miR-34b/c Regulates Wnt1 and Enhances Mesencephalic Dopaminergic Neuron Differentiation


DOI
10.1016/j.stemcr.2018.02.006

Publication date
2018

Document Version
Final published version

Published in
Stem Cell Reports

License
CC BY-NC-ND

Citation for published version (APA):

General rights
It is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), other than for strictly personal, individual use, unless the work is under an open content license (like Creative Commons).

Disclaimer/Complaints regulations
If you believe that digital publication of certain material infringes any of your rights or (privacy) interests, please let the Library know, stating your reasons. In case of a legitimate complaint, the Library will make the material inaccessible and/or remove it from the website. Please Ask the Library: https://uba.uva.nl/en/contact, or a letter to: Library of the University of Amsterdam, Secretariat, Singel 425, 1012 WP Amsterdam, The Netherlands. You will be contacted as soon as possible.
miR-34b/c Regulates Wnt1 and Enhances Mesencephalic Dopaminergic Neuron Differentiation

Roberto De Gregorio,1,11,12 Salvatore Pulcrano,1,3,12 Claudia De Sanctis,1,2 Floriana Volpicelli,1,3 Ezia Guatteo,8,9 Lars von Oertel,4 Emanuele Claudio Latagliata,8 Roberta Esposito,1 Rosa Maria Piscitelli,8,9 Carla Perrone-Capano,1,3 Valerio Costa,1 Stefano Puglisi-Allegra,8 Marten P. Smidt,4 Umberto di Porzio,1 Massimiliano Caiazzo,6 Nicola Biagio Mercuri,8,10 Meng Li,7 and Gian Carlo Bellenchi1,*

1Institute of Genetics and Biophysics, “Adriano Buzzati Traverso”, CNR, 80131 Naples, Italy
2Neuromed IRCCS, 86077 Pozzilli (IS), Italy
3Department of Pharmacy, University of Naples Federico II, 80131 Naples, Italy
4Swammerdam Institute for Life Sciences, University of Amsterdam, 1090 GE Amsterdam, the Netherlands
5Institute of Biotechnology, University of Helsinki, 00014 Helsinki, Finland
6Department of Pharmaceutics, Utrecht Institute for Pharmaceutical Sciences, 3584 CG Utrecht, the Netherlands
7Neuroscience and Mental Health Research Institute, School of Medicine and School of Bioscience, Cardiff University, Cardiff CF24 4HQ, UK
8Fondazione Santa Lucia IRCCS, 00143 Rome, Italy
9Parthenope University, Department of Motor Science and Wellness, 80133 Naples, Italy
10University of Tor Vergata, Department of Systems Medicine, 00133 Rome, Italy
11Present address: Department of Molecular Pharmacology, Albert Einstein College of Medicine, New York, NY 10461, USA
12Co-first author
*Correspondence: giancarlo.bellenchi@igb.cnr.it
https://doi.org/10.1016/j.stemcr.2018.02.006

SUMMARY

The differentiation of dopaminergic neurons requires concerted action of morphogens and transcription factors acting in a precise and well-defined time window. Very little is known about the potential role of microRNA in these events. By performing a microRNA-mRNA paired microarray screening, we identified miR-34b/c among the most upregulated microRNAs during dopaminergic differentiation. Interestingly, miR-34b/c modulates Wnt1 expression, promotes cell cycle exit, and induces dopaminergic differentiation. When combined with transcription factors ASCL1 and NURR1, miR-34b/c doubled the yield of transdifferentiated fibroblasts into dopaminergic neurons. Induced dopaminergic (iDA) cells synthesize dopamine and show spontaneous electrical activity, reversibly blocked by tetrodotoxin, consistent with the electrophysiological properties featured by brain dopaminergic neurons. Our findings point to a role for miR-34b/c in neuronal commitment and highlight the potential of exploiting its synergy with key transcription factors in enhancing in vitro generation of dopaminergic neurons.

INTRODUCTION

MicroRNAs (miRNAs) are a class of small non-coding single-strand RNA (~22 nucleotides) acting as post-transcriptional regulators of gene expression (Bartel and Chen, 2004). They are crucial players in several aspects of brain development such as neurogenesis, neuronal maturation, synapse formation, axon guidance, and neuronal plasticity (Kapsimali et al., 2007; McNeill and Van Vactor, 2012). Disruption of miRNA biogenesis by genetic deletion of Dicer has been a widely used strategy to investigate the role of miRNAs in neurodevelopmental processes (Cuellar et al., 2008; Davis et al., 2008; Kawase-Koga et al., 2009). Cell-type-specific deletion of miRNAs involved in mesencephalic dopaminergic (mDA) neurons causes progressive loss of these cells (Kim et al., 2007), suggesting a pivotal role for miRNA in mDA neuron formation, survival, and function. A number of laboratories attempted the identification of miRNAs involved in mDA neuron differentiation and diseases affecting the mDA system such as Parkinson’s disease (Kim et al., 2007; Miñones-Moyano et al., 2011; Saba et al., 2012; Tobon et al., 2012; Yang et al., 2012).

