Turner, J Gécz

Background: Börjeson-Forssman-Lehmann syndrome (BFLS; MIM 301900) is an infrequently described X linked disorder caused by mutations in PHF6, a novel zinc finger gene of unknown function.

Objective: To present the results of mutation screening in individuals referred for PHF6 testing and discuss the value of prior X-inactivation testing in the mothers of these individuals.

Results: 25 unrelated individuals were screened (24 male, one female). Five PHF6 mutations were detected, two of which (c.940A→G and c.27_28insA) were novel. One of these new mutations, c.27_28insA, was identified in a female BFLS patient. This was shown to be a de novo mutation arising on the paternal chromosome. This is the first report of a clinically diagnosed BFLS female with confirmed PHF6 mutation. In addition, the X-inactivation status of the mothers of 19 males with suggested clinical diagnosis of BFLS was determined. Skewed (>70%) X-inactivation was present in five mothers, three of whom had sons in whom a PHF6 mutation was detected. The mutation positive female also showed skewing.

Conclusions: The results indicate that the success of PHF6 screening in males suspected of having BFLS is markedly increased if there is a positive family history and/or skewed X-inactivation is found in the mother.

Börjeson-Forssman-Lehmann syndrome was first described in 1962 in a family with severe intellectual disability, characteristic craniofacial features, microcephaly, hypogonadism, obesity, short stature, and epilepsy. Identification of the BFLS gene, PHF6, has enabled the causal mutation in this original family to be determined. A recent clinical review of 25 affected males from nine unrelated families with PHF6 mutations showed that the phenotype is seldom as severe as that seen in the original patients. Clinical manifestations are quite variable, with the most consistent features being initial hypotonia, mild to moderate intellectual disability, large fleshy ears, underdeveloped genitilia, gynaecomastia, truncal obesity, tapering fingers, and shortening of the fourth and fifth toes. Heterozygote females may have a milder similar clinical phenotype, which can include hypothyroidism; however, many carrier females appear unaffected. Our initial research indicated that the majority of females carrying known PHF6 mutations show highly skewed X-inactivation in their blood leucocytes. Plenge et al analysed X-inactivation skewing in female carriers from 24 families segregating 20 distinct X linked mental retardation (XLMR) disorders. They found that approximately 50% of XLMR carriers showed markedly (>80%) skewed patterns of X-inactivation, compared with approximately 10% of the control population. In several instances where obligate carriers were found to have pronounced skewing, the pattern of skewing was consistent throughout the family. Screening of PHF6 in individuals with a suggested clinical diagnosis of BFLS immediately following characterisation of the gene identified mutations in all seven of a group of familial BFLS cases, but in only two of 13 sporadic cases. Based on our initial observation of frequently and consistently skewed X-inactivation in PHF6 mutation carriers, we investigated the use of X-inactivation screening in the mothers of males with a suggested clinical diagnosis of BFLS as an indicator of their PHF6 mutation carrier status. Here we outline our experience with PHF6 mutation screening, supply a brief family history of the previously unreported BFLS patients identified in this study, and compile all currently reported PHF6 mutations.

METHODS

Patients
Since publication of the BFLS gene PHF6, 25 patients with clinical features suggestive of BFLS have been referred for mutation testing to our laboratory. Photographs and detailed clinical history of the five patients found to have a PHF6 mutation were obtained from the referring clinicians before mutation detection. A brief family history of the three unpublished patients is outlined in the results section of this paper. Patient consent and ethics approval were obtained locally by the referring clinician for all cases.

X-inactivation studies
X chromosome inactivation status was studied in DNA extracted from white blood cells from the mothers of boys with a suggested clinical diagnosis of BFLS, and in the single girl clinically diagnosed as having BFLS (QIAamp DNA blood maxi kit, Qiagen Inc, Valencia, California, USA). The X-inactivation analysis was undertaken using three polymerase chain reaction (PCR) based assays: HpaII digestion of DNA followed either by amplification of the androgen receptor (AR) gene polymorphic CAG repeat, or of the FMR1 gene variable CGG repeat, or amplification of the phosphoglycerate kinase-1 (PGK-1) gene followed by digestion of the polymorphic BstXI site. PGK-1 analysis was carried out only when the patient was uninformative at both AR and FRAAXA loci. Initial analysis of AR and FMR1 repeat regions involved incorporation of radioactive 32P into each amplion and electrophoresis of resultant labelled products on denaturing gels. Conclusions: X-inactivation analysis was carried out only when the patient was uninformative at both AR and FRAAXA loci.

