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Breaking the vicious cycle of epileptogenesis

Focus on brain inflammation and matrix metalloproteinases

Broekaart, D.W.M.

Publication date

2020

Document Version

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Citation for published version (APA):

Broekaart, D. W. M. (2020). *Breaking the vicious cycle of epileptogenesis: Focus on brain inflammation and matrix metalloproteinases*. [Thesis, fully internal, Universiteit van Amsterdam].

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CHAPTER

8

Discussion

Epilepsy

Epilepsy is one of the most common neurological diseases, currently affecting 65 million people worldwide ¹. Despite the availability of a wide range of antiepileptic drugs (AED), around one-third of all epilepsy patients remain intractable ²⁻⁴. Moreover, there are currently no drugs available that can stop the development or progression of epilepsy. Therefore, the need to identify novel therapeutic targets is urgent. The main aim of this thesis was to investigate several neurobiological processes that are altered in the epileptogenic brain, such as brain inflammation, blood-brain barrier (BBB) disruption and extracellular matrix (ECM) dysregulation, and to use this knowledge to discover novel treatment strategies.

Brain inflammation in epilepsy

The immune system serves to defend the host from external or internal pathogenic threats. In the central nervous system (CNS), this system can be activated in response to infectious agents or pathological events such as hypoxia, ischemia, traumatic brain injury (TBI) and (prolonged) seizure activity ⁵⁻⁸, all of which can lead to the development of epilepsy. The importance of the immune system in epilepsies has been increasingly recognized in the last two decades ⁶⁻⁹. In chapter 2, we aimed to investigate inflammatory markers of both the innate and the adaptive immune response in human and experimental temporal lobe epilepsy (TLE). We observed that especially markers of the innate immune response were persistently upregulated in the epileptogenic hippocampus of TLE patients as well as during epileptogenesis in a rat model of TLE. The activation of the innate immune response, involving brain resident cells such as microglia and astrocytes, has previously been suggested to play a role in epilepsy ¹⁰⁻¹⁶. Accordingly, the presence of reactive astrocytes and microglia has been shown in the brain of patients with TLE ^{13,17-19}, tuberous sclerosis complex (TSC) ²⁰⁻²³ and malformations of cortical development (MCD) ^{24,25}. In these pathologies, cytokines and danger signal molecules are synthesized by activated cells of the innate immune system ^{18,22,24,26-29}. Looking at the adaptive immune system on the other hand, we observed that T-lymphocytes and dendritic cells were not or scarcely present in the hippocampus of TLE patients and in the post-status epilepticus (SE) rat model. Although cells of the adaptive immune system were more evident in other epilepsies, such as post-encephalitis epilepsy ⁸, focal cortical dysplasia (FCD) type II and TSC ^{8,30} as well as in several epilepsy rodent models ^{16,31,32}, our data suggest a major proepileptogenic role of the innate immune response in TLE. However, we cannot exclude the contribution of the adaptive immune system in the development of epilepsy, since the infiltration of cells belonging to the adaptive immune system is associated with BBB dysfunction and epileptogenesis ³¹⁻³³.

During the activation of the immune response, the proteasome serves important roles, being the most common intracellular degradative system. In chapter 3 and 4, we aimed to further investigate the proteasomal system in epilepsies. We found that the constitutive subunits $\beta 1$ and $\beta 5$ and the inducible subunits $\beta 1i$ and $\beta 5i$ were higher expressed in neurons and glia in the epileptogenic foci of patients with epilepsy secondary

to cortical malformations such as FCD and TSC and in TLE patients compared to their respective controls. Under normal physiological conditions, the constitutive form of the proteasome is ubiquitously expressed in non-immune cells where it modulates oxidative stress (OS), synaptic plasticity and gene transcription^{34,35} while the catalytically more efficient immunoproteasome is limited to immune cells. Consistent with our observations, expression of the immunoproteasome is induced in non-immune cells in response to brain inflammation and accompanying inducing factors such as cytokines, danger associated molecular pattern molecules (DAMPs)³⁶⁻³⁸, OS³⁹ and activation of the mammalian target of rapamycin (mTOR) signalling pathway⁴⁰. In order to investigate the involvement of the (immuno)proteasome during the development of epilepsy, we studied the expression of constitutive and inducible subunits in the post-SE rat model for TLE. We observed higher expression as compared to controls of both the constitutive and the immunoproteasome in the acute phase of epileptogenesis (1 day after SE induction) and an even more prominent expression during the latent phase (1 week post-SE) which continued throughout the chronic phase (3 months post-SE) when the animals experienced spontaneous recurrent seizures. This pattern coincides with the inflammatory response that peaked in the latent phase and remained high during the chronic phase. Moreover, we observed that expression of constitutive and inducible subunits was correlated with seizure frequency in post-SE rats, as well as in patients with MCD. Together, this suggests the involvement of the proteasomal system in progression of epilepsy.

The main function of the proteasome is to degrade pathogens and misfolded and oxidized proteins, thereby protecting the cell from possible pathogenic influences. However, its involvement has also been recognized in a range of pathological diseases including epilepsy (reviewed by^{41,42}). With the incorporation of the inducible subunits, the catalytic activity of the proteasome alters; an increase in trypsin- and chymotrypsin-like activity arises along with a decrease in caspase-like activity^{43,44}. The cells may therefore be more capable of degrading a wider variety of disease-related misfolded and damaged proteins, which primarily could serve a beneficial cause. In relationship to epilepsy, the proteasome is known to be involved in the turnover of proteins associated with neuronal survival, neurotransmission, spine- and dendritic growth^{35,45-48}. Rather than securing effective degradation, excessive proteasomal activity might also lead to uncontrolled off-target degradation. Moreover, the immunoproteasome has also been shown to activate the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) signalling pathway and to modulate proinflammatory conditions and OS responses⁴⁹⁻⁵⁴. Furthermore, the more diverse pool existing of pure constitutive, pure immuno- and all intermediate-type proteasomes gives the opportunity to a wide range of epitopes being presented on the cell surface⁵⁵. Together, this might lead to the prolonged activation and further recruitment of immune cells causing the enhancement of the immune response in the epileptogenic brain. These factors might result in a prolonged decrease of seizure threshold and the persistence of epileptic seizures.

Brain inflammation and blood-brain barrier dysfunction

During brain inflammation, microglia and astrocytes are assisted by immunocompetent cells such as neurons and cells of the BBB; endothelial cells and pericytes. Clustering of pericytes with microglia has been observed in the epileptogenic brain, likely contributing to BBB disruption^{56,57}. When endothelial cells become reactive, they do not only increase their expression of cytokines such as interleukin (IL)-1 β and its receptor¹⁸, but also the expression of adhesion molecules that facilitate the passage of blood-borne cells into the parenchyma^{30,97}. In chapter 2, we observed increased expression of osteopontin, a protein that promotes the migration of macrophages, in the human and rat epileptogenic brain along with a positive correlation between perivascular macrophages and BBB permeability.

BBB dysfunction does not only occur as a consequence of brain inflammation, but can also lead to the activation of astrocytes and microglia, which release cytokines and chemokines⁵⁸⁻⁶⁰ thereby inducing brain inflammation. It is known that extravasation of albumin can lead to the induction of these proinflammatory mediators⁶¹⁻⁶³. Additionally, inflammatory regulators, including cytokines, are known to travel through the compromised BBB and enhance local inflammatory responses. We observed increased expression of the chemokine CCL2, a chemoattractant for cells of the monocyte lineage including monocytes, macrophages and microglia, that can further induce the release of proinflammatory cytokines and is associated with BBB dysfunction^{30,64-68} as it also attracts blood monocytes^{69,70}. Similar to CCL2, other chemokines such as CCL3, CCL4, CX3CL1, CXCL12 and their receptors expressed by activated and functionally altered glial cells and neurons are known to affect BBB function, such as by attracting blood leucocytes into the brain^{31,71-74}.

Blood-brain barrier dysfunction and extracellular matrix organization

BBB dysfunction is observed in several neurological diseases, including patients with drug-resistant epilepsy^{11,18,75-91}. Additionally, we have shown in this thesis that BBB dysfunction was evident in cortical tubers of TSC patients. It is one of the earliest neuropathological characteristics after an insult such as SE, stroke or TBI and can last for hours to weeks and even months, implying that BBB dysfunction has a role in the development of epilepsy.

BBB dysfunction can occur as a direct result of the insult which can lead to trauma or ischemia to endothelial cells, but it can also be due to secondary events such as increased blood-pressure, hypoxia, decreased blood pH, increased metabolic activation and (as discussed in the previous paragraph) brain inflammation. The integrity of the BBB can be regulated by ECM modulatory molecules that regulate cell-cell and cell-matrix interactions. The remodelling of the ECM has been suggested to play a role in epileptogenesis⁹², which is confirmed by previous studies of our group^{21,93,94}. RNA sequencing studies in cortical tubers have indicated that ECM proteins and ECM remodelling are important processes in TSC patients^{21,93}. Additionally, by performing an RNA microarray study, Gorter *et al.* showed that several ECM regulators were increased during epileptogenesis in a post-SE rat model,

including several matrix metalloproteinases (MMPs). Similarly, using five different models of brain injury, Kim *et al.* showed that albumin exposure leads to transcriptional changes in genes related to ECM modulation, among which several MMPs⁵⁸. MMPs are a family of extracellular endopeptidases, responsible for the degradation of the virtually all proteins in the ECM and are thereby involved in many different processes such as synaptic plasticity and migration⁹⁵⁻⁹⁸. In this thesis, we have shown that gene and protein expression of MMP₂, MMP₃, MMP₉ and MMP₁₄ were increased both in cortical tubers of TSC patients and resected hippocampi of drug-resistant TLE patients. Interestingly, we observed increased MMP expression in regions of the brain where the BBB was disrupted, such as in cortical tubers and subependymal giant cell astrocytomas (SEGAs) of TSC patients (unpublished observations). In addition, we observed that expression of MMPs positively correlated with the presence of albumin in the hippocampus of TLE patients and patients who died after SE. This suggests that MMP dysregulation and BBB dysfunction are interrelated. MMPs are directly involved in the (dys)regulation of BBB integrity as their degradome includes several proteins of the basal membrane such as laminin, fibronectin, heparin sulphate and type IV collagen^{99-101,270}, and target tight junction proteins such as zona occludens-1, occludin and claudins²⁹³. This is supported by studies using animals models that have shown that disruption of the BBB is mediated by MMPs^{267,294,295}. Furthermore, using the post-SE rat model, we observed higher MMP expression compared to controls in animals experiencing spontaneous recurrent seizures as well as prior to the occurrence of spontaneous seizures (during the acute and latent phase after SE induction), suggesting that MMPs play a role in the development and progression of epilepsy. In case of a dysfunctional BBB, infiltration of leucocytes can take place, which are also a major source for MMP₉ providing a positive feedback loop for MMP-dependent proteolysis^{102,103}.

The activity of MMPs is governed by the four members of the endogenous tissue inhibitors of metalloproteinases (TIMPs). The relevance of TIMPs in human epilepsies has not been extensively studied, and only increased expression of TIMP₂ has been documented in FCD patients¹⁰⁴. In chapter 5 and 7, we examined TIMP expression in TSC and TLE patients and observed higher expression of TIMP₁₋₄ compared to control cases¹⁰⁵. Overexpression of TIMPs is also observed in animal models of epilepsy. Higher expression of *Timp1* and *Timp2* was observed in the hippocampus and/or the temporal lobe 1 day and 1 week after electrically-induced SE⁹⁴ and 15 days after kainic acid (KA)-induced SE¹⁰⁶. The increased expression of TIMPs might be the result of a compensatory mechanism of the brain to overcome the increased MMP-mediated proteolysis. However, the relationship between MMPs and TIMPs goes beyond mere inhibition of activity. TIMPs have been shown to interact with the propeptide of MMPs that physically blocks the active site of the enzyme rendering it catalytically inactive. Controversially, TIMP₂, an effective inhibitor of all MMPs, is also responsible for the activation of pro-MMP₂ with the use of two MMP₁₄ molecules^{107,108}, leading to cleavage of the propeptide. Furthermore, TIMP₂, TIMP₃ and TIMP₄ have been

shown to interact with the propeptide of MMP2 while TIMP1 and TIMP3 are able to interact with pro-MMP9¹⁰⁹. In order to fully understand the effect of increased TIMP expression in epilepsy, further investigation about the spatial and temporal interaction with MMPs is necessary.

Matrix metalloproteinases and brain inflammation

In chapter 5, 6, and 7, we observed increased MMP expression not only in neurons, but very prominently in glia cells as well, implying that they might be involved in processes governed by these cell types. Interestingly, higher expression of MMPs is observed in TLE patients with hippocampal sclerosis (HS) compared to TLE patients without HS, a pathology that is characterized by hippocampal neuronal cell death and reactive gliosis¹¹⁰. Various stimuli, such as cytokines, growth factors, reactive oxygen species, or cell–ECM interactions are known to be responsible for the transcriptional activation of MMPs^{111–114}. It is therefore tempting to speculate that the increase of MMPs in, particularly but not restricted to, glial cells such as astrocytes might be the result of the proinflammatory environment. Accordingly, we have shown that, in human foetal astrocytes and tuber-derived astrocyte-enriched cultures, MMP expression can be increased by mimicking a proinflammatory state through stimulation with IL-1 β , whose signalling pathway has been shown to be prominently activated in TSC^{21,22} and TLE^{18,115} and in experimental models of epilepsy^{18,116–121}. Transcription factors activator protein 1 (AP-1) and NF- κ B, have been associated with the innate immune response, and inflammation^{122,123} and are also known activators of several MMPs^{124,130}. Remarkably, MMPs are not only activated in response to brain inflammation, they also contribute to the aggravation of the inflammatory response. MMPs can activate cytokines such as tumour necrosis factor (TNF)- α , IL-1 β and IL-8 by cleavage of the propeptide^{131–133}, thereby sustaining their own activity in a proinflammatory environment.