Interestingly, recent works highlighted the importance of miRNAs, miR-135a2 in particular, in determining midbrain size and the allocation of prospective mDA precursors by modulating the extent of the Wnt signaling. The latter takes place through direct regulation of Lmx1b, in a context where excessive Wnt signaling has been shown to lead to improper mDA specification (Joksimovic and Awatramani, 2014; Nouri et al., 2015). Thus, manipulation of Wnt signaling has become an attractive strategy to define new in vitro differentiation protocols to increase the fraction of mDA neurons.

In addition, miRNAs are also emerging as possible components in both direct and indirect cell reprogramming strategies. Specific miRNAs are able to potentiate transcription factor-mediated conversion of mouse embryonic fibroblasts (MEFs) into induced pluripotent stem cells (Judson et al., 2009; Subramanyam et al., 2011). While two miRNAs, miR-9/9* and miR-124, are able to convert fibroblasts into neurons (Yoo et al., 2011) or into a specific subset of neurons if combined with defined transcription factors (Victor et al., 2014), to date, the ability of miRNAs to enhance dopaminergic...
(DA) transdifferentiation (Caiazzo et al., 2011) is largely unexplored.

MiR-34b-5p and miR-34c-5p (hereafter named miR-34b/c) belong to a large family of miRNAs that include miR-34a-5p and the miR-449 cluster (449a-5p, 449b-5p, and 449c-5p). They all share the same seed sequence and are considered as having overlapping roles. More recently it has been shown that miR-34a/b/c share only 20% of targets, suggesting that individual miR-34 family members exert largely unique roles (Navarro and Lieberman, 2015). Interestingly, miR-34b/c expression was altered in Parkinson’s disease (PD), where it was found significantly downregulated in brain areas of PD patients with different degrees of pathology, prior to the appearance of motor dysfunction (Míñones-Moyano et al., 2011).

In this work, we identified the miR-34b/c cluster as an additional modulator of the Wnt signaling pathway. Enforced expression of miR-34b/c significantly increases in vitro differentiation of DA neurons. We show that miR-34b/c is able to directly bind the Wnt1 3’UTR and to regulate its expression. Consistently, overexpression of miR-34b/c leads to reduction of Wnt1 and Lmx1b in DA-differentiated mouse embryonic stem cells (mESCs). In combination with the induced dopaminergic (iDA)-specific transcription factors ASCL1 and NURR1 (Caiazzo et al., 2011), miR-34b/c doubles the transdifferentiation efficiency, leading to the generation of functional iDA neurons with characteristic electrophysiological properties.

**RESULTS**

**Modulation of the Wnt Pathway during mDA Neuron Differentiation**

To investigate the roles of miRNAs in DA neuron fate specification, we performed a microarray screen and measured the expression of both mRNAs and miRNAs during DA differentiation of epiblast-derived stem cells (epiSCs) using a monolayer differentiation (MD) method developed previously (Jaeger et al., 2011). As a control, the same cells were differentiated in the basal condition without Sonic Hedgehog (SHH) and fibroblast growth factor 8 (FGF8), leading to the production of mostly GABAergic neurons. Samples were harvested at day 9 and day 14 MD (Figure 1A) and the RNAs processed for gene expression for mRNAs and miRNAs by microarray. To evaluate the activation of specific differentiation programs, we analyzed the expression of genes involved in developmental fate specifications and neuronal maturation at day 9 and day 14 MD in DA and control samples. In SHH/FGF8-treated cells mDA-associated transcripts were upregulated at both day 9 and day 14 while the expression of markers for GABAergic, glutamatergic, or adrenergic neurons were downregulated. Furthermore, we did not find any difference between the two conditions in the expression of markers for anterior/posterior identity (Figure 1B). The induction of DA fate was further confirmed by following the expression of selected mDA markers such as Th, Nurr1, Pitx3, Dmrt5, Foxa2a, and Lmx1a. All these gene markers showed a robust upregulation at the end the differentiation protocol in cultures treated with SHH and FGF8 (Figure 1C). In contrast, general neuronal markers, such as Tubb3, Ascl1, Map2, and Ngn2, have a similar expression profile in both DA and control conditions (Figure 1D). Consistent with the transcript analysis, immunostaining for tyrosine hydroxylase (TH) and GFP (for PITX3-GFP) at the end of the differentiation protocol confirmed the generation of DA neurons (Figure 1E).

