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Increased expression of miR142 and miR155 in glial and immune cells after traumatic brain injury may contribute to neuroinflammation via astrocyte activation

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INTRODUCTION

Traumatic brain injury (TBI) is a major cause of death and disability in humans, which is estimated to affect more than 50 million people worldwide every year (12,32). It involves a number of pathological alterations as a result of external damage to the brain (36,40). Numerous molecular changes following the primary brain injury contribute to the secondary brain injury, which includes brain inflammation, reactive gliosis, blood-brain barrier (BBB) dysfunction, axonal injury, progressive neuronal loss and remodeling of the extracellular matrix (ECM) (36,41,45,53). Initial brain inflammation is mediated by immune-competent cells, such as microglia and astrocytes, which produce pro-inflammatory mediators, including cytokines interleukin 1 beta (IL-1β) and tumor necrosis factor alpha (TNF-α) (49,63,64). Sustained brain inflammation leads to chronic activation of glial cells and infiltration of neutrophils, lymphocytes and macrophages in the brain parenchyma (19,49). This negatively affects the neuronal survival and promotes BBB dysfunction, further contributing to the secondary injury post-TBI. A better understanding of the molecular and cellular alterations involved in brain inflammation is crucial for the development of effective and adequate management of post-TBI neuropathology.
Brain inflammation can be regulated by microRNAs (miRNAs) (15,50), which constitute a class of small non-coding RNAs, capable of posttranscriptional regulation of gene expression networks (7). MiRNAs in mammals suppress target gene expression by directing the RNA-induced silencing complex toward the target messenger RNA (mRNA) through binding to the complementary regions in its 3′-untranslated region. MiRNAs have been shown to regulate numerous biological processes within the central nervous system both under normal and pathological conditions (26). Transcriptomic studies have revealed many deregulated miRNAs in the cerebral cortex and hippocampus in animal models for TBI (13,28,43,44,61). Among them are two miRNAs that are crucially involved in the immune response and brain inflammation: miR155 (14) and miR142 (48). These miRNAs may participate in the secondary injury post-TBI through the regulation of inflammation in the brain. Both these miRNAs have been found to be upregulated following the controlled cortical impact (CCI) injury in rats (16,17,38,54,62), but their expression in other post-TBI models has not been characterized. Furthermore, the validation of these findings and the cell-type specific characterization of miR155 and miR142 in the human post-TBI brain are lacking. Finally, circulating miRNAs in blood have been hypothesized to reflect neuropathological changes occurring after brain injury (4,13,42). Interestingly, increased expression of miR142 was observed in serum 1 day after TBI in rats (6). Therefore, the potential of these miRNAs to serve as biomarkers deserves further investigation.

We hypothesized that the inflammation-associated miR155 and miR142 are involved in the secondary brain injury and their expression in blood reflects the post-TBI pathological processes in brain. Therefore, we investigated the expression and cellular localization of miR155 and miR142 in the perilesional cortex of rats after lateral fluid percussion injury (FPI) and in the human autopsic brain post-TBI. The expression of miR155 and miR142 was analyzed in rat plasma post-TBI. Additionally, the potential of miR155 and miR142 to modulate inflammatory response in human astrocytes was investigated in vitro.

**MATERIALS AND METHODS**

The detailed description of methods is available as Supporting Materials and Methods.

**Human brain tissue**

The cases included in this study were obtained from the archives of the department of Neuropathology of the Amsterdam UMC, the Netherlands. Human postmortem brain samples (n = 3-6, who died from TBI (Table 1), were included in this study, as well as five controls (n = 5) without a history of neurological diseases. The neuropathological evaluation of human TBI samples was performed by two trained neuropathologists and assessed according to a routine protocol. TBI was confirmed, which was associated with cerebral hemorrhage, gliosis and axonal injury in all TBI samples. The protocol for detecting axonal injury included luxol fast blue-hematoxylin and eosin stainings, as well as immunohistochemistry for β-APP and neurofilament proteins. Tissue was obtained and used in accordance with the Declaration of Helsinki and the Amsterdam UMC Research Code provided by the Medical Ethics Committee. All cases were reviewed by a trained neuropathologist. Brain tissue was fixed in 10% buffered formalin and embedded in paraffin.

**Human primary cells**

Primary fetal astrocyte-enriched cell cultures were derived from human fetal brain tissue (14–20 weeks of gestation) obtained from medically induced abortions. All material was collected from donors from whom written informed consent was obtained. The detailed description of materials and methods is available as Supporting Materials and Methods.