Abbreviations: AR, androgen receptor; BFLS, Börjeson-Forssman-Lehmann syndrome; ORF, open reading frame; XLMR, X linked mental retardation

www.jmedgenet.com
5% acrylamide gels. We have also used AR and FMR1 amplification with fluorescently labelled primers (FAM and HEX, respectively) with subsequent analysis of labelled amplicons on a 3100-Avant genetic analyser (Applied Biosystems, Foster City, California, USA). The percentage of skewed X-inactivation in the original cell population from which the genomic DNA was derived was then determined by comparison of peak areas in digested versus undigested samples. Digestion conditions, primer sequences, and PCR conditions are available on request.

PHF6 mutation screening

Mutation screening was undertaken on either the genomic DNA or EBV transformed lymphoblast cell line RNA of the individuals with a suggested clinical diagnosis of BFLS. Genomic DNA was isolated from patient’s white blood cells using the QIAamp DNA blood maxi kit (Qiagen). Screening was undertaken on exons 2 to 10, a region that encompasses the entire PHF6 coding region. Primers were designed in intronic sequences such that coding sequences and a minimum of 20 base pairs (bp) of flanking intronic sequence could be analysed by direct sequencing (table 1). Cycling conditions for exons 2, 3, 5, 7, and 8 were as follows: 10 cycles of 94°C for 30 seconds, 60°C for 30 seconds, and 72°C for 30 seconds, followed by 25 cycles of 94°C for 30 seconds, 55°C for 30 seconds, and 72°C for 30 seconds. Exons 4, 6, 9, and 10 were amplified with 94°C for 30 seconds, 55°C for 30 seconds, and 72°C for 30 seconds for 35 cycles. Final extension for all PHF6 genomic PCRs was 72°C for seven minutes. Total RNA was isolated, when available, from patient lymphoblast cell lines with the RNeasy mini kit (Qiagen); 1 μg of total RNA was oligo-dT reverse transcribed using SuperScript II RNAse H- reverse transcriptase (Gibco BRL, Gaithersburg, Maryland, USA) following the manufacturer’s instructions. The PHF6 open reading frame (ORF) was