By comparing gene expression profiles at day 9 and day 14 MD in SHH/FGF8-treated samples (david.ncifcrf.gov) (Huang et al., 2007a), we noticed that the Wnt signaling pathway becomes one of the most affected KEGG (Kyoto Encyclopedia of Genes and Genomes) pathways along the DA differentiation (Figure 2A).

We have previously performed microarray screening comparing the floor plate (ventral midbrain, mDA) versus basal plate (non-DA) transcripts at three embryonic time points: embryonic day 9.5 (E9.5), E10.5, and E12.5 (Gennet et al., 2011). Reanalysis of this dataset revealed that only five Wnts (Wnt1, Wnt5a, Wnt5b, Wnt7a, and Wnt9a) are expressed at E9.5 but their expression decreases progressively until E12.5 (Figure 2B). The same genes were also affected in our microarray dataset, from day 9 and day 14 MD. By comparing epiSC differentiated toward an mDA phenotype with cells differentiated in control condition, giving rise to mainly GABAergic neurons, we found by qPCR that Wnt5a, Wnt5b, and Wnt9a increased during mDA differentiation while Wnt1 was maintained at a lower level (Figure 2C). The reduction of Wnt1 was paralleled by that of Lmx1b (Figure 2C), a transcription factor known to be involved in early stages of MHB formation (Guo et al., 2007) and was recently associated with Wnt1 as part of a regulatory loop together with miR-135a2-5p (Anderegg et al., 2013). Interestingly, the analysis of the array data show that most of the genes that positively affect the Wnt signaling were downregulated at day 14 MD, while some known inhibitors of the Wnt pathway, such as Gsk3b or Ctnnbip1, were upregulated (Figure 2D and scheme in Figure 2E). Together, these data suggest that the regulation of Wnt signaling during DA differentiation is needed to achieve proper generation of mDA neurons.

**miR-34b/c Targets Wnt1 and Is Enriched in Purified PITX3-GFP* mDA Neurons**

To test the hypothesis that negative regulation of the Wnt pathway could be achieved through the action of miRNAs, we screened miRNA-specific array data for miRNAs upregulated during DA differentiation. By using an available
miRNA target prediction tool (targetscan.org/mmu_71/) we scanned for all miRNAs upregulated both at day 9 and at day 14, in SHH/FGF8-treated cells compared with controls, which are predicted to bind to the 3’ UTR of downregulated genes in the Wnt signaling pathway. MiR-34b/c and miR-148a-3p both scored in

Figure 1. Profiling of epiSC-Derived Dopaminergic Cells
(A) Schematic representation of the protocols used to profile miRNAs and mRNAs expression during DA differentiation of mouse epiSCs. RNA samples were harvested in triplicate on day 9 MD and day 14 MD.
(B) The expression of genes representative for specific developmental fate and for different neuronal populations is shown at day 9 and day 14 MD. Data are presented as log(DA/Ctrl).
(C and D) qPCR for the expression of (C) DA-specific markers or (D) general neuronal markers over the differentiation protocol from day 6 MD to day 14 MD. All qPCR data have been normalized to the average of the reference gene Hmbs. The highest value for each gene (among both +SHH/FGF8 and −SHH/FGF8 samples) was set to 1. All other values are expressed as a ratio of 1. Data represent mean ± SEM from three independent experiments.
(E) Pitx3-GFP epiSC at day 14 MD differentiated in the presence (+) or absence (−) of SHH and FGF8 and immunostained for TH (red) and GFP (green) expression. High-magnification images for TH (yellow) with DAPI (blue) counterstain. Scale bars represent 200 μm and 100 μm, respectively, for the low- and high-magnification images.
Figure 2. The Wnt Pathway during mDA Differentiation of epiSCs

(A) Top ten categories of KEGG pathways associated with differentially expressed genes between day 9 MD and day 14 MD of the dopaminergic differentiation protocol (+SHH/FGF8). The p values were calculated using hypergeometric test and corrected by Benjamini-Hochberg adjustment.

(B) Relative expression of the most expressed Wnt genes in floor plate and basal plate during embryonic development, from E9.5 to E12.5.

(C) qPCR for the expression of Wnt5a, Wnt5b, Wnt7a, Wnt9a, Wnt1, and Lmx1b during epiSC differentiation into DA (+SHH/FGF8) or control (−SHH/FGF8) neurons. Data represent mean ± SEM from three independent experiments.