<table>
<thead>
<tr>
<th>Sample#</th>
<th>Gender</th>
<th>Age</th>
<th>Post-TBI interval</th>
<th>Brain region</th>
<th>TBI cause</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control 1</td>
<td>F</td>
<td>27</td>
<td>–</td>
<td>Front</td>
<td>–</td>
</tr>
<tr>
<td>Control 2</td>
<td>M</td>
<td>60</td>
<td>–</td>
<td>Front</td>
<td>–</td>
</tr>
<tr>
<td>Control 3</td>
<td>F</td>
<td>35</td>
<td>–</td>
<td>Front</td>
<td>–</td>
</tr>
<tr>
<td>Control 4</td>
<td>M</td>
<td>31</td>
<td>–</td>
<td>Front</td>
<td>–</td>
</tr>
<tr>
<td>Control 5</td>
<td>M</td>
<td>67</td>
<td>–</td>
<td>Temp</td>
<td>–</td>
</tr>
<tr>
<td>TBI 6*</td>
<td>M</td>
<td>35</td>
<td>6 mon</td>
<td>Front</td>
<td>Horse kick</td>
</tr>
<tr>
<td>TBI 7</td>
<td>F</td>
<td>65</td>
<td>6 d</td>
<td>Front</td>
<td>Traffic accident</td>
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<tr>
<td>TBI 8</td>
<td>M</td>
<td>32</td>
<td>1 d</td>
<td>Temp-par</td>
<td>Traffic accident</td>
</tr>
<tr>
<td>TBI 9</td>
<td>M</td>
<td>34</td>
<td>6 d</td>
<td>Front</td>
<td>Traffic accident</td>
</tr>
<tr>
<td>TBI 10</td>
<td>M</td>
<td>20</td>
<td>5 d</td>
<td>Front</td>
<td>Traffic accident</td>
</tr>
<tr>
<td>TBI 11†</td>
<td>M</td>
<td>67</td>
<td>1 mon</td>
<td>Front-par</td>
<td>Traffic accident</td>
</tr>
<tr>
<td>TBI 12</td>
<td>M</td>
<td>52</td>
<td>38 y</td>
<td>Temp</td>
<td>Traffic accident</td>
</tr>
</tbody>
</table>

Post-mortem human brain cortex of patients post-TBI (n = 7) and post-mortem control brain cortex (n = 5) samples were used in this study; F = female; M = male; TBI = traumatic brain injury, d = day(s); mon = months; y = years; Front = frontal cortex, Temp = temporal cortex; Par = parietal cortex.

*The case represented in Figure 2.
†The case represented in Figure 5.
consent for the use of the material for research purposes was obtained by the Bloemenhove clinic, the Netherlands. Tissue was obtained in accordance with the Declaration of Helsinki and the Amsterdam UMC Research Code provided by the Medical Ethics Committee. Human primary peripheral blood mononuclear cells (PBMCs) and T cells were isolated from blood using the Macs Pan T Cell Isolation Kit (Miltenyi Biotec, Bergisch Gladbach, Germany) and human primary microglia was isolated as previously described (35).

Animals

Adult male Sprague Dawley rats (n = 20) (Envigo, Horst, the Netherlands) were housed in a controlled environment (temperature 22 ± 1 °C; humidity 50–60%; lights on from 07:00 to 19:00 h). At the time of the experiments the animals weighed 350–400 g. Water and pellet food were provided ad libitum. All animal procedures were approved by the Animal Ethics Committee of the Provincial Government of Southern Finland. All animal work was carried out in accordance with the guidelines of the European Community Council Directives 2010/63/EU.

Lateral fluid-percussion injury

Rats were randomly assigned to groups as follows: rats which received lateral FPI (n = 8), sham-operated rats (n = 5) and naïve rats (n = 3). The rats were subjected to lateral FPI as described previously (22,34). Brain injury was induced with a mean pressure of 3.18 ± 0.08 atm. Sham-operated animals received anesthesia and underwent all surgical procedures without lateral FPI. The animals were sacrificed 2 weeks post-TBI. Acute mortality within 48 h post-TBI was 15% (3/20). One rat was excluded from the analysis because of a broken dura mater after TBI.

Sample collection

Rats were anesthetized with 5% isoﬂurane and decapitated with a guillotine. The brain was dissected and the cortex (~1 to ~4 from Bregma) was snap-frozen in liquid nitrogen. The rest of the brain was fixed in 10% formalin for 3 days at 4°C, cryoprotected in 20% glycerol and stored at −80°C thereafter. Coronal sections (10 µm) were prepared from the caudal part of the brain.

Blood sampling was done according to previously described guidelines (60). Briefly, rats were anesthetized with 5% isoflurane and blood was collected from the tail vein prior to TBI and 2 weeks post-TBI. Hemolyzed samples were excluded from further analysis.

Size-exclusion chromatography (SEC) of plasma

SEC analysis was performed as previously described (8,21). Briefly, plasma samples (200 µL) from four rats (two TBI, one sham and one naïve rat) were passed through sepharose CL-2B (GE Healthcare; Uppsala, Sweden) chromatography columns. A total of 25 fractions of 0.5 mL each were collected.