The chicken or the egg? A vicious circle during epileptogenesis

We know that brain inflammation, BBB dysfunction and MMP dysregulation are associated with epilepsy and are likely to be involved in epileptogenesis. These processes are associated with a dysfunctional neuronal microenvironment which increases the propensity for seizures to occur. However, at this point, the chicken or the egg dilemma arises. Do these processes have a causal role in epileptogenesis or are they merely the result of the aberrant network activity present in the epileptogenic brain?

Several studies have shown that the activation of inflammatory responses can have both beneficial, neuroprotective effects as well as exacerbating effects leading to increased neuronal excitability (reviewed by^{9,134}). It has been shown that activation of the immune system occurs after (induced) seizure activity^{5,6,9,10,18,28,135}. Models of acquired epilepsy have indicated a rapid activation of danger signals such as HMGB-1 and associated toll-like receptors (TLRs) along with an induction of prostaglandins and molecules of the complement system^{17,136}. Moreover, an eminent increase in cytokine and chemokine

production is seen in response to epileptiform activity^{6,10,116-118,120,137-141}. Indeed, mRNA expression of cyclooxygenase (COX)-2, an important mediator of neuroinflammation, is rapidly upregulated (within 1 hour) after SE induction^{117,142-144}. A similarly acute response is seen for cytokine expression including IL-1 β and TNF- α after SE induction^{117,118,143-145}. However, activation of the inflammatory system is also known to result in seizure activity, a clear example being the increased preposition to seizures that is observed in response to fever^{146,147}. Furthermore, diseases in which a compromised immune system is the primary pathology, such as limbic encephalitis and several autoimmune diseases, are associated with increased susceptibility to seizures and epilepsy¹⁴⁸⁻¹⁵⁰. Local or systemic injection of lipopolysaccharide (LPS), a prototypical inducer of inflammation, can lead to decreased seizure threshold through the activation of TLR4 and the thereby induced release of DAMPs^{27,151}. Interestingly, proinflammatory molecules such as cytokines have a dichotomous role in epilepsy showing anticonvulsive effects at low molecular concentrations. For example, daily intraventricular injections of IL-1 β delayed amygdala kindling in rats and intrahippocampal administration of nanomolar amounts of TNF- α reduced seizures in mice¹⁵²⁻¹⁵⁴. However, those concentrations are greatly lower than those endogenously produced by seizures and administration of cytokines with seizure-relevant concentrations have led to a reduced seizure threshold and increased seizure duration^{137,138,155-157}, also mediated by uncoupling of astrocytes¹⁵⁸. The effect of cytokines on seizure induction is strengthened by the fact that treatment with cytokine-inhibiting molecules has anticonvulsive effects^{137,138,159}. Moreover, it has been shown that cytokines can act indirectly and directly on glutamate, NMDA, AMPA and GABA receptors leading to enhanced excitatory and diminished inhibitory transmission¹⁶⁰⁻¹⁶⁶. Since activated cells of the innate immune system can produce large quantities of cytokines, chronic activation of these cells can contribute to epileptogenesis.

In humans, BBB dysfunction is one of the earliest neuropathological alterations in response to brain insults such as SE, TBI or stroke^{11,167-175}. BBB dysfunction is well documented in animal models of epilepsy, in which it is not only observed during the chronic phase when animals have recurrent spontaneous seizures but already during the first hours to days after SE^{11,85,176-180}. It has been observed as early as 5 minutes after bicuculline-induced seizures in a guinea pig model¹⁸⁰. In response to altered neuronal excitability, hypoperfusion of blood vessels can occur, leading to cellular damage and inflammation^{174,181}. This indicates that BBB dysfunction can occur as a result of seizure activity¹⁸². However, when the BBB is opened in animals with recurrent seizures, it leads to an increase in seizure frequency¹¹. This suggests that in the presence of neuropathological alterations such as brain inflammation, BBB dysfunction can play a significant role in epileptogenesis. Likewise, focal BBB damage in post-traumatic epilepsy patients is associated with abnormal EEG activity^{169,170,183} and areas with a compromised BBB show focal slowing and more spikes^{184,185}. Under healthy physiological conditions, artificial opening of the BBB does not typically lead to seizures^{11,186-189}, though acute seizures have been observed occasionally^{14,188}. This underlines that the presence of a pathology is important in how the tissue reacts to BBB dysfunction¹⁸². The

molecular mechanisms by which BBB dysfunction can lead to epileptic activity are not fully unravelled. It is proposed that, on the membrane of astrocytes, extravasated albumin binds to transforming growth factor (TGF)- β receptors and affects potassium buffering leading to increased extracellular potassium levels. This not only lowers the action potential threshold and increases neuronal excitability¹⁹⁰, but also leads to proinflammatory and synaptogenesis, resulting in gradual development of epilepsy^{61,187,191}. It is also suggested that albumin uptake by neurons, mostly found in regions in which cell death occurs, participated in the apoptotic processes involved in epilepsy^{11,85}.

Due to the strong interactions with both brain inflammation and BBB dysfunction, it is not unreasonable that also MMPs are readily increased in response to seizure activity. Indeed, several animal studies support the induction of multiple members of the MMP family acutely after an initial insult such as an SE^{94,192-199}. For example, increased MMP2 and MMP9 activity has been observed as early as 8 hours after KA-induced in SE¹⁹⁷. Furthermore, in TBI patients, after which acute seizures or even epilepsy can develop, the largest increase in MMP9 concentrations in the pericontusional cortex was found during the first 24 h post-injury²⁰⁰. On the other hand, upregulation of MMPs can also result in seizure activity as shown by genetic manipulation in rodent models. Overexpression of *Mmp9* leads to higher susceptibility to kindling-induced epileptogenesis²⁰¹. Likewise, *Mmp9*-overexpressing mice had more seizures in response to TBI²⁰². Besides the aggravation of BBB dysfunction and neuroinflammation, MMPs are able to directly affect neuronal excitability because of their wide range of substrates. Due to their capabilities to degrade the ECM proteins of perineuronal nets (PNNs), a structure of specialized ECM, MMPs can cause dysregulation of the inhibitory interneurons surrounded by PNNs²⁰³⁻²⁰⁵. This leads to reduced inhibitory transmission which is a known pathological factor in epilepsy and brain injury^{58,206}. Furthermore, MMPs can target the propeptides of several growth factors, such as the brain-derived growth factor (BDNF), resulting in cell proliferation²⁰⁷⁻²¹⁰. By activating BDNF, MMP9 might be one of the driving forces behind mossy fibre sprouting and the thereby caused synchronized firing seen in the epileptogenic brain²¹¹⁻²¹³. MMPs are also involved in spine formation and remodelling, due to their abilities to cleave membrane-associated adhesion molecules and via integrin-signalling pathways^{214,215}. Similar to cytokines, MMPs have also been shown to directly act on AMPA²¹⁶ and NMDA receptors²¹⁴, indicating that they play an important role in the generation of seizures and development of epilepsy²¹⁷⁻²¹⁹.

Overall, it seems that although brain inflammation, BBB disruption and ECM dysregulation have important and essential functions under normal physiological conditions, the persistent activation of these processes can have deleterious effects especially in an already compromised environment such as the epileptogenic brain. As they can occur as a consequence of (prolonged) seizure activity, but more importantly, can also cause a reduced seizure threshold, their aberrant activation can lead to the aggravation of pathology and progression of epilepsy (Fig. 1). Therefore, interfering with this vicious cycle provides an interesting therapeutic target in order to inhibit or block epileptogenesis.

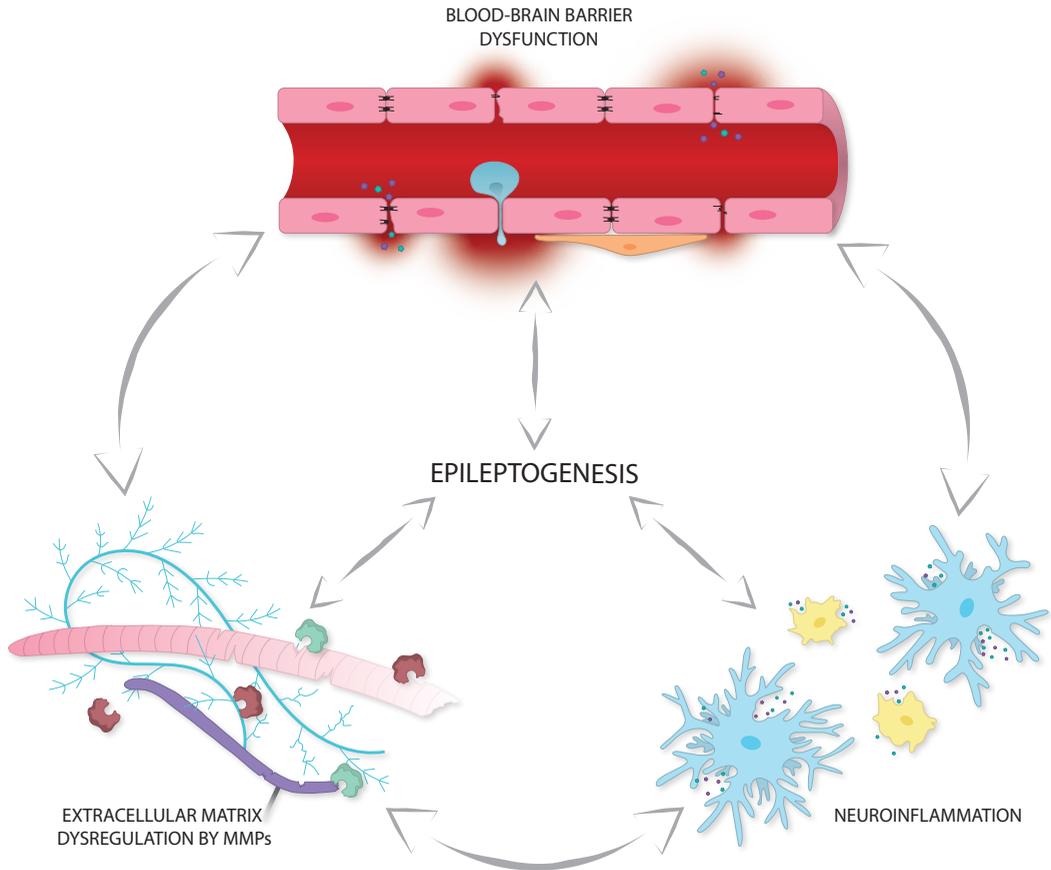


Figure 1. Schematic representation of the vicious circle consisting of blood-brain barrier dysfunction, neuroinflammation and extracellular matrix dysregulation by MMPs in the epileptogenic brain.

Blood-brain barrier (BBB) dysfunction: the BBB consists of several cells such as endothelial cells (pink) and pericytes (orange) that control the influx and efflux of molecules between the brain parenchyma and blood. Upon BBB dysfunction, infiltration of cells (blue) and permeability for blood-borne molecules (green/purple) increases. **Neuroinflammation:** among the many alterations during a proinflammatory state, reactive astrocytes (blue) and microglia (yellow) synthesize and secrete high number of inflammatory factors (green/purple) such as cytokines, chemokines and danger-associated molecules. **Extracellular matrix dysregulation by MMPs:** persistently activated matrix metalloproteinases (MMPs; green/red) can lead to dysregulation of extracellular matrix (ECM) proteins such as collagen (pink), fibronectin (purple) and proteoglycans (blue). As previously explained in detail, BBB dysfunction, neuroinflammation, ECM dysregulation by MMPs are interconnected and reciprocally aggravate each other. These pathological alterations can be enhanced by seizure activity, but most importantly can also lead to seizures and the development of epilepsy.

Therapeutic strategies targeting inflammatory processes and matrix metalloproteinases

The data presented in this thesis in combination with the available literature indicates that proinflammatory conditions, BBB dysfunction, and ECM dysregulation by MMPs are intertwined and are of great importance in the development and progression of epilepsy. The fact that increased expression and activation of markers of inflammation, BBB disruption and ECM remodelling are already evident before spontaneous seizures occur in animal models of epilepsy, indicates that there is a therapeutic time window. Some of the possible treatments targeting the aforementioned processes will be discussed in the following paragraphs.

Anti-inflammatory/immune-suppressive therapy

Anti-inflammatory/immune-suppressive therapy has been shown to be successful in several epilepsy cases. For example, treatment with the corticosteroid dexamethasone resulted in relief of seizure load, improved EEG and cognitive function in almost half of the included patients with refractory epileptic encephalopathy with continuous spike-and-wave during sleep ²²⁰. In another study, dexamethasone reduced the seizure frequency in 5/13 children with drug-resistant epilepsy ²²¹. In animals, dexamethasone reduced BBB leakage, ictogenesis and epileptogenesis after pilocarpine-induced SE in rats ¹⁵. However, the severe side effects of steroids have prevented long-term or widespread use of these drugs ²²². Moreover, anti-inflammatory treatment in animal models of epilepsy has proven challenging as different inflammatory mediators are induced at different time points. An initial wave of inflammation, including the induction of cytokines and its receptors is observed rapidly after experimental SE, followed by subsequent wave with COX-2 and prostaglandin induction as well as the upregulation of molecules of the complement system ^{5,9,116,223,224}. Therefore, great consideration has to be taken in terms of timing when targeting the inflammatory system as treatment for epilepsy. Several therapies targeting proinflammatory modulators, such as cytokines and chemokines, are reviewed in detail elsewhere ^{10,225}.

Potential novel therapeutic targets

The immunoproteasome

Enhanced activity of the proteasomal system might lead to uncontrolled off-target degradation, prolonged proinflammatory conditions and further recruitment of immune cells sustaining BBB dysfunction. Using hippocampal slices, Engel *et al.* (2017) investigated the effects of inhibition of the immunoproteasome and observed that KA- or glutamate-induced cell death was rescued with pretreatment of two different inhibitors, epoxomicin and MG132 ²²⁶. Also *in vivo*, intracerebroventricular pretreatment with epoxomicin resulted in less neurodegeneration 24 hours following KA-induced SE in mice, however without having an anticonvulsant effect. Specific inhibition of subunit β_5 or β_5i was investigated by Mishto *et al.* (2015) using brain slices from pilocarpine-induced SE rats with chronic epilepsy.