(legend continued on next page)
our list with 18 and 13 predicted binding sites, respectively (Figure 3A), and were predicted also to directly target Wnt1 (Figure 3B).

Pitx3-GFP is a mouse line in which the mDA neurons are exclusively labeled by the knockin GFP reporter (Zhang et al., 2004). Interestingly both miR-34b/c and miR-148a-3p were enriched in fluorescence-activated cell sorting (FACS)-purified GFP + cells obtained from E12.5 and E13.5 Pitx3-GFP embryos (Figure 3C), confirming their expression in mDA neurons in vivo. Hence, we investigated whether miR-34b/c and miR-148a-3p could act as post-transcriptional regulators of Wnt1. To this purpose we performed luciferase assays by transfecting plasmids expressing both miRNAs with a pmirGlo-reporter containing Wnt1 3′ UTR in HeLa cells. Both miR-34b/c and miR-148a-3p were able to significantly reduce luciferase activity (34.6% and 20%, respectively). The effect was abolished after mutation of the predicted binding site for miR-34b/c but not for miR-148a-3p, suggesting that only miR-34b/c effectively binds to its predicted site at Wnt1 3′ UTR (Figures 3B and 3D).

To further confirm that miR-34b/c and the Wnt signaling pathway could be involved in early phases of mDA differentiation, we used a dual-fluorescent GFP-reporter/monomeric red fluorescent protein (mRFP)-sensor plasmid (De Pietri Tonelli et al., 2006), which allows the detection of miRNAs at single-cell resolution. We cloned a tandem cassette complementary to miR-34b/c in the 3′ UTR of the mRFP sensor (pDSV3-34) or mutated in the region corresponding to the “seed sequence” of miR-34b/c (pDSV3-34mut). This approach has been described as very efficient for monitoring the endogenous expression of miRNAs both in vitro and in vivo. In differentiating mESCs transfected with pDSV3-34, the expression of the mRFP sensor was strongly reduced 72 hr after transfection (left column in Figure 3E). This effect was abolished with pDSV3-34mut (Figure 3E), thus suggesting that miR-34b/c is expressed in vitro during the DA differentiation of mESCs. Transfection of mESCs with the empty plasmids (pDSV2 and pDSV3) does not affect GFP or mRFP-sensor expression (Figure S1).

The endogenous miR-34b/c was also able to downregulate the expression of the mRFP sensor containing the entire Wnt1 3′ UTR sequence downstream of the coding sequence (CDS) (pDSV2-UTR). A reduction for the mRFP sensor was clearly visible in mESCs transfected with pDSV2-UTR 72 hr after transfection (second right column in Figure 3E). Mutation in the binding site for miR-34b/c (pDSV2-UTRmut) did not affect the mRFP-sensor expression (right column in Figure 3E).

**Mir-34b/c Enhances mESC Dopaminergic Differentiation**

Our data suggest that miR-34b/c has a role in DA neuron differentiation. To further corroborate this finding, we cloned ~900-bp genomic DNA encompassing the miR-34b/c cluster into an inducible lentiviral vector upstream of an IRES-GFP sequence (lenti-miR34b/c-Ires-GFP) and overexpressed the miR-34b/c cluster during the in vitro DA differentiation of mESCs. mESCs were infected with the lenti-miR34b/c-Ires-GFP vector (or with an inducible IRES-GFP as control), treated with doxycycline for 1 day and FACS purified for GFP. GFP + mESCs were amplified in the absence of doxycycline and then differentiated toward the DA phenotype in the presence or absence of doxycycline for 14 days (see scheme in Figure 4A). At the end of the differentiation, the expression of miR-34c was enriched 16-fold (Figure 4B), while no difference was observed for miR-34c in mock-infected GFP + mESCs (Figure 4B).

The overexpression of miR-34b/c cluster induced a downregulation in the mRNA levels for both Wnt1 and Lmx1b (Figure 4C). Consistent with the RNA data, immunostaining revealed a significant reduction of LMX1B + cells (Figure S2). In parallel, we observed a reduced amount of miRNAs for Lef1 and Axin2, thus supporting a role of miR-34b/c as a negative regulator of the Wnt signaling (Figure 4C). The overexpression of miR-34b/c also promoted DA differentiation. Indeed, we found a significant upregulation of mDA markers such as Th, Dat, Vmat2, and Pitx3 (Figure 4D). In addition, miR34b/c overexpression resulted in a ~60% increase in the number of TH + cells compared with control cultures (Figures 4E and 4F).