Plasmids

MiRNA expressing vectors were prepared by cloning the DNA fragments encoding stem-loop pre-miRNA sequences including their ~100-200 bp flanking regions into the multiple cloning site of a pCDH-EF1a plasmid with green fluorescent protein from copepod Pontellina plumata (copGFP) as reporter (a kind gift from Dr. J. Kluiver and Dr. J. Guikema, Amsterdam UMC, Amsterdam, the Netherlands) using NheI and NotI restriction sites. The DNA fragments were amplified from human genomic DNA for hsa-mir-142 and from C. elegans genomic DNA for cel-mir-59, used as a negative control (NC).

Cell culture

The culture medium contained Dulbecco’s Modified Eagle Medium (DMEM) for HEK 293T cells, DMEM/F10 Ham (1:1) for astrocytes or Roswell Park Memorial Institute (RPMI) 1640 for THP-1 cells (Gibco/Thermo Fisher Scientific, Waltham, MA, USA), supplemented with 2 mM L-glutamine, 100 units/mL penicillin, 100 µg/mL streptomycin and 10% heat-inactivated fetal calf serum (FCS) (Gibco, Life Technologies, Grand Island, NY, USA).

A HEK 293T cell line was used to produce lentiviral particles: 6.8 µg plasmid DNA, 1.7 µg pMD2.G/VSV-G (Addgene plasmid #12259) and 3.4 µg psPAX2 (Addgene plasmid # 12260) DNA using Genius transfection reagent (Westburg, Leusden, the Netherlands) according to the manufacturer’s instructions.

The human monocytic cell line THP-1 was used to generate cell lines overexpressing hsa-mir-142 and cel-miR-39. The cells expressing copGFP were sorted on a Sony SH800S cell sorter (San Jose, CA, USA). Further, THP-1 cells were used to produce macrophage-conditioned medium (MCM). Briefly, THP-1 cells (1.5 × 10⁶ per well in 6-well plates) were differentiated into macrophage-like cells by stimulation with 80 nM phorbol 12-myristate 13-acetate (PMA; Sigma-Aldrich, St. Louis, MO, USA) for 12 h, followed by 24 h incubation with fresh medium. To induce a pro-inflammatory state, the cells were stimulated with 10 ng/mL lipopolysaccharide (LPS from E. coli O55:B5, Sigma-Aldrich, St. Louis, MO, USA) for 1 h, washed twice, incubated with fresh medium (2 mL per well) for 24 h and the supernatants were collected. The MCM was centrifuged at 1500 × g for 5 minutes and filtered through a 0.22 µm filter. Levels of TNF-α were measured in culture supernatants (MCM) using the PeliKine Compact TNF-α ELISA kit (Sanquin, Amsterdam, the Netherlands) according to the manufacturer’s instructions.

Human primary fetal astrocyte-enriched cell cultures were isolated and maintained as previously described (23). For treatment with MCM, astrocytes were seeded as 50 000 cells per well in 12-well plates, allowed to attach for 24 h and stimulated MCM/fresh medium 1:1 mix. Cells were incubated for 6 h and harvested for RNA extraction. For LPS-stimulated MCM, astrocytes were additionally treated with 100 ng/mL of Toll-like receptor (TLR) 4 antagonist LPS-rs (LPS from Rhodobacter sphaeroides; Invivogen, Toulouse, France).
**RNA extraction**

RNA isolation from rat cortex (5–30 mg) was done using phenol/chloroform extraction following the previously described protocol (39). RNA isolation from rat plasma (100 µL), human cortex (30–50 mg) and primary cells was done using the miRNeasy Mini kit (Qiagen Benelux, Venlo, the Netherlands) according to the manufacturer’s instructions. A spike-in exogenous control (5.6 × 10⁶ copies/mL of cel-miR-39-3p; miRNeasy Serum/Plasma Spike-In Control Cat. #219610, Qiagen Benelux, Venlo, the Netherlands) and carrier RNA from bacteriophage MS2 (1 µg/mL; Roche) were added during RNA isolation from plasma and primary human cells for subsequent normalization of RT-qPCR data. RNA isolation from cell culture material was done using the standard phenol/chloroform isolation procedure. The concentration and purity of RNA were determined using a Nanodrop 2000 spectrophotometer (Thermo Fisher Scientific, Wilmington, DE, USA). The Qubit microRNA Assay Kit (#Q32880, Thermo Fisher Scientific) was used to measure RNA concentration in samples obtained by SEC. The protein concentration in the fractions was determined by a Pierce BCA protein assay kit (#23225, Thermo Fisher Scientific) according to the manufacturer’s instructions.

**Reverse transcription (RT)**

For the analysis of mRNA expression, 250 ng of total RNA was reverse-transcribed using oligo-dT primers. For the analysis of miRNA expression, the TaqMan miRNA RT Kit (#4366596, Applied Biosystems, Foster City, CA, USA) was used with the following primers: rno-miR-155-5p (assay #002571), hsa-miR-155-5p (assay #002623), hsa-miR-142-3p (assay #000464), hsa-miR-142-5p (assay #002248) and cel-miR-59-3p (assay #001362). For normalization of data, the expression of U6 small nuclear RNA (RNU6) (assay #001973), cel-miR-39-3p (assay #000200) and hsa-miR-23a-3p (assay #000399) was evaluated.