<table>
<thead>
<tr>
<th>Primer name</th>
<th>Sequence (5’ to 3’)</th>
<th>Size (bp)</th>
<th>Product (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PHF6ex1-F</td>
<td>GTA TCA ACG CTG GGG TC</td>
<td>20</td>
<td>273</td>
</tr>
<tr>
<td>PHF6ex1-R</td>
<td>GCC CAC GCG CAT CCT CC</td>
<td>17</td>
<td></td>
</tr>
<tr>
<td>PHF6ex2-F</td>
<td>AAA ATT AAC ATT GTC GCC CTT C</td>
<td>22</td>
<td>287</td>
</tr>
<tr>
<td>PHF6ex2-R</td>
<td>GAA CAT TCA TGT GTI ATT AAG AG</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6ex3-F</td>
<td>GCC ATT TTT ACT AGA AAA TTA CC</td>
<td>23</td>
<td>235</td>
</tr>
<tr>
<td>PHF6ex3-R</td>
<td>TTC TCA AGA GCC AGA AAT ACC</td>
<td>21</td>
<td></td>
</tr>
<tr>
<td>PHF6ex4-F</td>
<td>TTT TCA ATA ACC AAT TIG TTT TCC</td>
<td>24</td>
<td>310</td>
</tr>
<tr>
<td>PHF6ex4-R</td>
<td>TTT TTA CAA AAG CCC AAA GAA AG</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6ex5-F</td>
<td>TGT GGT GAA GTG TAA AGC TG</td>
<td>20</td>
<td>245</td>
</tr>
<tr>
<td>PHF6ex5-R</td>
<td>GAA TGT TGA GAT TCT TAT TG</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6ex6-F</td>
<td>AAC TAA TAC TTA TTT TGA GAT TGG</td>
<td>24</td>
<td>314</td>
</tr>
<tr>
<td>PHF6ex6-R</td>
<td>CAT TTC AAA TGA TGA ACT TTA CC</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6ex7-F</td>
<td>ATG TTA AGT AAG CTT GAA ATA CC</td>
<td>23</td>
<td>250</td>
</tr>
<tr>
<td>PHF6ex7-R</td>
<td>CAA AAT TGG GCT TAA AAG AAC C</td>
<td>22</td>
<td></td>
</tr>
<tr>
<td>PHF6ex8-F</td>
<td>CAT TTA ATG TTT CTC TCA TAA GG</td>
<td>23</td>
<td>265</td>
</tr>
<tr>
<td>PHF6ex8-R</td>
<td>ACT TTA AAT TTT CTT ATG ACT GG</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6ex9-F</td>
<td>TCT TTT TCA ATA GAA AAT AGC TG</td>
<td>23</td>
<td>276</td>
</tr>
<tr>
<td>PHF6ex9-R</td>
<td>TAA ACT AAT GTC ATC TAT TAA AGG</td>
<td>24</td>
<td></td>
</tr>
<tr>
<td>PHF6ex10-F</td>
<td>CAT CCA CTA ATG TTT GCA GG</td>
<td>20</td>
<td>261</td>
</tr>
<tr>
<td>PHF6ex10-R</td>
<td>ATA TAT CAG TGT GTA TGG TAT CC</td>
<td>23</td>
<td></td>
</tr>
<tr>
<td>PHF6polyA-F</td>
<td>TTT TGT TCT AAG AGA CAT TTA GG</td>
<td>23</td>
<td>285</td>
</tr>
<tr>
<td>PHF6polyA-R</td>
<td>AAA GCA AGA CTA AAA GGA ACA C</td>
<td>22</td>
<td></td>
</tr>
<tr>
<td>PHF6cDNAa-F</td>
<td>CAT TCT TIG AGA CTT AAA GTG G</td>
<td>22</td>
<td>605</td>
</tr>
<tr>
<td>PHF6cDNAa-R</td>
<td>GTT CCA TGG GAG GAT GTG G</td>
<td>19</td>
<td></td>
</tr>
<tr>
<td>PHF6cDNAb-F</td>
<td>AAC TGG AGC CCT CAT CAC C</td>
<td>19</td>
<td>756/1086*</td>
</tr>
<tr>
<td>PHF6cDNAb-R</td>
<td>GGC AGT AAA AAG TTA CAA ACC C</td>
<td>22</td>
<td></td>
</tr>
</tbody>
</table>

*Product sizes differ depending on the alternative splicing of intron between exons 10 and 11 (330 bp). bp, base pair.

Figure 1 Schematic representation of our experience with screening of the PHF6 gene in patients clinically diagnosed with Börjeson-Forssman-Lehmann syndrome.
amplified in two PCR products with PHF6-cDNA specific primers (table 1), under the following cycling conditions: initial denaturation at 94°C for two minutes, followed by 35 cycles of 94°C for 30 seconds, 56°C for 30 seconds, and 72°C for one minute. All amplifications were carried out on 100 ng of genomic DNA or 40 ng of cDNA, in a total PCR volume of 50 μl. PCR products were column purified (Qiagen PCR purification kit, Qiagen) and one or both strands were directly sequenced using Big Dye Terminator v3.0 or v3.1 sequencing chemistry (PE Applied Biosystems).

The following GenBank reference sequences were used: for PHF6-cDNA, NM_032335; for PHF6 protein, NM_032335; and for PHF6 genomic DNA, AL591668.

RESULTS
X-inactivation analysis of PHF6 mutation carrier females
We had access to DNA from 19 mothers of the 25 clinically referred BFLS patients. We have identified five mothers with skewed (>70%) X-inactivation, three of whom have sons with PHF6 mutations. In addition, we tested for, and found, highly (>90%) skewed X-inactivation in the only one BFLS diagnosed female. This pick up rate differs significantly from that seen for isolated males with a suggested clinical diagnosis of BFLS for whom the mother’s X-inactivation status was unknown, uninformative, or not skewed. In the 16 such cases screened, we failed to detect any PHF6 mutation (fig 1).