Pretreatment with ONX-0914, a specific inhibitor of β_5 , but not with PR825, a specific inhibitor of β_5 , resulted in a delay or prevention of AP-4-induced seizure-like events³⁸. Unfortunately, these inhibitors have bad bioavailability and poor capacities to cross the BBB suggesting that different target techniques are needed.

Considering the potential of the mTOR signalling pathway in activating the immunoproteasome and the overactivation of this pathway in many epilepsies such as FCD and TSC, we investigated whether rapamycin, an inhibitor of the mTOR signalling pathway able to pass the BBB, could attenuate overexpression of the (immuno)proteasome. Indeed, treatment of FCD-derived astrocytes and human foetal astrocytes with rapamycin reduced IL-1 β -induced increase of (immuno)proteasome subunits. Moreover, rapamycin treatment after electrically-stimulated SE in rats did not only result in a decreased seizure frequency, but also in decreased neuronal expression of subunits β_5 and β_5i . The beneficial effect of rapamycin has already been evident from studies including rodent models of epilepsy^{76,227-232}. It has shown to suppress seizures in post-SE models and other models of acquired epilepsy, as well as attenuate mossy fibre sprouting, cell death and neurogenesis^{229,233-237}. Furthermore, in response to rapamycin treatment, anti-inflammatory effects²³⁸⁻²⁴¹ and improvement of BBB integrity^{77,229,242,243} have been documented. A rapamycin derivative, everolimus, has been approved by the FDA for treating TSC patients with SEGA^{244,245} and as adjunctive treatment for adults and paediatric patients (>2 years of age) with TSC-associated partial-onset seizures as it leads to attenuation of the seizure burden^{246,247}. Further studies are needed to unravel the precise mechanism by which rapamycin affects proteasome activity and proteasome-dependent inflammation and epileptogenesis.

microRNAs

The first study about the relevance of microRNAs (miRNAs) in epilepsy arose almost a decade ago²⁴⁸ and from that time on, multiple studies investigated the identification and expression patterns of miRNAs in epilepsy patients and in several rodent models of epilepsy. Because of the widespread range of miRNA targets, research has focused more and more on the therapeutic potential of miRNAs as either a target or therapeutic agent itself. A recent meta-analysis evaluated pathways associated with experimental and human TLE and showed that among the most-enriched pathways is the regulation of the ECM²⁴⁹. Targeting these miRNAs might therefore be beneficial in patients in which an altered ECM milieu has a causative role or is contributing to the pathology. Evaluating the therapeutic potential of proinflammatory miR155, we used IL-1 β -stimulated human foetal astrocytes and observed that while overexpression of miR155 resulted in increased MMP3 expression, silencing of miR155 using an antagomiR, that inhibits the miRNA to bind to its target mRNA, reduced the IL-1 β -induced increase of MMP3. The anti-inflammatory miRNAs 146a and 147b were studied in relation to TSC, where they are highly expressed in dysmorphic cells²⁵⁰. In cortical tuber-derived cells, overexpression of miR146a or miR147b successfully attenuated the IL-1 β -induced increase in MMP3. Furthermore, TIMP expression that was decreased

after IL-1 β stimulation could be attenuated by miR146a or miR147b overexpression. Together, this suggests that inhibition of proinflammatory miRNAs and overexpression of anti-inflammatory miRNAs could restore the MMP/TIMP imbalance and resulting ECM alterations that are seen in the brain of patients with epilepsy(-related) disorders such as TLE and TSC.

The antiepileptogenic potential of miRNA silencing or overexpression has been investigated in a limited number of animal studies. Iori and colleagues observed reduced neuronal excitability in hippocampal slice cultures of mice injected with a miR146a synthetic mimic. Furthermore, in the KA-induced SE mouse model, they observed that intracerebroventricular injection of miR146a mimic delayed the onset, frequency and duration of acute seizures as well as the occurrence of spontaneous seizures²⁵¹. In another study it was shown that delivery of miR155 antagomiR prior to pilocarpine-induced SE resulted in less behavioural seizure severity and a trend towards better survival rates²⁵². Other miRNAs with antiepileptogenic potential supported by *in vivo* studies, are miR134a and miR135a. miR134, which is involved in activity-dependent dendritogenesis²⁵³, is shown to be upregulated in human^{254,255} and experimental TLE (reviewed by²⁵⁶). Silencing of this miRNA resulted in reduced spontaneous seizures after SE and increased delay in seizure onset^{254,255,257,258}. Inhibition of miR135a, which is upregulated specifically in neurons in the epileptogenic brain, has antiseizure effects in the chronic stage of intrahippocampal KA mouse model, most likely due to reduced expression of *Mef2a*, a key regulator of excitatory synapse density²⁵⁹.

Besides directly affecting disease-relevant pathways, miRNA modulation can also be regarded as an add-on to current drug treatment. Interestingly, a recent study showed that miR298 binds to the P-glycoprotein gene, reducing its ability to pump AEDs out of the brain²⁶⁰. Thus far, preclinical studies have not led to clinical trials that aim at treating brain diseases by modification of miRNA expression. However, miR155 modification is being investigated in a phase II trial for patients with T-cell lymphoma and phase II trials are planned for a long non-coding RNA-based treatment for Dravet syndrome²⁶¹. Challenges that need to be considered when aiming for miRNA modulation in disease are adverse inflammatory effects, charge-related toxicity, targeting the diseased brain area and routes of administration²⁶². For the latter, promising results have been obtained in experimental models of epilepsy using intranasal delivery^{263,264}. With regard to epilepsy, more research is needed about the medical utility of miRNAs, though miR155, miR146a and miR147b are interesting candidates and deserve further investigation.

Metalloproteinases

As described previously, prolonged expression and increased activity of MMPs can be devastating for brain homeostasis and can influence cellular physiology and brain circuitry. Inhibition of excessive MMP activity might therefore be an interesting therapy in several

neuropathologies such as epilepsy. Genetic manipulation in experimental models of epilepsy have already shown the effectiveness of MMP activity management, showing it has both positive effects on seizures or epilepsy development^{201,202,211} as well as on attenuation of BBB dysfunction^{102,265-267}.

Also natural products, such as tetracycline –produced by *Streptomyces*– and its derivatives are considered to inhibit MMPs, though the exact mechanisms of function are unknown. Treatment with doxycycline and minocycline has proven to restore BBB dysfunction²⁶⁸⁻²⁷³, although the effects on epileptogenesis have been moderate and model- and dose-dependent. Unfortunately, treatment with tetracycline derivatives can lead to severe side effects²⁷⁴⁻²⁷⁹. A case report of a patient with astrocytoma and drug-resistant epilepsy shows that treatment with 50 mg minocycline twice daily greatly decreased seizures but resulted in severe skin irritations after one month²⁸⁰. In chapter 7, we treated rats with a sub-toxic dose (45 mg/kg) of minocycline daily during rapid kindling and did not observe decreased seizure severity compared to vehicle-treated animals. Importantly, MMP9 activity did not change, indicating that this dose might not be sufficient to inhibit MMP9 activity in the epileptogenic brain.

Nowadays, crystallography has provided more insight into the several pockets of the MMP enzymes, which can contribute to drug design methods^{281,282}. It is now thought that the previously seen side effects of the broad-spectrum MMP inhibitors were due to the compounds' lack of specificity. Broad-spectrum targeting of MMPs resulted in inhibition of non-pathological MMPs as well as non-MMP inhibition of members of the a disintegrin and metalloproteinase (ADAM) and ADAM with thrombospondin motifs (ADAMTS) families²⁸³. For this reason, next generation inhibitors were developed to more specifically bind a selection of MMPs. For example, a thiol-based inhibitor SB-3CT was developed that specifically binds MMP9 and MMP2. Preclinical studies have shown that SB-3CT has neuroprotective effects after ischemia^{284,285}, and reduces astro- and microgliosis, hippocampal cell loss and BBB permeability after TBI^{286,288}. DP-b99, a MMP9-inhibitor, delays the onset and severity of pentylenetetrazole (PTZ)-induced seizures in mice²⁸⁹ and shows neuroprotective effects in cultures²⁹⁰. However, despite encouraging preclinical and phase II clinical trial data, DP-b99 did not show efficacy in ischemic stroke patients²⁹¹.

The newly-developed MMP inhibitor IPR-179 has high bioavailability, moderate water solubility and is BBB permeable²⁹² making it an interesting replacement for earlier mentioned MMP-inhibitors. Its inhibiting capacities for MMP2 and MMP9 are in nanomolar range while IC₅₀ values for other MMPs and non-MMP proteases are >10 μM. Administration of IPR-179 had anticonvulsive effects on acute PTZ-induced seizures in rats²⁹². In this thesis, we administered IPR-179 in two rodent models of epilepsy; the rapid kindling rat model and the intrahippocampal KA mouse model. In the rapid kindling rat model, IPR-179-treated rats showed less severe seizures in response to electrical stimulation compared to vehicle-treated animals. In the absence of the drug, these animals still had less

severe behavioural electrically-induced seizures. In the intrahippocampal KA mouse model, IPR-179-treated animals showed less epileptiform epochs, together with decreased seizure duration and overall decreased amount of seizures compared to vehicle-treated animals, which lasted for 7 weeks after IPR-179 was discontinued. Seizure-induced cognitive deficits such as impaired spatial navigation and novel object recognition, were attenuated with IPR-179 treatment. Together, these data show that IPR-179 decreased MMP9 activity effectively in rodent models and has disease-modifying effects without adverse effects. This renders IPR-179 extremely interesting in the search for novel therapeutics for drug-resistant epilepsy patients. Due to the promising results of MMP inhibition by this drug, the company that developed IPR-179 is currently aiming at improving the oral bioavailability of the compound while preserving the compound's BBB permeability, low cytotoxicity and high stability in order to bring the drug into clinical trials.

Conclusion

Taken together, this thesis has provided evidence that brain inflammation, BBB dysfunction and ECM dysregulation by MMPs play important roles in epileptogenic brain. Reciprocally interacting with each other, a vicious circle is formed which leads to the aggravation of pathology and progression of epilepsy. Therefore, alleviating brain inflammation, BBB dysfunction and ECM dysregulation could be a novel therapeutic strategy to inhibit or block epileptogenesis. We have shown that rapamycin, inflammation-related miRNAs and MMP inhibitor IPR-179 have disease-modifying effects in experimental models of epilepsy. In this respect, inhibiting the (immuno)proteasome, restoring a dysfunctional BBB and inhibiting MMPs are promising new approaches, which should be further investigated. Hopefully, the research presented in this thesis has helped to shorten the path towards an effective treatment for drug-resistant epilepsy patients and will aid in the development of a drug that ideally prevents and cures epilepsy.