**Quantitative polymerase chain reaction (qPCR)**

To evaluate mRNA expression, qPCR was performed using SYBR Green I (Roche Applied Science, Indianapolis, IN, USA). The primers used are listed in Table 2. The expression was normalized using the geometric mean of Gapdh and Tbp expression in the rat brain tissue and the geometric mean of EF1a and GAPDH in the human tissue.

MiRNA expression was evaluated by the TaqMan microRNA assay (Applied Biosystems, Foster City, CA, USA) according to the manufacturer’s instructions. The expression was normalized using the geometric mean of hsa-miR-23a-3p and cel-miR-39-3p in plasma, RNU6 in brain tissue and culture samples and to the cel-miR-39-3p across various primary cell cultures.

RT-qPCRs for all samples were performed in triplicates. The PCRs were run on the Roche LightCycler 480 (Roche Applied Science, Basel, Switzerland) with a 384-multiwell format. Data analysis was performed using LinRegPCR software (46) as previously described (24).

**Droplet digital PCR (ddPCR)**

miRNA expression in the combined adjacent SEC fractions (n = 12) was evaluated with ddPCR. Reaction mixtures were prepared as described in Bio-Rad’s ddPCR Applications Guide (Bio-Rad, http://www.bio-rad.com/) and as previously
described (61). The PCR reaction was performed in 96-well plates using the PTC-200 Thermal Cycler (MJ Research, St. Bruno, Canada). The fluorescence was measured with a QX100 Droplet Reader (Bio-Rad Laboratories Inc., Hercules, CA, USA) and analyzed with QuantaSoft software v1.7 (Bio-Rad). All samples were run in duplicates.

**In situ hybridization (ISH)**

ISH was performed as previously described (23). The oligonucleotide probes (RiboTask ApS, Odense, Denmark; Table 2) containing digoxigenin (DIG) labels, were hybridized (100 nM for miR-155-5p and 250 nM for miR-142-3p) for 1 h at 58 °C. Hybridization was detected with a sheep alkaline phosphatase (AP)-labeled anti-DIG antibody (1:1500, Roche Applied Science, Basel, Switzerland). Nitro-blue tetrazolium chloride (NBT)/5-bromo-4-chloro-3'-indolyolphosphate p-toluidine salt (BCIP) was used as chromogenic substrate for AP (1:50 diluted in NTM-T buffer: 100 mM Tris, pH 9.5; 100 mM NaCl; 50 mM MgCl₂; 0.05% Tween 20).

**ISH with immunohistochemistry**

For double-labeling after ISH, slides were incubated for 1 h at room temperature with the following primary antibodies: mouse anti-glial fibrillary acidic protein (GFAP; 1:4000, Sigma-Aldrich, St. Louis, MO, USA), mouse anti-NeuN (1:2000, MAB377, Chemicon, Temecula, CA, USA), rabbit anti-ionized calcium binding adaptor molecule 1 (Iba1; 1:2000, Wako Chemicals, Neuuss, Germany), mouse anti-human leukocyte antigen (HLA-DR/DP/DQ; 1:100, clone CR3/43 Agilent, Santa Clara, CA, USA), rabbit anti-transmembrane protein 119 (TMEM119; 1:500, #HPA051870, Sigma-Aldrich, St. Louis, MO, USA), mouse anti-CD68 (1:200, clone KP1, Dako, Glostrup, Denmark) or rabbit anti-CD8 (1:200, #7103, Dako, Glostrup, Denmark). Secondary horseradish peroxidase (HRP)-conjugated antibodies (Brightvision plus kit, ImmunoLogic, Duiven, the Netherlands) were used. The visualization of the antibody-antigen binding was done using 3-amino-9-ethylcarbazole (AEC; Sigma-Aldrich, St. Louis, MO, USA).

**Statistical analysis**

Statistical analyses were performed using Graphpad prism 5. The Mann-Whitney U-test or Kruskal-Wallis nonparametric test with Dunn’s post hoc test were used for comparisons between groups. A value of $P < 0.05$ was assumed to indicate significant difference.

**RESULTS**

**Increased expression of miR155 in the rat cortex post-TBI**

We investigated the expression of miR155 in the perilesional cortex of rats 2 weeks post-TBI in the lateral FPI model. TaqMan RT-qPCR analysis showed higher expression of miR-155-5p (Figure 1A, $P < 0.001$) in post-TBI rats as compared to control rats.