Mutation analysis in PHF6
We have conducted mutation screening in a cohort of 25 unrelated individuals clinically suspected of having Börjeson-Forssman-Lehmann syndrome. Screening of the PHF6 ORF has identified mutations in five of these individuals, four male and one female. Two of the males had the recurrent PHF6 mutation (case 1). A brief clinical history of the other two males and the female BFLS patient is outlined below. The c.2T→C/p.M1T mutation seen in case 2 has been identified in a previously reported family. Both the c.940A→G/p.I314V and c.27_28insA mutations, identified in cases 1 and 3 respectively, are unique to this study. The c.940A→G mutation (case 1) results in a replacement of isoleucine for valine at position p.314. This isoleucine is 100% conserved among man, orang-utan, cow, chicken, rat, mouse, xenopus, pufferfish, and zebrafish (data not shown). Photographs of the BFLS female patient (case 3) are presented in fig 2A, reproduced with permission. Mutation identification in this individual is outlined in fig 1B and described in further detail below. Family history appears to be a strong indicator of subsequent PHF6 mutation detection, with mutations identified in three of the five familial cases studied (fig 1). There were no PHF6 nucleotide changes detected in the other 20 patients screened and none of the mutations identified in this study was seen on at least 50 control chromosomes.

Identification of a novel PHF6 mutation in a female BFLS patient
Case 3 was unique in that the BFLS presenting proband was female (fig 2A). We first assessed the X-inactivation of this patient. Upon finding highly (93%) skewed X-inactivation, we screened her PHF6 locus on genomic DNA. Exonic sequencing revealed a single nucleotide insertion in exon 2 (c.27_28insA, fig 2B), presumably leading to a subsequent frameshift and addition of 21 novel amino acids to the PHF6 protein (p.G10fsX21). This mutation, which apparently results in a premature stop codon in exon 2, was the only sequence variant detected in the PHF6 open reading frame. Sequencing of DNA from the proband’s parents showed that the c.27_28insA mutation arose de novo, as it was not present in either parent. X-inactivation analysis indicated that the proband had the maternal chromosome active in 93% of peripheral blood cells (data not shown). The mother of the proband also had skewed X-inactivation (28:72). To determine the parental chromosome on which the c.27_28insA arose, we have taken advantage of an intronic SNP (intron 2, IVS2+203G/A) in close distance from the exon 2, where the c.27_28insA mutation lies. While the paternal chromosome had allele G at position IVS2+203 and the maternal chromosomes were both IVS2+203A, the proband is an IVS+203G/A heterozygote. Subcloning and subsequent sequencing of genomic PCR products across exon 2, intron 2, and exon 3 (primers PHF6ex2-F and PHF6ex3-R, 597 bp product) showed that the c.27_28insA mutation was on the chromosome with the IVS2+203G allele—that is, on the paternal chromosome (results not shown).

Brief family history of newly identified BFLS patients
Case 1 (c.940A→G/p.I314V)
The proband was born after 42 weeks with a birth weight of 3750 g. Psychomotor developmental delay was recognised early; he did not walk until two years three months and did not begin to talk until four years. He has an IQ of 61 and has major behavioural problems, one of the most pronounced being lack of any inhibition in sexual contacts. At 18 years he was 171.5 cm tall with a head circumference of 55.7 cm, moderate obesity, and marked gynaecomastia. External

Figure 2  Female patient with Börjeson-Forssman-Lehmann syndrome and an identified PHF6 mutation (case 3). (A) Two photographs of the patient, a side profile showing a shallow forehead, large ears, and fleshy ear lobes, and a frontal photograph showing a shallow forehead, deep set eyes, and prominent supraorbital ridges. (B) Sequencing traces illustrating reverse sequencing reactions of PHF6 exon 2 product of case 3 together with a control normal male trace; n, normal allele; m, mutant allele with the single nucleotide insertion, c.27_28insA; the arrow indicates the position of insertion of ‘A’ where the sequences of the n and m alleles start to differ; arrowheads indicate the orientation of the sequencing reaction; the filled circle indicates the proband, the empty square a control male. The written permission of the patient’s parents was obtained for reproducing the photographs in panel A.

www.jmendedgenet.com
genitalia showed normal post-pubertal development for age. His limbs showed hypermobility with Y-shaped cutaneous syndactyly on both feet. The X-inactivation testing on the mother of the proband showed a 100% skewed X-inactivation pattern.

Case 2 (c.2T→C/p.M1T)
The proband from this family was born at term after an uneventful pregnancy. His early milestones were delayed; he walked at four years and began saying a few words at five years. At approximately 15 years his height was 140 cm (<3rd centile), weight was 45 kg (5th centile), and occipito-frontal head circumference (OFC), 53.3 cm (10th–25th centile). On physical examination there was a proportionate short stature, triangular face, hyperplastic supraorbital ridges, deep set eyes, large ears, gynaecomastia, and lower abdominal obesity. External genital examination revealed a small penis and small testes with no evidence of pubertal change. The X-inactivation testing on the mother of the proband showed random pattern of X-inactivation.