**Mir-34b/c Enhances Fibroblast Transdifferentiation into Induced Dopaminergic Neurons**

It has been shown that direct reprogramming represents a powerful source of iDA, useful for functional studies and cell replacement approaches. Hence, we further investigated whether miR-34b/c could increase the yield of DA neurons in such an experimental paradigm. To this purpose, we infected MEFs derived from mice expressing GFP under the control of the Th promoter (Sawamoto et al., 2001) with inducible lentiviruses expressing miR-34b/c cluster in combination with the reprogramming transcription factors Ascl1 and Nurr1 (renamed also A and N) (Caiazzo et al., 2011).
The expression of miR-34b/c in combination with Ascl1 and Nurr1 was induced 1 day after the infection by adding doxycycline to the culture medium. Cells were then differentiated for 14 days (day 14 MD) before analyzing the amount of TH⁺ and GFP⁺ cells. FACS analysis revealed that the combination of miR-34b/c with ASCL1 and NURR1 (AN versus AN + 34b/c) increased the number of GFP⁺ cells from 10.1% ± 1.7% to 19.5% ± 2.4% (Figures 5A and 5B) while miR-34b/c alone or in combination with ASCL1 was unable to induce GFP expression (our unpublished data). Such a difference was further verified by an increased level of TH mRNA (Figure 5H) and the number of TH⁺ cells following quantification of captured images (Figures 5C and 5E).

The addition of miR-34b/c affects terminal differentiation of DA neurons. Indeed, the expression of late mDA-specific markers such as Dat and Vmat2 were increased at mRNA level when miR-34b/c was combined with ASCL1 and NURR1 (AN34 versus AN in Figures S1 and S5).
To understand whether miR-34b/c acts by affecting cell cycle progression, we infected the neuronal cell line A1 (Colucci-D’Amato et al., 1999) with an inducible lentiviral vector expressing miR-34b/c upstream of an Ires-GFP sequence. Cells were FACs purified, amplified, and differentiated toward the DA phenotype.

To investigate whether the modulation of Wnt signaling may have a role in the transdifferentiation process, we infected MEFs derived from TH-GFP mice with Ascl1, Nurr1, and miR-34b/c in the presence of CHIR99021 (chiron), a selective inhibitor of GSK3β that can mimic Wnt activation. We found that while chiron did not alter the level of Wnt1 (AN34C in Figure 5F), it elicited a strong upregulation of Lef1 (AN34C in Figure 5G), suggesting an activation of Wnt signaling as expected. This effect was accompanied by a significant reduction in both the number of TH+ neurons (AN34C in Figures 5C, 5D, and 5H) and the transcript level of several mDA markers such as Vmat2 and Dat (AN34C in Figures 5H and 5J).

**Figure 4. Enforced Expression of miR-34b/c Promotes ESC DA Differentiation by Downregulating Wnt Signaling**

(A) Schematic representation of the experimental procedure. mESCs were infected with an inducible lentiviral vector expressing miR-34b/c upstream of an Ires-GFP sequence. Cells were FACS purified, amplified, and differentiated toward the DA phenotype.

(B) TaqMan assay for miR-34c in FACS purified mESCs in presence or absence of doxycycline (DOX). Data were normalized to the average of the reference sno-202. Data represent mean ± SEM. *p < 0.01 (Student’s t test).

(C and D) qPCR analysis of genes related to the Wnt pathway (Wnt1, Lmx1b, Axin2, and Lef1; C) and dopaminergic lineage (Th, Vmat2, Dat, and Pitx3; D) at day 14 MD in the presence or absence of DOX. Data represent mean ± SEM from three independent experiments. *p < 0.01 (Student’s t test).

(E and F) Immunostaining and quantifications for TH in mESCs at day 14 MD (E). Counting was performed from 20 randomly selected fields for each condition, in three independent experiments. Data represent mean ± SEM. *p < 0.05 relative to −DOX (Student’s t test). A higher-magnification image of DA-differentiated ESCs is shown in (F). TH (red) and TUBB3 (green). Scale bars represent 200 μm in (E) and 100 μm in (F).
Mir-34b/c Programmed iDA Cells Are Functionally Active

MEF-derived iDA by miR-34b/c direct reprogramming also contains higher amounts of dopamine, as shown after staining with an anti-DA antibody (Figures 6A and 6B). Higher amounts of double-positive (DA⁺, TH-GFP⁺) cells (16.1% ± 2.3% versus 6.9% ± 2.5%) were identified when miR-34b/c was included in the reprogramming cocktail (AN34 versus AN) (Figure 6B). Similarly, a higher content of dopamine was also shown by high-performance liquid chromatography (HPLC) measurements (Figure 6C), suggesting an increased dopamine synthesis in miR34-b/c-derived iDA.