References

1. Fiest KM, Sauro KM, Wiebe S, Patten SB, Kwon CS, Dykeman J, et al. Prevalence and incidence of epilepsy: A systematic review and meta-analysis of international studies. *Neurology*. 2017;88(3):296-303.
2. Kwan P, Arzimanoglou A, Berg AT, Brodie MJ, Allen Hauser W, Mathern G, et al. Definition of drug resistant epilepsy: consensus proposal by the ad hoc Task Force of the ILAE Commission on Therapeutic Strategies. *Epilepsia*. 2010;51(6):1069-77.
3. Sisodiya SM, Lin WR, Harding BN, Squier MV, Thom M. Drug resistance in epilepsy: expression of drug resistance proteins in common causes of refractory epilepsy. *Brain*. 2002;125(Pt 1):22-31.
4. Brodie MJ, Barry SJ, Bamagous GA, Norrie JD, Kwan P. Patterns of treatment response in newly diagnosed epilepsy. *Neurology*. 2012;78(20):1548-54.
5. Vezzani A. Inflammation and epilepsy. *Epilepsy Curr*. 2005;5(1):1-6.
6. Vezzani A, Aronica E, Mazarati A, Pittman QJ. Epilepsy and brain inflammation. *Experimental neurology*. 2013;244:11-21.
7. Vezzani A, Fujinami RS, White HS, Preux PM, Blumcke I, Sander JW, et al. Infections, inflammation and epilepsy. *Acta neuropathologica*. 2016;131(2):211-34.
8. Bauer J, Becker AJ, Elyaman W, Peltola J, Ruegg S, Titulaer MJ, et al. Innate and adaptive immunity in human epilepsies. *Epilepsia*. 2017;58 Suppl 3:57-68.
9. Vezzani A, French J, Bartfai T, Baram TZ. The role of inflammation in epilepsy. *Nature reviews Neurology*. 2011;7(1):31-40.
10. van Vliet EA, Aronica E, Vezzani A, Ravizza T. Review: Neuroinflammatory pathways as treatment targets and biomarker candidates in epilepsy: emerging evidence from preclinical and clinical studies. *Neuropathology and applied neurobiology*. 2018;44(1):91-111.
11. van Vliet EA, da Costa Araújo S, Redeker S, Aronica E, Gorter JA. Blood-brain barrier leakage may lead to progression of temporal lobe epilepsy. *Brain*. 2007;130(Pt 2):521-34.
12. Vezzani A, Granata T. Brain inflammation in epilepsy: experimental and clinical evidence. *Epilepsia*. 2005;46(11):1724-43.
13. Crespel A, Coubes P, Rousset MC, Brana C, Rougier A, Rondouin G, et al. Inflammatory reactions in human medial temporal lobe epilepsy with hippocampal sclerosis. *Brain research*. 2002;952(2):159-69.
14. Marchi N, Angelov L, Masaryk T, Fazio V, Granata T, Hernandez N, et al. Seizure-Promoting Effect of Blood-Brain Barrier Disruption. *Epilepsia*. 2007;48(4):732-42.
15. Marchi N, Granata T, Freri E, Cusani E, Ragona F, Puvenna V, et al. Efficacy of anti-inflammatory therapy in a model of acute seizures and in a population of pediatric drug resistant epileptics. *PLoS one*. 2011;6(3):e18200.
16. Marchi N, Johnson AJ, Puvenna V, Johnson HL, Tierney W, Ghosh C, et al. Modulation of peripheral cytotoxic cells and ictogenesis in a model of seizures. *Epilepsia*. 2011;52(9):1627-34.
17. Aronica E, Boer K, van Vliet EA, Redeker S, Baayen JC, Spliet WGM, et al. Complement activation in experimental and human temporal lobe epilepsy. *Neurobiology of disease*. 2007;26(3):497-511.
18. Ravizza T, Gagliardi B, Noe F, Boer K, Aronica E, Vezzani A. Innate and adaptive immunity during epileptogenesis and spontaneous seizures: evidence from experimental models and human temporal lobe epilepsy. *Neurobiology of disease*. 2008;29(1):142-60.
19. van Gassen KL, de Wit M, Koerkamp MJ, Rensen MG, van Rijen PC, Holstege FC, et al. Possible role of the innate immunity in temporal lobe epilepsy. *Epilepsia*. 2008;49(6):1055-65.
20. Maldonado M, Baybis M, Newman D, Kolson DL, Chen W, McKhann G, 2nd, et al. Expression of ICAM-1, TNF-alpha, NF kappa B, and MAP kinase in tubers of the tuberous sclerosis complex. *Neurobiology of disease*. 2003;14(2):279-90.
21. Boer K, Crino PB, Gorter JA, Nellist M, Jansen FE, Spliet WG, et al. Gene expression analysis of tuberous sclerosis complex cortical tubers reveals increased expression of adhesion and inflammatory factors. *Brain Pathol*. 2010;20(4):704-19.
22. Boer K, Jansen F, Nellist M, Redeker S, van den Ouweland AM, Spliet WG, et al. Inflammatory processes in cortical tubers and subependymal giant cell tumors of tuberous sclerosis complex. *Epilepsy research*. 2008;78(1):7-21.
23. Iyer AM, Zurolo E, Boer K, Baayen JC, Giangaspero F, Arcella A, et al. Tissue plasminogen activator and urokinase plasminogen activator in human epileptogenic pathologies. *Neuroscience*. 2010;167(3):929-45.
24. Iyer A, Zurolo E, Spliet WG, van Rijen PC, Baayen JC, Gorter JA, et al. Evaluation of the innate and adaptive immunity in type I and type II focal cortical dysplasias. *Epilepsia*. 2010;51(9):1763-73.
25. Ravizza T, Boer K, Redeker S, Spliet WG, van Rijen PC, Troost D, et al. The IL-1beta system in epilepsy-associated malformations of cortical development. *Neurobiology of disease*. 2006;24(1):128-43.
26. Maroso M, Balosso S, Ravizza T, Iori V, Wright CI, French J, et al. Interleukin-1beta biosynthesis inhibition reduces acute seizures and drug resistant chronic epileptic activity in mice. *Neurotherapeutics : the journal of the American Society for Experimental NeuroTherapeutics*. 2011;8(2):304-15.
27. Maroso M, Balosso S, Ravizza T, Liu J, Aronica E, Iyer AM, et al. Toll-like receptor 4 and high-mobility group box-

- 1 are involved in ictogenesis and can be targeted to reduce seizures. *Nat Med.* 2010;16(4):413-9.
28. Aronica E, Bauer S, Bozzi Y, Caleo M, Dingledine R, Gorter JA, et al. Neuroinflammatory targets and treatments for epilepsy validated in experimental models. *Epilepsia.* 2017;58 Suppl 3:27-38.
29. Aronica E, Ravizza T, Zurolo E, Vezzani A. Astrocyte immune responses in epilepsy. *Glia.* 2012;60(8):1258-68.
30. Muhlechner A, van Scheppingen J, Hulshof HM, Scholl T, Iyer AM, Anink JJ, et al. Novel Histopathological Patterns in Cortical Tubers of Epilepsy Surgery Patients with Tuberous Sclerosis Complex. *PLoS one.* 2016;11(6):e0157396.
31. Fabene PF, Navarro Mora G, Martinello M, Rossi B, Merigo F, Ottoboni L, et al. A role for leukocyte-endothelial adhesion mechanisms in epilepsy. *Nat Med.* 2008;14(12):1377-83.
32. Zattoni M, Mura ML, Deprez F, Schwendener RA, Engelhardt B, Frei K, et al. Brain infiltration of leukocytes contributes to the pathophysiology of temporal lobe epilepsy. *The Journal of neuroscience : the official journal of the Society for Neuroscience.* 2011;31(11):4037-50.
33. Rana A, Musto AE. The role of inflammation in the development of epilepsy. *Journal of neuroinflammation.* 2018;15(1):144.
34. Speese SD, Trotta N, Rodesch CK, Aravamudan B, Broadie K. The ubiquitin proteasome system acutely regulates presynaptic protein turnover and synaptic efficacy. *Current biology : CB.* 2003;13(11):899-910.
35. Hegde AN. The ubiquitin-proteasome pathway and synaptic plasticity. *Learning & memory (Cold Spring Harbor, NY).* 2010;17(7):314-27.
36. Grimm S, Ott C, Horlacher M, Weber D, Hohn A, Grune T. Advanced-glycation-end-product-induced formation of immunoproteasomes: involvement of RAGE and Jak2/STAT1. *The Biochemical journal.* 2012;448(1):127-39.
37. Pla A, Pascual M, Renau-Piqueras J, Guerri C. TLR4 mediates the impairment of ubiquitin-proteasome and autophagy-lysosome pathways induced by ethanol treatment in brain. *Cell death & disease.* 2014;5:e1066.
38. Mishto M, Raza ML, de Biase D, Ravizza T, Vasuri F, Martucci M, et al. The immunoproteasome beta5i subunit is a key contributor to ictogenesis in a rat model of chronic epilepsy. *Brain, behavior, and immunity.* 2015;49:188-96.
39. Kotamraju S, Matalon S, Matsunaga T, Shang T, Hickman-Davis JM, Kalyanaraman B. Upregulation of immunoproteasomes by nitric oxide: potential antioxidative mechanism in endothelial cells. *Free radical biology & medicine.* 2006;40(6):1034-44.
40. Yun YS, Kim KH, Tschida B, Sachs Z, Noble-Orcutt KE, Moriarity BS, et al. mTORC1 Coordinates Protein Synthesis and Immunoproteasome Formation via PRAS40 to Prevent Accumulation of Protein Stress. *Molecular cell.* 2016;61(4):625-39.
41. Limanaqi F, Biagioni F, Gaglione A, Busceti CL, Fornai F. A Sentinel in the Crosstalk Between the Nervous and Immune System: The (Immuno)-Proteasome. *Frontiers in immunology.* 2019;10:628.
42. Schmidt M, Finley D. Regulation of proteasome activity in health and disease. *Biochimica et biophysica acta.* 2014;1843(1):13-25.
43. Klare N, Seeger M, Janek K, Jungblut PR, Dahlmann B. Intermediate-type 20 S proteasomes in HeLa cells: "asymmetric" subunit composition, diversity and adaptation. *J Mol Biol.* 2007;373(1):1-10.
44. Aki M, Shimbara N, Takashina M, Akiyama K, Kagawa S, Tamura T, et al. Interferon-gamma induces different subunit organizations and functional diversity of proteasomes. *J Biochem.* 1994;115(2):257-69.
45. Cajigas LJ, Will T, Schuman EM. Protein homeostasis and synaptic plasticity. *The EMBO journal.* 2010;29(16):2746-52.
46. Colledge M, Snyder EM, Crozier RA, Soderling JA, Jin Y, Langeberg LK, et al. Ubiquitination regulates PSD-95 degradation and AMPA receptor surface expression. *Neuron.* 2003;40(3):595-607.
47. Arancibia-Carcamo IL, Kittler JT. Regulation of GABA(A) receptor membrane trafficking and synaptic localization. *Pharmacology & therapeutics.* 2009;123(1):17-31.
48. Hamilton AM, Oh WC, Vega-Ramirez H, Stein IS, Hell JW, Patrick GN, et al. Activity-dependent growth of new dendritic spines is regulated by the proteasome. *Neuron.* 2012;74(6):1023-30.
49. Groettrup M, Kirk CJ, Basler M. Proteasomes in immune cells: more than peptide producers? *Nat Rev Immunol.* 2010;10(1):73-8.
50. McCarthy MK, Weinberg JB. The immunoproteasome and viral infection: a complex regulator of inflammation. *Frontiers in microbiology.* 2015;6:21.
51. Ebstein F, Kloetzel PM, Kruger E, Seifert U. Emerging roles of immunoproteasomes beyond MHC class I antigen processing. *Cellular and molecular life sciences : CMLS.* 2012;69(15):2543-58.
52. Basler M, Kirk CJ, Groettrup M. The immunoproteasome in antigen processing and other immunological functions. *Current opinion in immunology.* 2013;25(1):74-80.
53. Wamatsch A, Bergann T, Kruger E. Oxidation matters: the ubiquitin proteasome system connects innate immune mechanisms with MHC class I antigen presentation. *Molecular immunology.* 2013;55(2):106-9.
54. Maldonado M, Kappahh RJ, Terluk MR, Heuss ND, Yuan C, Gregerson DS, et al. Immunoproteasome deficiency modifies the alternative pathway of NFkappaB signaling. *PLoS one.* 2013;8(2):e56187.
55. Kloetzel PM. Antigen processing by the proteasome.

- Nature reviews Molecular cell biology. 2001;2(3):179-87.
56. Giannoni P, Badaut J, Dargazanli C, De Maudave AF, Klement W, Costalat V, et al. The pericyte-glia interface at the blood-brain barrier. *Clin Sci (Lond)*. 2018;132(3):361-74.
 57. Klement W, Garbelli R, Zub E, Rossini L, Tassi L, Girard B, et al. Seizure progression and inflammatory mediators promote pericytosis and pericyte-microglia clustering at the cerebrovasculature. *Neurobiology of disease*. 2018;113:70-81.
 58. Kim SY, Senatorov VV, Jr., Morrissey CS, Lippmann K, Vazquez O, Milikovsky DZ, et al. TGFbeta signaling is associated with changes in inflammatory gene expression and perineuronal net degradation around inhibitory neurons following various neurological insults. *Sci Rep*. 2017;7(1):7711.
 59. Zhu Y, Culmsee C, Klumpp S, Kriegstein J. Neuroprotection by transforming growth factor-beta1 involves activation of nuclear factor-kappaB through phosphatidylinositol-3-OH kinase/Akt and mitogen-activated protein kinase-extracellular-signal regulated kinase1,2 signaling pathways. *Neuroscience*. 2004;123(4):897-906.
 60. Davies M, Robinson M, Smith E, Huntley S, Prime S, Paterson I. Induction of an epithelial to mesenchymal transition in human immortal and malignant keratinocytes by TGF-beta1 involves MAPK, Smad and AP-1 signalling pathways. *Journal of cellular biochemistry*. 2005;95(5):918-31.
 61. Cacheaux LP, Ivens S, David Y, Lakhter AJ, Bar-Klein G, Shapira M, et al. Transcriptome profiling reveals TGF-beta signaling involvement in epileptogenesis. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2009;29(28):8927-35.
 62. Ralay Ranaivo H, Patel F, Wainwright MS. Albumin activates the canonical TGF receptor-smad signaling pathway but this is not required for activation of astrocytes. *Experimental neurology*. 2010;226(2):310-9.
 63. Ralay Ranaivo H, Wainwright MS. Albumin activates astrocytes and microglia through mitogen-activated protein kinase pathways. *Brain research*. 2010;1313:222-31.
 64. Deng YY, Lu J, Ling EA, Kaur C. Monocyte chemoattractant protein-1 (MCP-1) produced via NF-kappaB signaling pathway mediates migration of amoeboid microglia in the periventricular white matter in hypoxic neonatal rats. *Glia*. 2009;57(6):604-21.
 65. Sheehan JJ, Zhou C, Gravanis I, Rogove AD, Wu YP, Bogenhagen DF, et al. Proteolytic activation of monocyte chemoattractant protein-1 by plasmin underlies excitotoxic neurodegeneration in mice. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2007;27(7):1738-45.
 66. Dinkel K, MacPherson A, Sapolsky RM. Novel glucocorticoid effects on acute inflammation in the CNS. *J Neurochem*. 2003;84(4):705-16.
 67. Jiang Y, Beller DI, Frenzl G, Graves DT. Monocyte chemoattractant protein-1 regulates adhesion molecule expression and cytokine production in human monocytes. *Journal of immunology (Baltimore, Md : 1950)*. 1992;148(8):2423-8.
 68. Yadav A, Saini V, Arora S. MCP-1: chemoattractant with a role beyond immunity: a review. *Clin Chim Acta*. 2010;411(21-22):1570-9.
 69. Varvel NH, Neher JJ, Bosch A, Wang W, Ransohoff RM, Miller RJ, et al. Infiltrating monocytes promote brain inflammation and exacerbate neuronal damage after status epilepticus. *Proceedings of the National Academy of Sciences of the United States of America*. 2016;113(38):E5665-74.
 70. Bozzi Y, Caleo M. Epilepsy, Seizures, and Inflammation: Role of the C-C Motif Ligand 2 Chemokine. *DNA Cell Biol*. 2016;35(6):257-60.
 71. David Y, Cacheaux LP, Ivens S, Lapilover E, Heinemann U, Kaufer D, et al. Astrocytic dysfunction in epileptogenesis: consequence of altered potassium and glutamate homeostasis? *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2009;29(34):10588-99.
 72. Coulter DA, Steinhauser C. Role of astrocytes in epilepsy. *Cold Spring Harbor perspectives in medicine*. 2015;5(3):a022434.
 73. Fabene PF, Bramanti P, Constantini G. The emerging role for chemokines in epilepsy. *Journal of neuroimmunology*. 2010;224(1-2):22-7.
 74. Jabs R, Seifert G, Steinhauser C. Astrocytic function and its alteration in the epileptic brain. *Epilepsia*. 2008;49 Suppl 2:3-12.
 75. Marchi N, Tierney W, Alexopoulos AV, Puvion V, Granata T, Janigro D. The etiological role of blood-brain barrier dysfunction in seizure disorders. *Cardiovasc Psychiatry Neurol*. 2011;2011:482415.
 76. van Vliet EA, Otte WM, Wadman WJ, Aronica E, Kooij G, de Vries HE, et al. Blood-brain barrier leakage after status epilepticus in rapamycin-treated rats I: Magnetic resonance imaging. *Epilepsia*. 2016;57(1):59-69.
 77. van Vliet EA, Otte WM, Wadman WJ, Aronica E, Kooij G, de Vries HE, et al. Blood-brain barrier leakage after status epilepticus in rapamycin-treated rats II: Potential mechanisms. *Epilepsia*. 2016;57(1):70-8.
 78. van Vliet EA, Aronica E, Gorter JA. Blood-brain barrier dysfunction, seizures and epilepsy. *Seminars in cell & developmental biology*. 2015;38:26-34.
 79. Abbott NJ. Inflammatory mediators and modulation of blood-brain barrier permeability. *Cellular and molecular neurobiology*. 2000;20(2):131-47.
 80. Abbott NJ, Friedman A. Overview and introduction: the blood-brain barrier in health and disease. *Epilepsia*. 2012;53 Suppl 6:1-6.
 81. Pitkänen A, Immonen RJ, Grohn OH, Kharatishvili I. From