**Circulating miR-155-5p is associated with proteins, rather than extracellular vesicles (EVs)**

Next, we assessed the expression of miR155 in rat plasma 2 weeks post-TBI. RT-qPCR analysis did not show any differences in expression of miR-155-5p between post-TBI and control rats (Figure 1B). The ddPCR on plasma fractions obtained by SEC showed that miR-155-5p was enriched in the late-eluting fractions, associated with proteins (Figure 1C).

**MiR155 expression is observed in neurons, astrocytes and microglia**

Next, we investigated the cellular expression and distribution of miR-155-5p using ISH. A neuronal pattern of expression and strong hybridization signal of miR-155-5p were observed in naive (Figure 1D), sham (Figure 1E) and post-TBI rats (Figure 1F). Double-labeling showed miR-155-5p expression in neurons (Figure 1G), astrocytes (Figure 1H) and microglia (Figure 1I) located in close proximity to the lesion edge.

**Increased miR155 expression in the white matter (WM) of the human cortex post-TBI is associated with activated glial cells**

Next, the expression of miR-155-5p was analyzed in the cortex of humans post-TBI and controls using ISH. A strong hybridization signal of miR-155-5p was seen in neurons in control human cortex (Figure 2A). In the perilesional cortex, neuronal loss was present; however, a strong hybridization signal was observed in all samples in the surviving neurons as well as in cells with glial morphology (Figure 2B). In the WM of controls, a moderate expression was observed (Figure 2C). In contrast, in the WM of all post-TBI samples the ubiquitously present cells strongly expressed miR-155-5p (Figure 2D) and the morphology of these cells resembled astrocytes (Figure 2E). Double-labeling showed co-localization of miR155 with the neuronal marker NeuN (Figure 2F) in the WM and the gray matter (GM), as well as with the astrocytic marker GFAP (Figure 2G), the microglial markers Iba1 (Figure 2H) and HLA-DR/DP/DQ (Figure 2J) as well as the macrophage/monocyte marker CD68 (Figure 2K) and the marker of cytotoxic T cells, CD8 (Figure 2L).

**Pro-inflammatory conditioned medium from macrophage-like cells upregulates miR155 in human primary astrocytes**

A comparison of miR155 expression between various human primary cells demonstrated the highest expression in astrocytes, followed by T cells, PBMCs and microglia (Figure 3A).
This confirmed the observations in situ, and since miR155 was strongly expressed in activated astrocytes in the WM post-TBI, we further investigated miR155 in human astrocytes in vitro. A higher expression of inflammatory markers, including Il1b and Tnf was observed in the rat perilesional cortex (Supporting Figure S1). The activated microglia and macrophages exhibiting pro-inflammatory phenotypes may contribute to such expression profile. In
order to mimic these conditions, we incubated the astrocytes with the MCM produced by PMA-activated THP-1 cells following LPS stimulation (Figure 3B). TaqMan RT-qPCR showed higher expression of miR-155-5p in astrocytes \( (P < 0.001) \) following 6 h incubation with LPS-induced MCM compared to non-stimulated MCM (Figure 3C). This effect was the same when astrocytes were additionally treated with LPS-rs, signifying that the increase was not mediated by any carry-over of LPS to the MCM. We also observed higher expression of inflammatory markers: *IL1B* (Figure 3D), *TNF* (Figure 3E) and *PTGS2* (Figure 3F; all \( P < 0.001 \)), indicating that the LPS-stimulated MCM induced a pro-inflammatory phenotype in human astrocytes.

**Increased expression of miR142 in the rat cortex post-TBI**

We further investigated the expression of miR142 in the rat brain post-TBI. RT-qPCR analysis showed higher expression of miR-142-3p \( (P < 0.05, \text{Figure 4A}) \) and miR-142-5p \( (P < 0.05, \text{Supporting Figure S2A}) \) in the perilesional cortex of rats 2 weeks post-TBI compared to control rats. Since both strands of the mature miR142 have been reported to be functional, we compared the relative expression of miR-142-3p and miR-142-5p. The expression of the miR-142-3p strand was 200-fold higher compared to the miR-142-5p in the control and post-TBI rat brain \( (P < 0.001, \text{Supporting Figure S2B}) \). Therefore, we further focused on the miR-142-3p strand.

**Circulating miR142 is associated with EVs, rather than proteins**

Next, we assessed miR142 expression in rat plasma 2 weeks post-TBI. RT-qPCR analysis did not show any difference in expression of miR-142-3p (Figure 4B) and miR-142-5p (Supporting Figure S2C) between post-TBI and control rats. The expression of the miR-142-3p strand was