To directly demonstrate that miR-34b/c-derived iDA are functional, we measured the action potential by performing targeted whole-cell electrophysiological recordings. TH-GFP⁺ cells derived from miR-34b/c displayed active neuronal properties. In extracellular recordings, we recorded spontaneous action potential firing at 3.93 ± 2.28 Hz in four cells. These events were reversibly blocked by TTX (1 μM), suggesting that they were mainly mediated by sodium currents (Figure 6D). In whole-cell recordings, depolarizing current injection of increasing amplitudes from a holding potential of −55/−60 mV (1 s, Figure 6E) evoked trains of action potentials in five cells, which were
completely blocked by TTX (Figure 6E, right), similar to those seen in ex vivo DA neurons. In response to the same protocol, another group of cells displayed membrane depolarization that did not reach the threshold for action potential generation (not shown).

In voltage-clamp recordings (Figure 6F, $V_h = -60\, \text{mV}$), we applied hyperpolarizing voltage steps (to $-120\, \text{mV}$, 20-mV increments) to activate the hyperpolarization-activated inward current, $I_h$, largely expressed in vivo by DA neurons of the substantia nigra pars compacta (Grace and Onn, 1989; Mercuri et al., 1995) and to a lesser extent by some neurons of the ventral tegmental area (Krashia et al., 2017) Two out of the 25 recorded GFP$^+$ cells displayed a small $I_h$ (Figure 6F) and a sag potential in response to hyperpolarizing current injections in current clamp mode (Figure 6G). Voltage-gated Na$^+$ and K$^+$ currents elicited by depolarizing voltage steps are also shown (Figure 6H). Taken together, these data confirm that miR-34b/c-derived iDA behave as functional active DA neurons.

**DISCUSSION**

**Mir-34b/c Is Expressed in mDA Neurons and Regulates Wnt1**

In the midbrain Wnt signaling is required for DA neurogenesis, and among the multiple canonical Wnts Wnt1 and
Wnt5α have a well-established role in promoting DA differentiation (Arenas, 2014). Here we investigate whether miRNAs could have a role in DA neuron differentiation alongside the Wnt signaling genes, since it was shown recently that restriction of Wnt1 and the downstream Wnt signaling occurs during mDA development and is achieved by a feedback loop including miR-135a2-5p that acts by repressing Lmx1b, and modulating WNT1/Wnt signaling (Anderegg et al., 2013). Interestingly we identified miR-135a2-5p as the miRNA with the highest number of predicted targets among the genes associated with Wnt pathway, which were downregulated at the late phases of our DA differentiation experiment (between day 9 and day 14 MD), confirming the relevance of our analysis aimed at identifying novel candidate miRNAs playing a role during DA differentiation. This approach allowed us to identify also miR-148a-3p and miR-34b/c, two miRNAs, enriched in FACS-purified mDA neurons, which are predicted to target Wnt1 3′UTR. Nevertheless only miR-34b/c binds the Wnt1 3′UTR and represses luciferase activity.

While the function of miR-34a in regulating Wnt signaling has been shown in both cancer progression and development (Kim et al., 2011, 2013), the involvement of miR-34b/c in this pathway has been largely neglected mainly because of the assumption that miRNAs sharing identical seed sequences target the same genes. It is now clear, however, that the role of individual miRNA should be considered relative to the cellular context and that sequence determinants outside the seed might profoundly affect miRNA binding through undefined mechanisms (Navarro and Lieberman, 2015; Ebner and Selbach, 2014).

Interestingly miR-34b/c was previously shown to be downregulated in postmortem brain of PD patients at different stages of pathology (Minones-Moyano et al., 2011). Nevertheless, few papers described the involvement of miR-34b/c in mDA neurons or reported DA-related phenotypes in miR-34b/c knockout mice (Comazzetto et al., 2014; Concepcion et al., 2012). This may be explained by the redundancy between the miR-34 family that includes three members of miR-449 (a, b, and c) in addition to miR-34a, b, and c. It is plausible that in vivo fine-tuning of the Wnt signaling might require the action of several miRNAs targeting simultaneously different actors of the pathway. In this context, it is not surprising that deletion of a single gene cluster has limited or no effects on mDA development.

Thus, although we identify Wnt1 as a target of miR-34b/c, it is unlikely that downregulation of Wnt1 by itself could drive mDA differentiation. We believe, indeed, that the miR-34b/c-mediated downregulation of Wnt1 is part of a complex series of events requiring the simultaneous intervention of other genes and pathways whose final aim is to define DA identity.