Case 3 (c.27_28insA/p.G10fsX21)
This proband is a girl who presented with BFLS-like symptoms. The pregnancy was uneventful. Her birth weight was 3972 g. She never crawled but she walked at seven to eight years; she has had a high level of thyroid labile, has poor spatial awareness and startles easily to noise. Testing of her blood DNA showed a 93% skewed X-inactivation pattern.

**PHF6 mutation summary**
Amalgamating the results of our study with those published by others gives a total of 12 familial and five sporadic cases of BFLS recognised so far with the identified PHF6 gene mutation. There are currently 12 different PHF6 mutations identified from 17 unrelated BFLS families, with four of these mutations (c.2T→C; c.134G→A, c.769A→G, and c.1024C→T) being recurrent (fig 3). Missense mutations are the most frequent ones, found in 10 of the 17 cases. Interestingly, PHF6 mutations are not clustered and they do not seem to predominantly affect the only two recognised functional domains of the protein, the PHD-like zinc fingers. In fact just the opposite is seen, with the majority of mutations found outside the PHD finger regions. These observations, along with the lack of notable genotype-phenotype correlations, lead us to propose that the majority, if not all, PHF6 mutations might cause loss of function of the PHF6 protein.

**DISCUSSION**
Since the identification of the causative gene for BFLS in 2002, we have analysed 25 clinically diagnosed cases received from colleagues worldwide (fig 1). Given the size of the gene (10 exons) we preferentially screened affected individuals on cDNA generated from lymphoblast cell line (LCL) RNA; however, as LCLs were not available for every patient, we also undertook screening on genomic DNA (supplementary table 1, which can be viewed on the journal website: http://www.jmedgenet.com/supplemental). Analysis of mutation detection outcomes indicated that, as expected, family history was a strong indicator of subsequent PHF6 mutation detection, with mutations identified in three of the five referred familial cases. Skewed X-inactivation in carrier females was another attribute that indicated increased likelihood of PHF6 mutation detection, with PHF6 mutations being carried by three of the five mothers with skewed X-inactivation. The single clinically diagnosed BFLS female in whom we identified a PHF6 mutation also showed a skewed pattern of X-inactivation. A detailed flow chart of the results of PHF6 screening is shown in fig 1.

Based on the analysis of the initial BFLS families we expected to see concordance between BFLS carrier status and skewing of X-inactivation. However, as the number of characterised BFLS cases has increased, the percentage of obligate carrier females with highly skewed (>90%) X-inactivation has decreased. Combining the data from this and our previous study, we have now data on carrier females...
from 13 unrelated BFLS families. Of these, in three families the carrier females show random X-inactivation and in a further three the carrier females show approximately 70% skewing. However, carriers in the seven remaining families show >90% skewed X-inactivation as tested on blood leucocyte DNA. Taken together, in about 50% of BFLS families obligate carriers show highly (>90%) skewed X-inactivation. There does not seem to be a correlation between the X-inactivation and the type or location of the **PHF6** mutation. While in three families with the c.1204C→T (p.R342X) mutation we see occasionally skewed (>70%), but more frequently highly skewed (>90%), X-inactivation in blood of obligate carriers, 13 for the c.2T→C (p.M1T) mutation one obligate carrier is 100% X-skewed and one shows random X-inactivation (case 2 in this study).

It must be recognised that although knowledge of X-inactivation skewing in the mother of the proband, together with familial occurrence of the BFLS phenotype, is the best predictor of the detection of a **PHF6** mutation, some **PHF6** mutations (including de novo mutations) may be missed if screening is based solely on these criteria. There also remains the question of the genetic cause in patients with clinically diagnosed BFLS for whom we have not detected a **PHF6** mutation. Underlying genetic heterogeneity (well known for XLMR disorders) provide one explanation. However, no BFLS families suggesting this have been reported. Another, more likely explanation is that the clinical features that raise the possibility of a BFLS diagnosis are so protean that over-diagnosis and overlap with Prader-Willi, Wilson-Turner, Cohen, and Klinefelter syndromes may be common. Alternatively, individuals diagnosed with BFLS and tested negative for **PHF6** mutations in the coding region could harbour mutations not detected by our screening strategy, such as changes in short or long range regulatory elements, copy number imbalances, or other rearrangements. Our current and previous 2 gene resequencing analyses of controls and patients have failed to detect any single nucleotide variation within the coding region of the **PHF6** gene, allowing us to rule out potential “splicing spoilers” as causative of BFLS.11