![Figure 2. Characterization of miR155 expression in the human cortex post-TBI. ISH of miR-155-5p in the GM (A, B) and WM (C, D) of the human cortex; a neuronal pattern of the miR-155-5p hybridization signal was observed in controls (A); the hybridization signal was stronger in individual cells in the the perilesional cortex (B); a moderate hybridization signal was detected in cells with glial morphology in the WM (C); a strong hybridization signal was observed in the WM post-TBI (D), arrowhead indicates a cell with astrocytic morphology shown in higher magnification in E; double-labeling showed that the miR-155-5p hybridization signal was co-localized with NeuN (F), GFAP (G, I), Iba1 (H), HLA-DP/DQ/DR (J), CD68 (K) and CD8 (L); scale bar in A = 50 μm applies to A-D; scale bars in E-L = 10 μm.](image-url)
~10-fold higher (P < 0.001; Supporting Figure S2D) as compared to the miR-142-5p strand in both control and post-TBI rat plasma. We further assessed what type of carrier the circulating miR-142-3p was associated with by performing a ddPCR analysis on plasma fractions obtained by SEC. MiR-142-3p was enriched in the early eluting fractions, associated with EVs (Figure 4C), whereas miR-142-5p was predominantly enriched in the late-eluting fractions, associated with proteins (Supporting Figure S2E).

**MiR142 expression is observed in neurons and microglia post-TBI**

Next, we investigated the cellular expression and distribution of miR-142-3p using ISH. The miR-142-3p hybridization signal was almost undetectable in the cortex of naive rats (Figure 4D), weak in sham-operated rats (Figure 4E), but strong in the perilesional cortex (Figure 4F). Double-labeling with cell-type specific markers revealed that miR-142-3p co-localized with the neuronal marker NeuN (Figure 4G) and the microglial marker Iba1 (Figure 4H), but did not co-localize with a marker of astrocytes GFAP in the perilesional cortex (Figure 4I).

**Expression of miR142 in the human brain post-TBI is associated with microglia and cells of hematopoietic origin**

We further investigated miR142 expression in the human brain. Similar to the findings in the rat cortex, the expression of the miR-142-3p strand was higher (P < 0.01) than miR-142-5p strand in the control human cortex (Supporting Figure S2F). ISH did not reveal a hybridization signal of miR-142-3p in the control human cortex in the GM
(Figure 5A) and WM (Figure 5C). A stronger miR-142-3p hybridization signal was seen in individual cells throughout the cortex of all samples post-TBI, around the lesions in the WM, as well as in the meninges and blood vessels (Figure 5B,D,E). Double-labeling of miR142 hybridization signal with cell-type specific markers revealed a weak hybridization signal located in the cells expressing NeuN in the GM (Figure 5F). The hybridization signal was not co-localized with GFAP in the GM (Figure 5G); however, a weak hybridization signal was co-localized with GFAP
in the WM astrocytes in the vicinity of lesions. The stronger hybridization signal was co-localized with the microglial markers Iba1 (Figure 5H), TMEM119 (Figure 5I) and HLA-DR/DP/DQ (Figure 5J). A strong hybridization signal co-localized with the macrophage/monocyte marker CD68 in cells with macrophage morphology (Figure 5K) as well as with CD8-positive cells inside and along the walls of blood vessels in (Figure 5L).

miR142 overexpression in the human macrophage/microglia in vitro model increases production of TNF-α

We further sought to identify whether increased expression of miR142 could evoke an inflammatory response in vitro. A comparison of miR142 expression between various human primary cells showed that miR142 was highly expressed in T cells, PBMCs and microglia, but lowly in astrocytes (Figure 6A, Supporting Figure S2G), corresponding with the observations in situ. The expression of miR-142-3p in PBMCs and microglia was comparable with the expression in the THP-1 cell line (Supporting Figure S3); therefore, we used THP-1 as a macrophage/microglia cell model to stably overexpress miR142 with the lentiviral vector (THP1mir142). THP-1 cells were differentiated into macrophage-like cells by the treatment with PMA and the pro-inflammatory state was induced by LPS stimulation (Figure 6B). TaqMan RT-qPCR showed higher expression of miR-142-3p in the THP1mir142 cells ($P < 0.001$) compared to the NC (THP1celmir59) (Figure 6C). ELISA analysis showed that the conditioned medium produced by the THP1mir142 cells following LPS stimulation had higher level of a major pro-inflammatory cytokine TNF-α (Figure 6D).

MiR142-overexpression in THP-1 cells induces a pro-inflammatory phenotype in astrocytes

We further hypothesized that the THP1mir142 MCM could potentiate the pro-inflammatory phenotype in human astrocytes (Figure 7A). The incubation of human astrocytes
miR142 and miR155 in the brain post-TBI

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Figure 6. miR142 overexpression increases production of TNF-α in THP-1 cells. (A) TaqMan RT-qPCR analysis of miR-142-3p expression in various cell types relative to cel-miR-39-3p spike-in control: miR-142-3p was strongly expressed in primary human T cells, PBMCs and microglia, but a relatively low expression was observed in astrocytes; (B) schematic representation of the in vitro experiment: THP-1 cell line stably overexpressing miR142 was converted into macrophage-like phenotype using 80 nM PMA treatment, followed by stimulation with 10 ng/mL LPS; (C) RT-qPCR analysis showed higher basal expression of miR-142-3p \((P < 0.001)\) in the THP1mir142 cells relative to the NC cells; (D) ELISA analysis showed a higher level of TNF-α \((P < 0.01)\) produced by THP1mir142 cells as compared to the NC cells; Mφ = macrophage-like; MCM = macrophage conditioned medium; NC = negative control cells (THP1celmir59); mir142 = cells overexpressing miR142 (THP1mir142); **P < 0.01, ***P < 0.001; Kruskal-Wallis with Dunn’s post-hoc test in A; Mann-Whitney U-test in C, D.