**Mir-34b/c Potentiates iDA Reprogramming**

Direct reprogramming has been used to achieve differentiation of fibroblasts into specific subtypes of mature neurons (Vierbuchen et al., 2010) and is considered a promising approach in terms of both regenerative medicine and modeling human diseases. For DA neurons this result has been achieved by combinatorial expression of early and late mDA transcription factors such as Ascl1, Lmx1a, and Nurr1 (Calazzo et al., 2011). miRNAs are considered a very promising additional tool to increase the yield of specific neuronal populations, as was shown by miR-9 and miR-124 (Victor et al., 2014). A similar mechanism may be proposed also for miR-34b/c, which increases mDA differentiation efficiency by facilitating cell cycle exit through the upregulation of a potent cyclin-dependent kinase inhibitor p21 (our unpublished data). The expression of miR-34b/c during the differentiation phases in combination with Ascl1 and NURR1 represses proliferation, promotes terminal differentiation, and downregulates Wnt1, which was maintained at high level by constant expression of Ascl1 (Rheinbay et al., 2013). The importance of miR-34b/c has also emerged in previous reports that placed miR-34 downstream of p53 (Kim et al., 2011). miRNAs might thus be required to shut down molecular programs to counteract differentiation. In DA neurons this is the case for miR-124 (Jiang et al., 2015) and miR-34b/c, which facilitate the generation of functional active DA neurons.

In the brain, miRNAs confer robustness to specific developmental processes and contribute to the maturation of specific neuronal circuits (Amin et al., 2015; Choi et al., 2008; Conaco et al., 2006; Cuellar et al., 2008; Tan et al., 2013, Stark et al., 2005). It is thus possible that miR-34b/c acts, at least in part, along the same axis by restraining Wnt signaling in the midbrain, thus facilitating cell cycle exit and allowing progenitor differentiation toward mature DA neurons. Taken together, our findings suggest that the achievement of functional mDA neurons requires the control over time of different stimuli (such as those mediated by the Wnt signaling) that are essential during early progenitor differentiation. It is plausible that other miRNAs are involved in the DA-differentiation process through the regulation of different pathways. It will be thus compelling to attain their identification in order to further understand the process of DA differentiation and increase the overall yield of DA neurons, as well as to improve their features in terms of both molecular and physiological properties.
EXPERIMENTAL PROCEDURES

Ethics Statement
All procedures involving mice were carried out in strict accordance with national and European law.

EpiSC DA Differentiation
Dopaminergic differentiation of epiSC was performed as previously described (Jaeger et al., 2011). The sequential steps of the protocol are described in the Supplemental Experimental Procedures.

RNA Extraction and qPCR
RNA was extracted from cells and tissues using the TRI-Reagent (Sigma, Milan, Italy) or mirVana miRNA isolation kit (Ambion, Milan, Italy). qPCR was carried out at least in triplicate samples using Power SYBR Green or TaqMan microRNA assays (Applied Biosystems, Milan, Italy) and analyzed as described in Supplemental Experimental Procedures.

Microarray Data Analysis
Microarray data were performed on epiSC differentiated toward the DA phenotype as described in Supplemental Experimental Procedures. Data are available at GEO: GSE110270.

Gene ontology and pathway analysis were performed on microarray data using the DAVID functional annotation tool (https://david.ncifcrf.gov) (Huang et al., 2007b) to find over-represented biological themes. Default DAVID parameters were used. To identify the pathways altered, we used the online tool available from Kanehsa laboratories, KEGG Mapper (Kanehsa et al., 2010). Regarding putative miRNA-mRNA interaction, for the identification of miRNA responsive elements within the 3’ UTRs of protein coding genes, we used TargetScan algorithm (Lewis et al., 2005).

PITX3-GFP* Cell Analysis
Freshly dissected ventral midbrains of Pitx3-GFP embryos were dissociated using a Papain dissociation system (Worthington, Milan, Italy). GFP-positive and -negative cells were isolated by a Cytopeia Influx Cell sorter or BD FACSARia III using previously described settings (Jacobs et al., 2011) and collected in RNAlater (Thermo Fisher Scientific, Milan, Italy). RNA was then purified by an miRVana miRNA isolation kit and analyzed by TaqMan MicroRNA Assays as described in Supplemental Experimental Procedures.

Immunocytochemistry
The following primary antibodies were used with the protocol described in Supplemental Experimental Procedures: anti-TUJ1 mouse 1:1000 (Covance, Rome, Italy, catalog #MMS-435P), anti-TH rabbit 1:500 (Chemicon, Milan, Italy, cat. #AB152), anti-LMX1B rabbit (a gift from Dr. Antonio Simeone), and anti-GFP mouse (Thermo Fisher Scientific, cat. #A11120). Dopamine staining was performed with an anti-dopamine rabbit (IS1005) + STAINperfect immunostaining kit A (SP-A-1000) (ImmunoSrol SAS, Bordeaux, France) according to the manufacturer’s instruction.