- traumatic brain injury to posttraumatic epilepsy: what animal models tell us about the process and treatment options. *Epilepsia*. 2009;50 Suppl 2:21-9.
82. Pitkänen A, Roivainen R, Lukasiuk K. Development of epilepsy after ischaemic stroke. *The Lancet Neurology*. 2015.
83. Rosell A, Ortega-Aznar A, Alvarez-Sabin J, Fernandez-Cadenas I, Ribo M, Molina CA, et al. Increased brain expression of matrix metalloproteinase-9 after ischemic and hemorrhagic human stroke. *Stroke*. 2006;37(6):1399-406.
84. Mihaly A, Bozoky B. Immunohistochemical localization of extravasated serum albumin in the hippocampus of human subjects with partial and generalized epilepsies and epileptiform convulsions. *Acta Neuropathol (Berl)*. 1984;65(1):25-34.
85. Rigau V, Morin M, Rousset MC, de Bock F, Lebrun A, Coubes P, et al. Angiogenesis is associated with blood-brain barrier permeability in temporal lobe epilepsy. *Brain*. 2007;130(Pt 7):1942-56.
86. Kastanauskaitė A, Alonso-Nanclares L, Blazquez-Llorca L, Pastor J, Sola RG, DeFelipe J. Alterations of the microvascular network in sclerotic hippocampi from patients with epilepsy. *J Neuropathol Exp Neurol*. 2009;68(8):939-50.
87. Marchi N, Teng Q, Ghosh C, Fan Q, Nguyen MT, Desai NK, et al. Blood-brain barrier damage, but not parenchymal white blood cells, is a hallmark of seizure activity. *Brain research*. 2010;1353:176-86.
88. Michalak Z, Lebrun A, Di Miceli M, Rousset MC, Crespel A, Coubes P, et al. IgG leakage may contribute to neuronal dysfunction in drug-refractory epilepsies with blood-brain barrier disruption. *Journal of neuropathology and experimental neurology*. 2012;71(9):826-38.
89. Raabe A, Schmitz AK, Pernhorst K, Grote A, von der Brélie C, Urbach H, et al. Cliniconeuropathologic correlations show astroglial albumin storage as a common factor in epileptogenic vascular lesions. *Epilepsia*. 2012;53(3):539-48.
90. Liu JY, Thom M, Catarino CB, Martinian L, Figarella-Branger D, Bartolomei F, et al. Neuropathology of the blood-brain barrier and pharmaco-resistance in human epilepsy. *Brain*. 2012;135(Pt 10):3115-33.
91. Schmitz AK, Grote A, Raabe A, Urbach H, Friedman A, von Lehe M, et al. Albumin storage in neoplastic astroglial elements of gangliogliomas. *Seizure*. 2013;22(2):144-50.
92. Dityatev A. Remodeling of extracellular matrix and epileptogenesis. *Epilepsia*. 2010;51 Suppl 3:61-5.
93. Mills JD, Iyer AM, van Scheppingen J, Bongaarts A, Anink JJ, Janssen B, et al. Coding and small non-coding transcriptional landscape of tuberous sclerosis complex cortical tubers: implications for pathophysiology and treatment. *Sci Rep*. 2017;7(1):8089.
94. Gorter JA, Van Vliet EA, Rauwerda H, Breit T, Stad R, van Schaik L, et al. Dynamic changes of proteases and protease inhibitors revealed by microarray analysis in CA3 and entorhinal cortex during epileptogenesis in the rat. *Epilepsia*. 2007;48 Suppl 5:53-64.
95. Nagy V, Bozdagi O, Matyenia A, Balcerzyk M, Okulski P, Dzwonek J, et al. Matrix metalloproteinase-9 is required for hippocampal late-phase long-term potentiation and memory. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2006;26(7):1923-34.
96. Ethell IM, Ethell DW. Matrix metalloproteinases in brain development and remodeling: synaptic functions and targets. *Journal of neuroscience research*. 2007;85(13):2813-23.
97. Bajor M, Michaluk P, Gulyassy P, Kekesi AK, Juhasz G, Kaczmarek L. Synaptic cell adhesion molecule-2 and collapsin response mediator protein-2 are novel members of the matrix metalloproteinase-9 degradome. *J Neurochem*. 2012;122(4):775-88.
98. Jovanov Milosevic N, Judas M, Aronica E, Kostovic I. Neural ECM in laminar organization and connectivity development in healthy and diseased human brain. *Progress in brain research*. 2014;214:159-78.
99. Giannelli G, Falk-Marzillier J, Schiraldi O, Stetler-Stevenson WG, Quaranta V. Induction of cell migration by matrix metalloproteinase-2 cleavage of laminin-5. *Science (New York, NY)*. 1997;277(5323):225-8.
100. Kelly MA, Shuaib A, Todd KG. Matrix metalloproteinase activation and blood-brain barrier breakdown following thrombolysis. *Experimental neurology*. 2006;200(1):38-49.
101. Kim EM, Hwang O. Role of matrix metalloproteinase-3 in neurodegeneration. *J Neurochem*. 2011;116(1):22-32.
102. Gidday JM, Gasche YG, Copin JC, Shah AR, Perez RS, Shapiro SD, et al. Leukocyte-derived matrix metalloproteinase-9 mediates blood-brain barrier breakdown and is proinflammatory after transient focal cerebral ischemia. *Am J Physiol Heart Circ Physiol*. 2005;289(2):H558-68.
103. Justicia C, Panes J, Sole S, Cervera A, Deulofeu R, Chamorro A, et al. Neutrophil infiltration increases matrix metalloproteinase-9 in the ischemic brain after occlusion/reperfusion of the middle cerebral artery in rats. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*. 2003;23(12):1430-40.
104. Konopka A, Grajkowska W, Ziemianska K, Roszkowski M, Daszkiewicz P, Rysz A, et al. Matrix metalloproteinase-9 (MMP-9) in human intractable epilepsy caused by focal cortical dysplasia. *Epilepsy research*. 2013;104(1-2):45-58.
105. Broekaert DWM, van Scheppingen J, Anink JJ, Wierts L, van Het Hof B, Jansen FE, et al. Increased matrix metalloproteinases expression in tuberous sclerosis complex: modulation by microRNA 146a and 147b in vitro.

- Neuropathology and applied neurobiology. 2019.
106. Motti D, Le Duigou C, Eugene E, Chemaly N, Wittner L, Lazarevic D, et al. Gene expression analysis of the emergence of epileptiform activity after focal injection of kainic acid into mouse hippocampus. *Eur J Neurosci*. 2010;32(8):1364-79.
 107. Itoh Y, Takamura A, Ito N, Maru Y, Sato H, Suenaga N, et al. Homophilic complex formation of MT1-MMP facilitates proMMP-2 activation on the cell surface and promotes tumor cell invasion. *The EMBO journal*. 2001;20(17):4782-93.
 108. Atkinson SJ, Crabbe T, Cowell S, Ward RV, Butler MJ, Sato H, et al. Intermolecular autolytic cleavage can contribute to the activation of progelatinase A by cell membranes. *The Journal of biological chemistry*. 1995;270(51):30479-85.
 109. Nagase H. Tailoring TIMPs for Selective Metalloproteinase Inhibition. In: Edwards D, editor. *The Cancer Degradome*. Springer; 2008. p. 787-810.
 110. Blumcke I, Thom M, Aronica E, Armstrong DD, Bartolomei F, Bernasconi A, et al. International consensus classification of hippocampal sclerosis in temporal lobe epilepsy: a Task Force report from the ILAE Commission on Diagnostic Methods. *Epilepsia*. 2013;54(7):1315-29.
 111. Kim EM, Hwang O. Role of matrix metalloproteinase-3 in neurodegeneration. *J Neurochem*. 2011;116(1):22-32.
 112. Witek-Zawada B, Koj A. Regulation of expression of stromelysin-1 by proinflammatory cytokines in mouse brain astrocytes. *Journal of physiology and pharmacology : an official journal of the Polish Physiological Society*. 2003;54(4):489-96.
 113. Delany AM, Brinckerhoff CE. Post-transcriptional regulation of collagenase and stromelysin gene expression by epidermal growth factor and dexamethasone in cultured human fibroblasts. *Journal of cellular biochemistry*. 1992;50(4):400-10.
 114. Kirstein M, Sanz L, Quinones S, Moscat J, Diaz-Meco MT, Saus J. Cross-talk between different enhancer elements during mitogenic induction of the human stromelysin-1 gene. *The Journal of biological chemistry*. 1996;271(30):18231-6.
 115. Lachos J, Zattoni M, Wieser HG, Fritschy JM, Langmann T, Schmitz G, et al. Characterization of the gene expression profile of human hippocampus in mesial temporal lobe epilepsy with hippocampal sclerosis. *Epilepsy research and treatment*. 2011;2011:758407.
 116. De Simoni MG, Perego C, Ravizza T, Moneta D, Conti M, Marchesi F, et al. Inflammatory cytokines and related genes are induced in the rat hippocampus by limbic status epilepticus. *Eur J Neurosci*. 2000;12(7):2623-33.
 117. Gorter JA, Van Vliet EA, Aronica E, Breit T, Rauwerda H, Lopes da Silva FH, et al. Potential new anti-epileptogenic targets indicated by microarray analysis in a rat model for temporal lobe epilepsy. *Journal of Neuroscience*. 2006;26(43):11083-110.
 118. Dhote F, Peinnequin A, Carpentier P, Baille V, Delacour C, Foquin A, et al. Prolonged inflammatory gene response following soman-induced seizures in mice. *Toxicology*. 2007;238(2-3):166-76.
 119. Dube CM, Ravizza T, Hamamura M, Zha Q, Keebaugh A, Fok K, et al. Epileptogenesis provoked by prolonged experimental febrile seizures: mechanisms and biomarkers. *The Journal of neuroscience: the official journal of the Society for Neuroscience*. 2010;30(22):7484-94.
 120. Voutsinos-Porche B, Koning E, Kaplan H, Ferrandon A, Guenounou M, Nehlig A, et al. Temporal patterns of the cerebral inflammatory response in the rat lithium-pilocarpine model of temporal lobe epilepsy. *Neurobiology of disease*. 2004;17(3):385-402.
 121. Arisi GM, Foresti ML, Katki K, Shapiro LA. Increased CCL2, CCL3, CCL5, and IL-1beta cytokine concentration in piriform cortex, hippocampus, and neocortex after pilocarpine-induced seizures. *Journal of neuroinflammation*. 2015;12:129.
 122. Ameyar M, Wisniewska M, Weitzman JB. A role for AP-1 in apoptosis: the case for and against. *Biochimie*. 2003;85(8):747-52.
 123. Barkett M, Gilmore TD. Control of apoptosis by Rel/NF-kappaB transcription factors. *Oncogene*. 1999;18(49):6910-24.
 124. Bhaumik D, Scott GK, Schokrpur S, Patil CK, Campisi J, Benz CC. Expression of microRNA-146 suppresses NF-kappaB activity with reduction of metastatic potential in breast cancer cells. *Oncogene*. 2008;27(42):5643-7.
 125. Yan C, Boyd DD. Regulation of matrix metalloproteinase gene expression. *Journal of cellular physiology*. 2007;211(1):19-26.
 126. Benbow U, Brinckerhoff CE. The AP-1 site and MMP gene regulation: what is all the fuss about? *Matrix biology : journal of the International Society for Matrix Biology*. 1997;15(8-9):519-26.
 127. Van Hove I, Lemmens K, Van de Velde S, Verslegers M, Moons L. Matrix metalloproteinase-3 in the central nervous system: a look on the bright side. *J Neurochem*. 2012;123(2):203-16.
 128. Crocker SJ, Milner R, Pham-Mitchell N, Campbell IL. Cell and agonist-specific regulation of genes for matrix metalloproteinases and their tissue inhibitors by primary glial cells. *J Neurochem*. 2006;98(3):812-23.
 129. Vincenti MP. The matrix metalloproteinase (MMP) and tissue inhibitor of metalloproteinase (TIMP) genes. Transcriptional and posttranscriptional regulation, signal transduction and cell-type-specific expression. *Methods in molecular biology (Clifton, NJ)*. 2001;151:121-48.