with the MCM produced by THP1mir142 cells led to a ~20% higher expression \((P < 0.05)\) of miR-155-5p as compared to the cells incubated with MCM from NC cells. However, the incubation of astrocytes with the MCM produced in THP1mir142 cells under LPS stimulation did not change miR-155-5p expression compared to the NC (Figure 7B). We further assessed the inflammatory gene expression pattern in astrocytes and found higher expression of \(IL1B\) (Figure 7C, \(P < 0.001)\), \(TNF\) (Figure 7D, \(P < 0.01)\) and \(PTGS2\) (Figure 7E, \(P < 0.001)\) in cells incubated with non-stimulated THP1mir142 MCM; as well as higher \(IL1B\) (Figure 7C, \(P < 0.001)\) and \(PTGS2\) (Figure 7E, \(P < 0.001)\), but not \(TNF\) (Figure 7D) in the cells incubated with LPS-stimulated THP1mir142 MCM.

DISCUSSION

We investigated the expression of miR155 and miR142 in the perilesional cortex and plasma of rats 2 weeks post-TBI, characterized their cell-type specific expression in the human cortex post-TBI and further studied these miRNAs in the human cells in vitro. Higher expression of both miRNAs in the brain post-TBI was associated with activated glial and infiltrating immune cells. MiR155 and
inflammatory genes were induced in astrocytes by a proinflammatory macrophage conditioned medium (MCM). This effect was further potentiated by the MCM from the cells overexpressing miR142, suggesting that miR142 may promote neuroinflammation post-TBI. These results are discussed in more detail in the following paragraphs.

Increased expression of miR155 and miR142 in the cortex post-TBI

Increased expression of miR142 and miR155 has been previously reported in the CCI post-TBI model (16,27,28,31,44,54,62). We corroborated these findings for both miRNAs in the perilesional cortex of rats after lateral FPI. The two miRNAs have different expression patterns, with miR142 virtually absent in control rat brain, but present in neurons and microglia post-TBI, and miR155 ubiquitously expressed in both control and post-TBI brain. The two miRNAs are important regulators of the innate immune response and inflammation (14,48), and their upregulation 2 weeks after the injury may indicate that they participate in the sustained inflammatory response.

Furthermore, the expression of these miRNAs in the human perilesional cortex resembles the patterns observed in the rat brain after TBI. Since the number of human TBI samples used for ISH is rather low we need to interpret these results with caution. Moreover, miR142 and miR155 expression was also validated by PCR using fresh brain tissue and in vitro by studying their expression in cultured cells. Altogether, this gives a complete and consistent picture. MiR155 is expressed only moderately by glial cells in the healthy cortex; however, a strong activation could be observed post-TBI,
especially in hypertrophic astrocytes in the WM. MiR142 could not be detected in the cortex of controls, recapitulating the observations from the rat brain. However, miR142-positive cells could be observed in the cortex post-TBI. These cells co-expressed the markers of microglia, which is in accordance with previous observations of miR142 expression in the macaque and rat microglia (9,33). Furthermore, we observed miR142 co-localization with the markers of macrophages and cytotoxic T cells, which is because of the enrichment of miR142 in the cells of hematopoietic origin (10).

The increased expression of the two miRNAs has been previously shown in several brain pathologies, all associated with strong inflammation in the brain, such as multiple sclerosis (2,20) and viral encephalitis (37) as well as temporal lobe epilepsy (3,18,24) and tuberous sclerosis complex (5,47). Therefore, since miR155 is overexpressed in activated astrocytes and miR142—in myeloid and lymphoid cells, these miRNAs may be involved in the regulation of brain inflammation after TBI.

MiR155 and miR142 may contribute to astrocyte reactivity post-TBI

Astrocytes are important mediators of the innate immune response and inflammation in the brain (11,29,51). The activation of astrocytes can be induced by pro-inflammatory signals from activated microglia (30). In a similar manner, we observed that miR155 could be induced in astrocytes by the pro-inflammatory medium from macrophage-like cells. This increase was further potentiated when the astrocytes were incubated with the medium from cells overexpressing miR142. These cells produce increased levels of TNF-α, which, upon acting on astrocytes, activates the pro-inflammatory signaling pathways associated with the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) and activator protein 1 (AP-1) transcription factors to induce miR155 expression (52,55). Indeed, the expression of pro-inflammatory genes, such as IL1b and PTGS2, was even higher in astrocytes following the stimulation by the medium from the miR142-overexpressing cells. This suggests that miR142-overexpressing cells observed in the brain post-TBI may promote a pro-inflammatory state in surrounding astrocytes. Previous evidence corroborates the idea that miR142 promotes inflammation in the brain, since miR142 knock-out eliminated inflammation and neurological impairment in the cerebellum of mice following experimental autoimmune encephalitis (33), and miR142 expression was associated with the cytotoxic macrophages in glioblastoma (65). The astrocytes activated by the pro-inflammatory milieu secrete cytokines, chemokines, prostaglandins and proteases (58,59), which further contribute to brain inflammation.