Image Quantification
Images were acquired using a Leica TCS SP5 confocal microscope. Quantification was performed either by using the cell counter plugin in the ImageJ software on randomly selected fields or by performing pixel analysis quantifications with a custom-made ImageJ plugin (rsb.info.nih.gov/ij/download/). In both cases quantification was performed from at least three independent experiments.

Luciferase Assay
The assay was performed by using the Luciferase Reporter Assay System (Promega, Milan, Italy), following the manufacturer’s instructions. The 3’ UTR-containing pmiR-Report was co-transfected with the Tet-O-FUW miRNA-overexpressing vector and the rtTA-expressing vector in HeLa cells. A pRL-SV40 Renilla luciferase reporter vector (Promega) was also used to quantify the transfection efficiency. Firefly luciferase luminescent signal was normalized on the Renilla luciferase signal. For each assay, a control experiment with the empty pmiR-Report vector or without overexpressing any miRNA were performed.

Dual-Fluorescent Reporter Sensor
DFRS plasmids were kindly provided by Prof. Wieland B. Huttner. Cloning strategy was performed as previously described (De Pietri Tonelli et al., 2014) using the oligos listed in Supplemental Experimental Procedures. Plasmids were transfected into control (basal) and differentiating mESCs by Lipofectamine. Fluorescence was monitored every day until 72 hr post transfection.

Lentivirus Preparation and Viral Infection
cDNAs for mAscl1, mNurr1, and 983 base pairs encompassing the pri-miRNA-34b/c cluster were cloned into Tet-O-FUW or Tet-O-FUW-Ires-GFP lentiviral vectors under the control of the tetracycline operator. Lentiviruses were packaged in HEK293T cells as previously described (Caiazzo et al., 2011) (see Supplemental Experimental Procedures for details). Gene expression was tested by qPCR, while miRNA levels were measured by TaqMan assay. Infections were performed in combination of rtTA transactivator viruses supplied with doxycycline (2 μg/mL, Clontech).

Fluorescence-Activated Cell Sorting
TH-GFP* iDA cells or mES-miR-34b/c-GFP* cells were trypsinized, washed, and sorted or analyzed with a BD FACSARia II. Cells were collected in TRI-Reagent (Sigma) for RNA extraction or in cell culture medium for subsequent amplification.

Induced Dopaminergic Neuron Generation
iDA neurons were generated from MEFs as previously described (Caiazzo et al., 2011). In brief, MEFs were infected with rtTA, Tet-O-FUW-mAscl1, Tet-O-FUW-mNurr1, and Tet-O-FUW-miR-34b/c lentiviruses in MEF medium with addition of doxycycline (2 μg/mL, Sigma). After 48 hr the medium was replaced with neuronal inducing medium (DMEM/F12, B27, and penicillin/streptomycin [all from Thermo Fisher Scientific]) containing doxycycline and cells cultured for 14 days.
**Electrophysiology Recordings**

Whole-cell patch-clamp and conventional single-unit extracellular recordings were performed according to published procedures (Guatteo et al., 2013, 2017) from visually identified TH-GFP+ cells plated on glass coverslips at 12–16 days post infection. See Supplemental Experimental Procedures for details.

**Statistical Analysis**

All statistical analyses were performed using GraphPad Prism (GraphPad Software). Significance of differences was assessed by one-way ANOVA followed by Newman-Keuls post hoc test for intergroup comparisons or by Student’s t test when cultures were compared with the corresponding control or with vehicle.

**ACCESSION NUMBERS**

The accession number for microarray data reported in this paper is GEO: GSE110270.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes Supplemental Experimental Procedures, three figures, and one table and can be found with this article online at https://doi.org/10.1016/j.stemcr.2018.02.006.

**AUTHOR CONTRIBUTIONS**


**ACKNOWLEDGMENTS**

We thank FIRB-RBIN062YH4, MERIT-RBNE08LNN4P-002, PRIN 2015SR9ASHHT_003, and Finanziamento Ricerca di Ateneo to C.P.C. for financial support. We also thank Sara Mancinelli who provided the Tet-O-FUW-Ires-GFP empty vector and Dr. Antonio Simeone for anti-LMX1B antibody. We are grateful to the FACS facility and the Integrated Microscopy Facility of the Institute of Genetics and Biophysics “Adriano Buzzati Traverso,” CNR, Naples, Italy.

Received: April 24, 2017
Revised: February 9, 2018
Accepted: February 9, 2018
Published: March 8, 2018

**REFERENCES**