130. Moon SK, Cha BY, Kim CH. ERK1/2 mediates TNF- α -induced matrix metalloproteinase-9 expression in human vascular smooth muscle cells via the regulation of NF- κ B and AP-1: Involvement of the ras dependent pathway. *Journal of cellular physiology*. 2004;198(3):417-27.
131. Van den Steen PE, Proost P, Wuyts A, Van Damme J, Opdenakker G. Neutrophil gelatinase B potentiates interleukin-8 tenfold by aminoterminal processing, whereas it degrades CTAP-III, PF-4, and GRO- α and leaves RANTES and MCP-2 intact. *Blood*. 2000;96(8):2673-81.
132. Schonbeck U, Mach F, Libby P. Generation of biologically active IL-1 β by matrix metalloproteinases: a novel caspase-1-independent pathway of IL-1 β processing. *Journal of immunology (Baltimore, Md : 1950)*. 1998;161(7):3340-6.
133. Mohan MJ, Seaton T, Mitchell J, Howe A, Blackburn K, Burkhart W, et al. The tumor necrosis factor- α converting enzyme (TACE): a unique metalloproteinase with highly defined substrate selectivity. *Biochemistry*. 2002;41(30):9462-9.
134. Vezzani A, Friedman A, Dingledine RJ. The role of inflammation in epileptogenesis. *Neuropharmacology*. 2013;69:16-24.
135. Xanthos DN, Sandkuhler J. Neurogenic neuroinflammation: inflammatory CNS reactions in response to neuronal activity. *Nat Rev Neurosci*. 2014;15(1):43-53.
136. Iori V, Frigerio F, Vezzani A. Modulation of neuronal excitability by immune mediators in epilepsy. *Current opinion in pharmacology*. 2016;26:118-23.
137. Vezzani A, Moneta D, Conti M, Richichi C, Ravizza T, De Luigi A, et al. Powerful anticonvulsant action of IL-1 receptor antagonist on intracerebral injection and astrocytic overexpression in mice. *Proceedings of the National Academy of Sciences of the United States of America*. 2000;97(21):11534-9.
138. Vezzani A, Conti M, De Luigi A, Ravizza T, Moneta D, Marchesi F, et al. Interleukin-1 β immunoreactivity and microglia are enhanced in the rat hippocampus by focal kainate application: functional evidence for enhancement of electrographic seizures. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 1999;19(12):5054-65.
139. Turrin NP, Rivest S. Innate immune reaction in response to seizures: implications for the neuropathology associated with epilepsy. *Neurobiology of disease*. 2004;16(2):321-34.
140. Jung KH, Chu K, Lee ST, Kim J, Sinn DJ, Kim JM, et al. Cyclooxygenase-2 inhibitor, celecoxib, inhibits the altered hippocampal neurogenesis with attenuation of spontaneous recurrent seizures following pilocarpine-induced status epilepticus. *Neurobiology of disease*. 2006;23(2):237-46.
141. Holtman L, van Vliet EA, van Schaik R, Queiroz CM, Aronica E, Gorter JA. Effects of SC58236, a selective COX-2 inhibitor, on epileptogenesis and spontaneous seizures in a rat model for temporal lobe epilepsy. *Epilepsy research*. 2009;84(1):56-66.
142. Chen J, Marsh T, Zhang JS, Graham SH. Expression of cyclo-oxygenase 2 in rat brain following kainate treatment. *Neuroreport*. 1995;6(2):245-8.
143. Pernot F, Heinrich C, Barbier L, Peinnequin A, Carpentier P, Dhote F, et al. Inflammatory changes during epileptogenesis and spontaneous seizures in a mouse model of mesiotemporal lobe epilepsy. *Epilepsia*. 2011;52(12):2315-25.
144. Jiang J, Yang MS, Quan Y, Gueorguieva P, Ganesh T, Dingledine R. Therapeutic window for cyclooxygenase-2 related anti-inflammatory therapy after status epilepticus. *Neurobiology of disease*. 2015;76:126-36.
145. Lehtimäki KA, Peltola J, Koskikallio E, Keränen T, Honkaniemi J. Expression of cytokines and cytokine receptors in the rat brain after kainic acid-induced seizures. *Brain research Molecular brain research*. 2003;110(2):253-60.
146. Patel N, Ram D, Swiderska N, Mewasingh LD, Newton RW, Offringa M. Febrile seizures. *BMJ (Clinical research ed)*. 2015;351:h4240.
147. Dube CM, Brewster AL, Baram TZ. Febrile seizures: mechanisms and relationship to epilepsy. *Brain & development*. 2009;31(5):366-71.
148. Bien CG, Urbach H, Schramm J, Soeder BM, Becker AJ, Voltz R, et al. Limbic encephalitis as a precipitating event in adult-onset temporal lobe epilepsy. *Neurology*. 2007;69(12):1236-44.
149. Vincent A, Bien CG. Anti-NMDA-receptor encephalitis: a cause of psychiatric, seizure, and movement disorders in young adults. *The Lancet Neurology*. 2008;7(12):1074-5.
150. Najjar S, Bernbaum M, Lai G, Devinsky O. Immunology and epilepsy. *Reviews in neurological diseases*. 2008;5(3):109-16.
151. Auvin S, Shin D, Mazarati A, Sankar R. Inflammation induced by LPS enhances epileptogenesis in immature rat and may be partially reversed by IL1RA. *Epilepsia*. 2010;51 Suppl 3:34-8.
152. Balosso S, Ravizza T, Aronica E, Vezzani A. The dual role of TNF- α and its receptors in seizures. *Experimental neurology*. 2013;247:267-71.
153. Balosso S, Ravizza T, Perego C, Peschon J, Campbell IL, De Simoni MG, et al. Tumor necrosis factor- α inhibits seizures in mice via p75 receptors. *Annals of neurology*. 2005;57(6):804-12.
154. Sayyah M, Beheshti S, Shokrgozar MA, Eslami-far A, Deljoo Z, Khabiri AR, et al. Antiepileptogenic and anticonvulsant activity of interleukin-1 β in amygdala-

- kindled rats. *Experimental neurology*. 2005;191(1):145-53.
155. Riazi K, Galic MA, Pittman QJ. Contributions of peripheral inflammation to seizure susceptibility: cytokines and brain excitability. *Epilepsy research*. 2010;89(1):34-42.
156. Shandra AA, Godlevsky LS, Vastyanov RS, Oleinik AA, Konovalenko VL, Rapoport EN, et al. The role of TNF-alpha in amygdala kindled rats. *Neuroscience research*. 2002;42(2):147-53.
157. Mittelman A, Puccio C, Gafney E, Coombe N, Singh B, Wood D, et al. A phase I pharmacokinetic study of recombinant human tumor necrosis factor administered by a 5-day continuous infusion. *Investigational new drugs*. 1992;10(3):183-90.
158. Bedner P, Dupper A, Huttman K, Muller J, Herde MK, Dublin P, et al. Astrocyte uncoupling as a cause of human temporal lobe epilepsy. *Brain*. 2015;138(Pt 5):1208-22.
159. Lagarde S, Villeneuve N, Trebuchon A, Kaphan E, Lepine A, McGonigal A, et al. Anti-tumor necrosis factor alpha therapy (adalimumab) in Rasmussen's encephalitis: An open pilot study. *Epilepsia*. 2016;57(6):956-66.
160. Devinsky O, Vezzani A, Najjar S, De Lanerolle NC, Rogawski MA. Glia and epilepsy: excitability and inflammation. *Trends Neurosci*. 2013;36(3):174-84.
161. Wang S, Cheng Q, Malik S, Yang J. Interleukin-1beta inhibits gamma-aminobutyric acid type A (GABA(A)) receptor current in cultured hippocampal neurons. *The Journal of pharmacology and experimental therapeutics*. 2000;292(2):497-504.
162. Roseti C, van Vliet EA, Cifelli P, Ruffolo G, Baayen JC, Di Castro MA, et al. GABAA currents are decreased by IL-1beta in epileptogenic tissue of patients with temporal lobe epilepsy: implications for ictogenesis. *Neurobiology of disease*. 2015;82:311-20.
163. Takeuchi H, Jin S, Wang J, Zhang G, Kawanokuchi J, Kuno R, et al. Tumor necrosis factor-alpha induces neurotoxicity via glutamate release from hemichannels of activated microglia in an autocrine manner. *The Journal of biological chemistry*. 2006;281(30):21362-8.
164. Bezzi P, Domercq M, Brambilla L, Galli R, Schols D, De Clercq E, et al. CXCR4-activated astrocyte glutamate release via TNFalpha: amplification by microglia triggers neurotoxicity. *Nat Neurosci*. 2001;4(7):702-10.
165. Stellwagen D, Beattie EC, Seo JY, Malenka RC. Differential regulation of AMPA receptor and GABA receptor trafficking by tumor necrosis factor-alpha. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2005;25(12):3219-28.
166. Wheeler D, Knapp E, Bandaru VV, Wang Y, Knorr D, Poirier C, et al. Tumor necrosis factor-alpha-induced neutral sphingomyelinase-2 modulates synaptic plasticity by controlling the membrane insertion of NMDA receptors. *J Neurochem*. 2009;109(5):1237-49.
167. Friedman A. Blood-brain barrier dysfunction, status epilepticus, seizures, and epilepsy: a puzzle of a chicken and egg? *Epilepsia*. 2011;52 Suppl 8:19-20.
168. Swissa E, Serlin Y, Vazana U, Prager O, Friedman A. Blood-brain barrier dysfunction in status epilepticus: Mechanisms and role in epileptogenesis. *Epilepsy Behav*. 2019;101(Pt B):106285.
169. Tomkins O, Feintuch A, Benifla M, Cohen A, Friedman A, Shelef I. Blood-brain barrier breakdown following traumatic brain injury: a possible role in posttraumatic epilepsy. *Cardiovasc Psychiatry Neurol*. 2011;2011:765923.
170. Tomkins O, Shelef I, Kaizerman I, Eliushin A, Afawi Z, Misk A, et al. Blood-brain barrier disruption in post-traumatic epilepsy. *Journal of neurology, neurosurgery, and psychiatry*. 2008;79(7):774-7.
171. Dadas A, Janigro D. Breakdown of blood brain barrier as a mechanism of post-traumatic epilepsy. *Neurobiology of disease*. 2019;123:20-6.
172. Sulhan S, Lyon KA, Shapiro LA, Huang JH. Neuroinflammation and blood-brain barrier disruption following traumatic brain injury: Pathophysiology and potential therapeutic targets. *Journal of neuroscience research*. 2020;98(1):19-28.
173. Lapolover EG, Lippmann K, Salar S, Maslarova A, Dreier JP, Heinemann U, et al. Peri-infarct blood-brain barrier dysfunction facilitates induction of spreading depolarization associated with epileptiform discharges. *Neurobiology of disease*. 2012;48(3):495-506.
174. Dreier JP. The role of spreading depression, spreading depolarization and spreading ischemia in neurological disease. *Nat Med*. 2011;17(4):439-47.
175. Schoknecht K, David Y, Heinemann U. The blood-brain barrier-gatekeeper to neuronal homeostasis: clinical implications in the setting of stroke. *Seminars in cell & developmental biology*. 2015;38:35-42.
176. Nodde-Ekane XE, Hayward N, Grohn O, Pitkanen A. Vascular changes in epilepsy: functional consequences and association with network plasticity in pilocarpine-induced experimental epilepsy. *Neuroscience*. 2010;166(1):312-32.
177. Pont F, Collet A, Lallement G. Early and transient increase of rat hippocampal blood-brain barrier permeability to amino acids during kainic acid-induced seizures. *Neurosci Lett*. 1995;184(1):52-4.
178. Lassmann H, Petsche U, Kitz K, Baran H, Sperk G, Seitelberger F, et al. The role of brain edema in epileptic brain damage induced by systemic kainic acid injection. *Neuroscience*. 1984;13(3):691-704.
179. Roch C, Leroy C, Nehlig A, Namer JJ. Magnetic