The **PHF6** mutations identified thus far are not clustered within a particular domain, for instance within the PHD-like zinc finger domains (in fact only one mutation occurs in this region), nor are they concentrated in any other region of the protein. The mutations occur along the length of **PHF6**, and include missense, nonsense, splice site, deletion, and insertion mutations (fig 3). There is no clear link between mutation type or location, and the severity of the disease.4 There is one potential mutational hot spot in exon 10, c.1024C→T, which has been found in three cases (of the 17 known) carrying a **PHF6** mutation. The nucleotide C at position c.1024 is part of a CpG dinucleotide, known to be often mutated in humans.15

The identification of a female BFLS patient with a **PHF6** mutation (case 3) is novel. Unlike many female **PHF6** mutation carriers studied so far, she is affected. Varying expression of clinical features in female carriers had been reported 10 and as such this case may represent an extreme. A likely explanation of such a varying heterozygous expression may lie in differences in X-inactivation skewing between tissues (for example, brain versus blood), or a probable non-cell-autonomous effect of the **PHF6** mutation harbouring cells on those with normal **PHF6**, similar to the observations made in Rett syndrome.10 16 The identification of additional female BFLS cases or the generation of a suitable animal model will help address this important issue.

In conclusion, we have screened a total of 25 unrelated individuals with suggested clinical diagnosis of BFLS (24 males and one female) and detected five **PHF6** mutations, two of which have not been identified previously (case 1 and case 3, fig 2). We assessed the X-inactivation status of the mothers of 19 of the clinically diagnosed BFLS patients. We saw skewed (>70%) X-inactivation in five of these cases, with three **PHF6** mutations subsequently detected (two p.R342X and one p.I314V (case 1)). We identified a further **PHF6** mutation in a family where the obligate carrier mother did not show a skewed pattern of X-inactivation (p.M1T (case 2)). The fifth mutation was identified in a female BFLS patient (p.G10fsX21; case 3, fig 2) and was identified as a de novo mutation arising on the paternal chromosome. Before the ascertainment of case 3, we had identified carrier females (ranging in phenotype from asymptomatic to mildly affected) only through an affected male relative. Case 3 was the only referred female BFLS patient. Clinically diagnosed female BFLS patients have previously been reported,47–49 but never with subsequent genetic confirmation of the diagnosis. Based on this finding and the published reports, mutation screening of the **PHF6** gene in clinically diagnosed BFLS female patients should be considered, particularly if they show a skewed pattern of X-inactivation.

**ACKNOWLEDGEMENTS**

We thank the members of the families and their clinicians for their participation in this study. Additionally we thank Bruce Bennetts for the information he provided with regard to the detection of non-random X chromosome inactivation. We would also like to thank Rebecca Clay, Sarah McDonnell, and Cathy Derwas for help with cell culture and Olivia McKenzie and Tod Fullston with molecular genetic analyses. This work was supported by the National Health and Medical Research Council of Australia.

The supplementary table can be seen on the journal website (http://www.jmedgenet.com/complemental).

**Authors’ affiliations**

J Crawford, K M Lower, J Gecz, Department of Genetic Medicine, Women’s and Children’s Hospital, and Department of Paediatrics, University of Adelaide, Adelaide, Australia

R C M Hennekam, Clinical and Molecular Genetics Unit, Great Ormond Street Hospital for Children, London, UK

H Van Esch, Centre for Human Genetics, University Hospital Leuven, Leuven, Belgium

A Mégarbané, Faculté de Médecine, Université Saint-Joseph, Beirut, Lebanon

S Ann Lynch, National Centre for Medical Genetics, Our Lady’s Hospital for Sick Children, Dublin, Ireland

G Turner, Hunter Genetics and University of Newcastle, New South Wales, Australia

Conflicts of interest: none declared

Correspondence to: A/Prof Jozef Gecz, Department of Genetic Medicine, Women’s and Children’s Hospital, 72 King William Road, North Adelaide, SA 5006, Australia; jozef.gecz@adelaide.edu.au

Received 21 March 2005
Revised version received 21 June 2005
Accepted for publication 23 June 2005
Published Online First 25 July 2005

**REFERENCES**