Circulating miR142 and miR155 in the rat plasma post-TBI

We did not find any biomarker potential for miR155 based on its expression in plasma post-TBI, and previous studies using human blood post-TBI support this conclusion (44,56). Although an increase in miR142 was previously observed in serum 1 day after TBI in rats (6), our experiments did not show any difference in plasma that was obtained 2 weeks post-TBI. Interestingly, we observed that miR-142-3p was enriched in the plasma fractions associated with EVs, rather than proteins. This is in accordance with previous studies (1,21) and it has been shown that only a minor portion of circulating miRNAs are associated with EVs, whereas a majority of circulating miRNAs could be found as bound to Argonaute2 protein (l). EVs, containing miRNAs and mRNAs, have been hypothesized to be actively released by the cells and participate in intercellular communication (25,57). Given the relative abundance of miR142 in the blood compared to the brain, the EV-associated miR142 from blood may target brain cells, and vice versa the EVs containing miR142 in association with brain-specific markers may be found in the blood after TBI, representing an interesting substrate for further investigation.

Conclusions

Overexpression of miR155 and miR142 in the rat and human brain post-TBI is associated with activated glial and immune cells, respectively. Human astrocytes acquire a pro-inflammatory phenotype and overexpress miR155 upon stimulation with a pro-inflammatory macrophage-conditioned medium. MiR142 expression may increase the pro-inflammatory action of macrophages, which may further potentiate astrocyte activation. We conclude that miR155 and miR142 promote neuroinflammation via astrocyte activation and may be involved in the secondary brain injury after TBI.

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CONFLICT OF INTEREST

The authors have no conflicts of interest to report. We confirm that we have read the Journal’s position on issues
involved in ethical publication and affirm that this report is consistent with those guidelines.

AUTHOR CONTRIBUTIONS
AK, NP, JK, EA\textsuperscript{V}, AP and EA conceived and designed the experiments. AK, NP, SDG, MH and JK performed the experiments. EA, EA\textsuperscript{V}, DWMB and JJA processed human brain tissue. J\textsuperscript{s}, JJA, NV, IH provided technical support. AK, EA, JDM, NP, AP and EA\textsuperscript{V} contributed to the data interpretation and manuscript preparation. All authors read, revised and approved the final manuscript.

DATA AVAILABILITY STATEMENT
The data that support the findings of this study are available from the corresponding author upon reasonable request.

REFERENCES
miR142 and miR155 in the brain post-TBI


miR142 and miR155 in the brain post-TBI

SUPPORTING INFORMATION

Additional supporting information may be found in the online version of this article at the publisher’s web site:

Figure S1. Gene expression analysis of pathological markers in the rat cortex post-TBI. Higher expression of Il1b, Tnf, Tgfβ1, Gfap (all p<0.001), as well as Pdgfrb (p<0.05) was found in the perilesional cortex of rats; *p<0.05, ***p<0.001; Mann-Whitney U-test

Figure S2. Expression miR-142-5p strand. (a) – The TaqMan RT-qPCR showed higher miR-142-5p expression (p<0.05) in the rat cortex post-TBI compared to control (naive+sham); (b) – expression of miR-142-3p strand was higher than miR-142-5p strand (p<0.001) in the rat control cortex and cortex post-TBI; (c) – miR-142-5p expression did not differ in the rat plasma post-TBI compared to control; (d) – miR-142-3p expression was higher (p<0.01) in rat plasma compared to miR-142-5p; (e) – miR-142-5p was enriched in the late-eluting plasma fractions associated with proteins; (f) – miR-142-3p was higher expressed compared to miR-142-5p in the control human cortex; (g) – miR-142-5p was highly expressed in primary human T cells, PBMCs and microglia, as well as THP-1 cell line, but not in astrocytes; *p<0.05, **p<0.01, ***p<0.001; Mann-Whitney U-test in b, c, d, f; Kruskall-Wallis with Dunn’s post-hoc test in g

Figure S3. Comparison of miR-142-3p expression between human cells. Expression of miR-142-3p in THP-1 cells was lower compared to the expression in T cells (p<0.001), however did not differ from the expression in primary microglia and PBMCs and was higher than in astrocytes (p<0.001); ***p<0.001; Mann-Whitney U-test