- resonance imaging in the study of the lithium-pilocarpine model of temporal lobe epilepsy in adult rats. *Epilepsia*. 2002;43(4):325-35.
180. Librizzi L, Noe F, Vezzani A, de Curtis M, Ravizza T. Seizure-induced brain-borne inflammation sustains seizure recurrence and blood-brain barrier damage. *Annals of neurology*. 2012;72(1):82-90.
181. Prager O, Kamintsky L, Hasam-Henderson LA, Schoknecht K, Wuntke V, Papageorgiou I, et al. Seizure-induced microvascular injury is associated with impaired neurovascular coupling and blood-brain barrier dysfunction. *Epilepsia*. 2019;60(2):322-36.
182. Loscher W, Friedman A. Structural, Molecular, and Functional Alterations of the Blood-Brain Barrier during Epileptogenesis and Epilepsy: A Cause, Consequence, or Both? *International journal of molecular sciences*. 2020;21(2).
183. Korn A, Golan H, Melamed I, Pascual-Marqui R, Friedman A. Focal cortical dysfunction and blood-brain barrier disruption in patients with Postconcussion syndrome. *J Clin Neurophysiol*. 2005;22(1):1-9.
184. Milikovsky DZ, Ofer J, Senatorov VV, Jr, Friedman AR, Prager O, Sheintuch L, et al. Paroxysmal slow cortical activity in Alzheimer's disease and epilepsy is associated with blood-brain barrier dysfunction. *Sci Transl Med*. 2019;11(521).
185. Cornford EM, Hyman S, Cornford ME, Landaw EM, Delgado-Escueta AV. Interictal seizure resections show two configurations of endothelial Glut1 glucose transporter in the human blood-brain barrier. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*. 1998;18(1):26-42.
186. Seiffert E, Dreier JP, Ivens S, Bechmann I, Tomkins O, Heinemann U, et al. Lasting blood-brain barrier disruption induces epileptic focus in the rat somatosensory cortex. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2004;24(36):7829-36.
187. Ivens S, Kaufer D, Flores LP, Bechmann I, Zumsteg D, Tomkins O, et al. TGF-beta receptor-mediated albumin uptake into astrocytes is involved in neocortical epileptogenesis. *Brain*. 2007;130(Pt 2):535-47.
188. Tomkins O, Friedman O, Ivens S, Reiffurth C, Major S, Dreier JP, et al. Blood-brain barrier disruption results in delayed functional and structural alterations in the rat neocortex. *Neurobiology of disease*. 2007;25(2):367-77.
189. van Vliet EA, Zibell G, Pekcec A, Schlichtiger J, Edelbroek PM, Holtman L, et al. COX-2 inhibition controls P-glycoprotein expression and promotes brain delivery of phenytoin in chronic epileptic rats. *Neuropharmacology*. 2010;58(2):404-12.
190. Friedman A, Kaufer D, Heinemann U. Blood-brain barrier breakdown-inducing astrocytic transformation: novel targets for the prevention of epilepsy. *Epilepsy research*. 2009;85(2-3):142-9.
191. Weissberg I, Wood L, Kamintsky L, Vazquez O, Milikovsky DZ, Alexander A, et al. Albumin induces excitatory synaptogenesis through astrocytic TGF-beta/ALK5 signaling in a model of acquired epilepsy following blood-brain barrier dysfunction. *Neurobiology of disease*. 2015;78:115-25.
192. Hunsberger JG, Bennett AH, Selvanayagam E, Duman RS, Newton SS. Gene profiling the response to kainic acid induced seizures. *Brain research Molecular brain research*. 2005;141(1):95-112.
193. Dubey D, McRae PA, Rankin-Gee EK, Baranov E, Wandrey L, Rogers S, et al. Increased metalloproteinase activity in the hippocampus following status epilepticus. *Epilepsy research*. 2017;132:50-8.
194. Kim GW, Kim HJ, Cho KJ, Kim HW, Cho YJ, Lee BI. The role of MMP-9 in integrin-mediated hippocampal cell death after pilocarpine-induced status epilepticus. *Neurobiology of disease*. 2009;36(1):169-80.
195. Penkowa M, Florit S, Giral M, Quintana A, Molinero A, Carrasco J, et al. Metallothionein reduces central nervous system inflammation, neurodegeneration, and cell death following kainic acid-induced epileptic seizures. *Journal of neuroscience research*. 2005;79(4):522-34.
196. Lee J, Lim E, Kim Y, Li E, Park S. Ghrelin attenuates kainic acid-induced neuronal cell death in the mouse hippocampus. *The Journal of endocrinology*. 2010;205(3):263-70.
197. Jourquin J, Tremblay E, Decanis N, Charton G, Hanessian S, Chollet AM, et al. Neuronal activity-dependent increase of net matrix metalloproteinase activity is associated with MMP-9 neurotoxicity after kainate. *Eur J Neurosci*. 2003;18(6):1507-17.
198. Zhang JW, Deb S, Gottschall PE. Regional and age-related expression of gelatinases in the brains of young and old rats after treatment with kainic acid. *Neurosci Lett*. 2000;295(1-2):9-12.
199. Zhang JW, Deb S, Gottschall PE. Regional and differential expression of gelatinases in rat brain after systemic kainic acid or bicuculline administration. *Eur J Neurosci*. 1998;10(11):3358-68.
200. Guilfoyle MR, Carpenter KL, Helmy A, Pickard JD, Menon DK, Hutchinson PJ. Matrix Metalloproteinase Expression in Contusional Traumatic Brain Injury: A Paired Microdialysis Study. *Journal of neurotrauma*. 2015;32(20):1553-9.
201. Wilczynski GM, Konopacki FA, Wilczek E, Lasiecka Z, Gorlewicz A, Michaluk P, et al. Important role of matrix metalloproteinase 9 in epileptogenesis. *The Journal of cell biology*. 2008;180(5):1021-35.
202. Pijet B, Stefaniuk M, Kostrzewska-Ksiezka A, Tsilibary PE, Tzinia A, Kaczmarek L. Elevation of MMP-9 Levels Promotes

- Epileptogenesis After Traumatic Brain Injury. *Mol Neurobiol*. 2018.
203. Bush JA, Li G. Regulation of the Mdr1 isoforms in a p53-deficient mouse model. *Carcinogenesis*. 2002;23(10):1603-7.
204. Morawski M, Reinert T, Meyer-Klaucke W, Wagner FE, Troger W, Reinert A, et al. Ion exchanger in the brain: Quantitative analysis of perineuronally fixed anionic binding sites suggests diffusion barriers with ion sorting properties. *Sci Rep*. 2015;5:16471.
205. Hartig W, Derouiche A, Welt K, Brauer K, Grosche J, Mader M, et al. Cortical neurons immunoreactive for the potassium channel Kv3.1b subunit are predominantly surrounded by perineuronal nets presumed as a buffering system for cations. *Brain research*. 1999;842(1):15-29.
206. Cammarota M, Losi G, Chiavagato A, Zonta M, Carmignoto G. Fast spiking interneuron control of seizure propagation in a cortical slice model of focal epilepsy. *J Physiol*. 2013;591(4):807-22.
207. Hwang JJ, Park MH, Choi SY, Koh JY. Activation of the Trk signaling pathway by extracellular zinc. Role of metalloproteinases. *The Journal of biological chemistry*. 2005;280(12):11995-2001.
208. Lee R, Kermani P, Teng KK, Hempstead BL. Regulation of cell survival by secreted proneurotrophins. *Science (New York, NY)*. 2001;294(5548):1945-8.
209. Martinez MA, Ubeda A, Trillo MA. Involvement of the EGF Receptor in MAPK Signaling Activation by a 50 Hz Magnetic Field in Human Neuroblastoma Cells. *Cellular physiology and biochemistry : international journal of experimental cellular physiology, biochemistry, and pharmacology*. 2019;52(4):893-907.
210. Sternlicht MD, Werb Z. How matrix metalloproteinases regulate cell behavior. *Annual review of cell and developmental biology*. 2001;17:463-516.
211. Mizoguchi H, Nakade J, Tachibana M, Ibi D, Someya E, Koike H, et al. Matrix metalloproteinase-9 contributes to kindled seizure development in pentylenetetrazole-treated mice by converting pro-BDNF to mature BDNF in the hippocampus. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2011;31(36):12963-71.
212. Dudek FE, Sutula TP. Epileptogenesis in the dentate gyrus: a critical perspective. *Progress in brain research*. 2007;163:755-73.
213. Dudek FE, Shao LR. Mossy fiber sprouting and recurrent excitation: direct electrophysiologic evidence and potential implications. *Epilepsy Curr*. 2004;4(5):184-7.
214. Tian L, Stefanidakis M, Ning L, Van Lint P, Nyman-Huttunen H, Libert C, et al. Activation of NMDA receptors promotes dendritic spine development through MMP-mediated ICAM-5 cleavage. *The Journal of cell biology*. 2007;178(4):687-700.
215. Michaluk P, Wawrzyniak M, Alot P, Szczot M, Wyrembek P, Mercik K, et al. Influence of matrix metalloproteinase MMP-9 on dendritic spine morphology. *J Cell Sci*. 2011;124(Pt 19):3369-80.
216. Lonskaya I, Partridge J, Lalchandani RR, Chung A, Lee T, Vicini S, et al. Soluble ICAM-5, a product of activity dependent proteolysis, increases mEPSC frequency and dendritic expression of GluA1. *PLoS one*. 2013;8(7):e69136.
217. Coenen AM, Van Lujtelaar EL. Genetic animal models for absence epilepsy: a review of the WAG/Rij strain of rats. *Behavior genetics*. 2003;33(6):635-55.
218. Schilditzki A, Twele F, Klee R, Waltl I, Romermann K, Broer S, et al. A combination of NMDA and AMPA receptor antagonists retards granule cell dispersion and epileptogenesis in a model of acquired epilepsy. *Sci Rep*. 2017;7(1):12191.
219. Szczurowska E, Mares P. NMDA and AMPA receptors: development and status epilepticus. *Physiological research*. 2013;62 Suppl 1:S21-38.
220. Chen J, Cai F, Jiang L, Hu Y, Feng C. A prospective study of dexamethasone therapy in refractory epileptic encephalopathy with continuous spike-and-wave during sleep. *Epilepsy Behav*. 2016;55:1-5.
221. Verhelst H, Boon P, Buyse G, Ceulemans B, D'Hooghe M, Meirleir LD, et al. Steroids in intractable childhood epilepsy: clinical experience and review of the literature. *Seizure*. 2005;14(6):412-21.
222. Xu D, Robinson AP, Ishii T, Duncan DS, Alden TD, Goings GE, et al. Peripherally derived T regulatory and gammadelta T cells have opposing roles in the pathogenesis of intractable pediatric epilepsy. *The Journal of experimental medicine*. 2018;215(4):1169-86.
223. Yoshikawa K, Kita Y, Kishimoto K, Shimizu T. Profiling of eicosanoid production in the rat hippocampus during kainic acid-induced seizure: dual phase regulation and differential involvement of COX-1 and COX-2. *The Journal of biological chemistry*. 2006;281(21):14663-9.
224. Xu JH, Long L, Tang YC, Zhang JT, Hut HT, Tang FR. CCR3, CCR2A and macrophage inflammatory protein (MIP)-1a, monocyte chemoattractant protein-1 (MCP-1) in the mouse hippocampus during and after pilocarpine-induced status epilepticus (PISE). *Neuropathology and applied neurobiology*. 2009;35(5):496-514.
225. Vezzani A, Balosso S, Ravizza T. Neuroinflammatory pathways as treatment targets and biomarkers in epilepsy. *Nature reviews Neurology*. 2019;15(8):459-72.
226. Engel T, Martinez-Villarreal J, Henke C, Jimenez-Mateos EM, Sanz-Rodriguez A, Alves M, et al. Spatiotemporal progression of ubiquitin-proteasome system inhibition

- after status epilepticus suggests protective adaptation against hippocampal injury. *Molecular neurodegeneration*. 2017;12(1):21.
227. Citraro R, Leo A, Constanti A, Russo E, De Sarro G. mTOR pathway inhibition as a new therapeutic strategy in epilepsy and epileptogenesis. *Pharmacol Res*. 2016;107:333-43.
228. Drion CM, Borm LE, Kooijman L, Aronica E, Wadman WJ, Hartog AF, et al. Effects of rapamycin and curcumin treatment on the development of epilepsy after electrically induced status epilepticus in rats. *Epilepsia*. 2016;57(5):688-97.
229. van Vliet EA, Forte G, Holtman L, den Burger JC, Sinjewel A, de Vries HE, et al. Inhibition of mammalian target of rapamycin reduces epileptogenesis and blood-brain barrier leakage but not microglia activation. *Epilepsia*. 2012;53(7):1254-63.
230. Huang X, Zhang H, Yang J, Wu J, McMahon J, Lin Y, et al. Pharmacological inhibition of the mammalian target of rapamycin pathway suppresses acquired epilepsy. *Neurobiology of disease*. 2010;40(1):193-9.
231. Zeng LH, Rensing NR, Wong M. The mammalian target of rapamycin signaling pathway mediates epileptogenesis in a model of temporal lobe epilepsy. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2009;29(21):6964-72.
232. Guo D, Zeng L, Brody DL, Wong M. Rapamycin attenuates the development of posttraumatic epilepsy in a mouse model of traumatic brain injury. *PLoS one*. 2013;8(5):e64078.
233. Butler CR, Boychuk JA, Smith BN. Effects of Rapamycin Treatment on Neurogenesis and Synaptic Reorganization in the Dentate Gyrus after Controlled Cortical Impact Injury in Mice. *Frontiers in systems neuroscience*. 2015;9:163.
234. Hester MS, Hosford BE, Santos VR, Singh SP, Rolle IJ, LaSarge CL, et al. Impact of rapamycin on status epilepticus induced hippocampal pathology and weight gain. *Experimental neurology*. 2016;280:1-12.
235. Buckmaster PS, Ingram EA, Wen X. Inhibition of the mammalian target of rapamycin signaling pathway suppresses dentate granule cell axon sprouting in a rodent model of temporal lobe epilepsy. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2009;29(25):8259-69.
236. Buckmaster PS, Lew FH. Rapamycin suppresses mossy fiber sprouting but not seizure frequency in a mouse model of temporal lobe epilepsy. *The Journal of neuroscience : the official journal of the Society for Neuroscience*. 2011;31(6):2337-47.
237. Zeng LH, McDaniel S, Rensing NR, Wong M. Regulation of cell death and epileptogenesis by the mammalian target of rapamycin (mTOR): A double-edged sword? *Cell Cycle*. 2010;9(12):2281-85.
238. Hailer NP. Immunosuppression after traumatic or ischemic CNS damage: it is neuroprotective and illuminates the role of microglial cells. *Progress in neurobiology*. 2008;84(3):211-33.
239. Erlich S, Alexandrovich A, Shohami E, Pinkas-Kramarski R. Rapamycin is a neuroprotective treatment for traumatic brain injury. *Neurobiology of disease*. 2007;26(1):86-93.
240. Dello Russo C, Lisi L, Tringali G, Navarra P. Involvement of mTOR kinase in cytokine-dependent microglial activation and cell proliferation. *Biochemical pharmacology*. 2009;78(9):1242-51.
241. Lu DY, Liou HC, Tang CH, Fu WM. Hypoxia-induced iNOS expression in microglia is regulated by the PI3-kinase/Akt/mTOR signaling pathway and activation of hypoxia inducible factor-1alpha. *Biochemical pharmacology*. 2006;72(8):992-1000.
242. Chi OZ, Mellender SJ, Barsoum S, Liu X, Damito S, Weiss HR. Effects of rapamycin pretreatment on blood-brain barrier disruption in cerebral ischemia-reperfusion. *Neurosci Lett*. 2016;620:132-6.
243. Van Skike CE, Jahrling JB, Olson AB, Sayre NL, Hussong SA, Ungvari Z, et al. Inhibition of mTOR protects the blood-brain barrier in models of Alzheimer's disease and vascular cognitive impairment. *Am J Physiol Heart Circ Physiol*. 2018;314(4):H693-h703.
244. Krueger DA, Care MM, Holland K, Agricola K, Tudor C, Mangeshkar P, et al. Everolimus for subependymal giant-cell astrocytomas in tuberous sclerosis. *The New England journal of medicine*. 2010;363(19):1801-11.
245. Franz DN, Agricola K, Mays M, Tudor C, Care MM, Holland-Bouley K, et al. Everolimus for subependymal giant cell astrocytoma: 5-year final analysis. *Annals of neurology*. 2015;78(6):929-38.
246. French JA, Lawson JA, Yapici Z, Ikeda H, Polster T, Nabbout R, et al. Adjunctive everolimus therapy for treatment-resistant focal-onset seizures associated with tuberous sclerosis (EXIST-3): a phase 3, randomised, double-blind, placebo-controlled study. *Lancet (London, England)*. 2016.
247. Curatolo P, Franz DN, Lawson JA, Yapici Z, Ikeda H, Polster T, et al. Adjunctive everolimus for children and adolescents with treatment-refractory seizures associated with tuberous sclerosis complex: post-hoc analysis of the phase 3 EXIST-3 trial. *The Lancet Child & adolescent health*. 2018;2(7):495-504.
248. Reschke CR, Henshall DC. microRNA and Epilepsy. *Adv Exp Med Biol*. 2015;888:41-70.
249. Korotkov A, Mills JD, Gorter JA, van Vliet EA, Aronica

- E. Systematic review and meta-analysis of differentially expressed miRNAs in experimental and human temporal lobe epilepsy. *Sci Rep.* 2017;7(1):11592.
250. van Scheppingen J, Mills JD, Zimmer TS, Broekaart DWM, Iori V, Bongaarts A, et al. miR147b: A novel key regulator of interleukin 1 beta-mediated inflammation in human astrocytes. *Glia.* 2018;66(5):1082-97.
251. Iori V, Iyer AM, Ravizza T, Beltrame L, Paracchini L, Marchini S, et al. Blockade of the IL-1R1/TLR4 pathway mediates disease-modification therapeutic effects in a model of acquired epilepsy. *Neurobiology of disease.* 2017;99:12-23.
252. Cai Z, Li S, Li S, Song F, Zhang Z, Qi G, et al. Antagonist Targeting microRNA-155 Protects against Lithium-Pilocarpine-Induced Status Epilepticus in C57BL/6 Mice by Activating Brain-Derived Neurotrophic Factor. *Front Pharmacol.* 2016;7:129.
253. Fiore R, Khudayberdiyev S, Christensen M, Siegel G, Flavell SW, Kim TK, et al. Mef2-mediated transcription of the miR379-410 cluster regulates activity-dependent dendritogenesis by fine-tuning Pumilio2 protein levels. *The EMBO journal.* 2009;28(6):697-710.
254. Jimenez-Mateos EM, Engel T, Merino-Serrais P, McKiernan RC, Tanaka K, Moury G, et al. Silencing microRNA-134 produces neuroprotective and prolonged seizure-suppressive effects. *Nat Med.* 2012;18(7):1087-94.
255. Reschke CR, Silva LF, Norwood BA, Senthilkumar K, Morris G, Sanz-Rodriguez A, et al. Potent Anti-seizure Effects of Locked Nucleic Acid Antagomirs Targeting miR-134 in Multiple Mouse and Rat Models of Epilepsy. *Mol Ther Nucleic Acids.* 2017;6:45-56.
256. Morris G, Reschke CR, Henshall DC. Targeting microRNA-134 for seizure control and disease modification in epilepsy. *EBioMedicine.* 2019;45:646-54.
257. Morris G, Brennan GP, Reschke CR, Henshall DC, Schorge S. Sparing CA1 pyramidal neuron function and hippocampal performance following antisense knockdown of microRNA-134. *Epilepsia.* 2018;59(8):1518-26.
258. Gao X, Guo M, Meng D, Sun F, Guan L, Cui Y, et al. Silencing MicroRNA-134 Alleviates Hippocampal Damage and Occurrence of Spontaneous Seizures After Intraventricular Kainic Acid-Induced Status Epilepticus in Rats. *Frontiers in cellular neuroscience.* 2019;13:145.
259. Vangoor VR, Reschke CR, Senthilkumar K, van de Haar LL, de Wit M, Giuliani G, et al. Antagonizing Increased miR-135a Levels at the Chronic Stage of Experimental TLE Reduces Spontaneous Recurrent Seizures. *The Journal of neuroscience : the official journal of the Society for Neuroscience.* 2019;39(26):5064-79.
260. Xie Y, Shao Y, Deng X, Wang M, Chen Y. MicroRNA-298 Reverses Multidrug Resistance to Antiepileptic Drugs by Suppressing MDR1/P-gp Expression in vitro. *Frontiers in neuroscience.* 2018;12:602.
261. Hsiao J, Yuan TY, Tsai MS, Lu CY, Lin YC, Lee ML, et al. Upregulation of Haploinsufficient Gene Expression in the Brain by Targeting a Long Non-coding RNA Improves Seizure Phenotype in a Model of Dravet Syndrome. *EBioMedicine.* 2016;9:257-77.
262. Rupaimoole R, Slack FJ. MicroRNA therapeutics: towards a new era for the management of cancer and other diseases. *Nat Rev Drug Discov.* 2017;16(3):203-22.
263. Henshall DC. Manipulating MicroRNAs in Murine Models: Targeting the Multi-Targeting in Epilepsy. *Epilepsy Curr.* 2017;17(1):43-7.
264. Tao H, Zhao J, Liu T, Cai Y, Zhou X, Xing H, et al. Intranasal Delivery of miR-146a Mimics Delayed Seizure Onset in the Lithium-Pilocarpine Mouse Model. *Mediators Inflamm.* 2017;2017:6512620.
265. Asahi M, Wang X, Mori T, Sumii T, Jung JC, Moskowitz MA, et al. Effects of matrix metalloproteinase-9 gene knock-out on the proteolysis of blood-brain barrier and white matter components after cerebral ischemia. *The Journal of neuroscience : the official journal of the Society for Neuroscience.* 2001;21(19):7724-32.
266. Svedin P, Hagberg H, Savman K, Zhu C, Mallard C. Matrix metalloproteinase-9 gene knock-out protects the immature brain after cerebral hypoxia-ischemia. *The Journal of neuroscience : the official journal of the Society for Neuroscience.* 2007;27(7):1511-8.
267. Gurney KJ, Estrada EY, Rosenberg GA. Blood-brain barrier disruption by stromelysin-1 facilitates neutrophil infiltration in neuroinflammation. *Neurobiology of disease.* 2006;23(1):87-96.
268. Burggraf D, Trinkl A, Dichgans M, Hamann GF. Doxycycline inhibits MMPs via modulation of plasminogen activators in focal cerebral ischemia. *Neurobiology of disease.* 2007;25(3):506-13.
269. Copin JC, Merlani P, Sugawara T, Chan PH, Gasche Y. Delayed matrix metalloproteinase inhibition reduces intracerebral hemorrhage after embolic stroke in rats. *Experimental neurology.* 2008;213(1):196-201.
270. Lee H, Park JW, Kim SP, Lo EH, Lee SR. Doxycycline inhibits matrix metalloproteinase-9 and laminin degradation after transient global cerebral ischemia. *Neurobiology of disease.* 2009;34(2):189-98.
271. Pires PW, Rogers CT, McClain JL, Garver HS, Fink GD, Dorrance AM. Doxycycline, a matrix metalloprotease inhibitor, reduces vascular remodeling and damage after cerebral ischemia in stroke-prone spontaneously hypertensive rats. *Am J Physiol Heart Circ Physiol.* 2011;301(1):H87-97.

272. Yrjanheikki J, Tikka T, Keinanen R, Goldsteins G, Chan PH, Koistinaho J. A tetracycline derivative, minocycline, reduces inflammation and protects against focal cerebral ischemia with a wide therapeutic window. *Proceedings of the National Academy of Sciences of the United States of America*. 1999;96(23):13496-500.
273. Bhatt LK, Addepalli V. Potentiation of aspirin-induced cerebroprotection by minocycline: a therapeutic approach to attenuate exacerbation of transient focal cerebral ischaemia. *Diabetes & vascular disease research*. 2012;9(1):25-34.
274. Ahmadirad N, Shojaei A, Javan M, Pourgholami MH, Mirnajafi-Zadeh J. Effect of minocycline on pentylenetetrazol-induced chemical kindled seizures in mice. *Neurol Sci*. 2014;35(4):571-6.
275. Arisi GM, Foresti ML, Montañez A, Shapiro LA. Minocycline ameliorates neuronal loss after pilocarpine-induced status epilepticus. *J Neurol Disord Stroke*. 2014;2(3):1055.
276. Beheshti Nasr SM, Moghimi A, Mohammad-Zadeh M, Shamsizadeh A, Noorbakhsh SM. The effect of minocycline on seizures induced by amygdala kindling in rats. *Seizure*. 2013;22(8):670-4.
277. Blum D, Chtarto A, Tenenbaum L, Brotchi J, Levivier M. Clinical potential of minocycline for neurodegenerative disorders. *Neurobiology of disease*. 2004;17(3):359-66.
278. Yang L, Sugama S, Chirichigno JW, Gregorio J, Lorenzl S, Shin DH, et al. Minocycline enhances MPTP toxicity to dopaminergic neurons. *Journal of neuroscience research*. 2003;74(2):278-85.
279. Bocker R, Estler CJ, Ludewig-Sandig D. Evaluation of the hepatotoxic potential of minocycline. *Antimicrobial agents and chemotherapy*. 1991;35(7):1434-6.
280. Nowak M, Strzelczyk A, Reif PS, Schorlemmer K, Bauer S, Norwood BA, et al. Minocycline as potent anticonvulsant in a patient with astrocytoma and drug resistant epilepsy. *Seizure*. 2012;21(3):227-8.
281. Jacobsen JA, Major Jourden JL, Miller MT, Cohen SM. To bind zinc or not to bind zinc: an examination of innovative approaches to improved metalloproteinase inhibition. *Biochimica et biophysica acta*. 2010;1803(1):72-94.
282. Blundell TL, Jhoti H, Abell C. High-throughput crystallography for lead discovery in drug design. *Nat Rev Drug Discov*. 2002;1(1):45-54.
283. Vandenbroucke RE, Libert C. Is there new hope for therapeutic matrix metalloproteinase inhibition? *Nat Rev Drug Discov*. 2014;13(12):904-27.
284. Cui J, Chen S, Zhang C, Meng F, Wu W, Hu R, et al. Inhibition of MMP-9 by a selective gelatinase inhibitor protects neurovasculature from embolic focal cerebral ischemia. *Molecular neurodegeneration*. 2012;7:21.
285. Ranasinghe HS, Scheepens A, Sirimanne E, Mitchell MD, Williams CE, Fraser M. Inhibition of MMP-9 activity following hypoxic ischemia in the developing brain using a highly specific inhibitor. *Developmental neuroscience*. 2012;34(5):417-27.
286. Jia F, Yin YH, Gao GY, Wang Y, Cen L, Jiang JY. MMP-9 inhibitor SB-3CT attenuates behavioral impairments and hippocampal loss after traumatic brain injury in rat. *Journal of neurotrauma*. 2014;31(13):1225-34.
287. Hadass O, Tomlinson BN, Gooyit M, Chen S, Purdy JJ, Walker JM, et al. Selective inhibition of matrix metalloproteinase-9 attenuates secondary damage resulting from severe traumatic brain injury. *PLoS one*. 2013;8(10):e76904.
288. Wu MY, Gao F, Yang XM, Qin X, Chen GZ, Li D, et al. Matrix metalloproteinase-9 regulates the blood brain barrier via the hedgehog pathway in a rat model of traumatic brain injury. *Brain research*. 2020;1727:146553.
289. Yeghiazaryan M, Rutkowska-Wlodarczyk I, Konopka A, Wilczynski GM, Melikyan A, Korkotian E, et al. DP-b99 modulates matrix metalloproteinase activity and neuronal plasticity. *PLoS one*. 2014;9(6):e99789.
290. Barkalifa R, Hershinkel M, Friedman JE, Kozak A, Sekler I. The lipophilic zinc chelator DP-b99 prevents zinc induced neuronal death. *European journal of pharmacology*. 2009;618(1-3):15-21.
291. Lees KR, Bornstein N, Diener HC, Gorelick PB, Rosenberg G, Shuaib A, et al. Results of Membrane-Activated Chelator Stroke Intervention randomized trial of DP-b99 in acute ischemic stroke. *Stroke*. 2013;44(3):580-4.
292. Bertran A, Khomiak D, Konopka A, Rejmak E, Bulska E, Seco J, et al. Design and synthesis of selective and blood-brain barrier-permeable hydroxamate-based gelatinase inhibitors. *Bioorganic chemistry*. 2019:103365.
293. Feng S, Cen J, Huang Y, Shen H, Yao L, Wang Y, et al. Matrix metalloproteinase-2 and -9 secreted by leukemic cells increase the permeability of blood-brain barrier by disrupting tight junction proteins. *PLoS one*. 2011;6(8):e20599.
294. Yang Y, Estrada EY, Thompson JF, Liu W, Rosenberg GA. Matrix metalloproteinase-mediated disruption of tight junction proteins in cerebral vessels is reversed by synthetic matrix metalloproteinase inhibitor in focal ischemia in rat. *Journal of cerebral blood flow and metabolism : official journal of the International Society of Cerebral Blood Flow and Metabolism*. 2007;27(4):697-709.
295. Higashida T, Kreipke CW, Rafols JA, Peng C, Schafer S, Schafer P, et al. The role of hypoxia-inducible factor-1 α , aquaporin-4, and matrix metalloproteinase-9 in blood-brain barrier disruption and brain edema after traumatic brain injury. *J Neurosurg*. 2011;114(1):92-101.