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Phase Separation by Entanglement of Active Polymerlike Worms

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We investigate the aggregation and phase separation of thin, living T. tubifex worms that behave as active polymers. Randomly dispersed active worms spontaneously aggregate to form compact, highly entangled blobs, a process similar to polymer phase separation, and for which we observe power-law growth kinetics. We find that the phase separation of active polymerlike worms does not occur through Ostwald ripening, but through active motion and coalescence of the phase domains. Interestingly, the growth mechanism differs from conventional growth by droplet coalescence: the diffusion constant characterizing the random motion of a worm blob is independent of its size, a phenomenon that can be explained from the fact that the active random motion arises from the worms at the surface of the blob. This leads to a fundamentally different phase-separation mechanism that may be unique to active polymers.

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Driven colloidal particles [1–3], self-propelled bots [4,5], cells [6,7], animals [8,9], and humans [10] belong to the field of active matter: interacting agents that extract energy from the environment to produce sustained motion or mechanical stresses [3,11,12]. Their collective behavior is fascinating, and the activity and interactions of the individual components give rise to highly nontrivial macroscopic phenomena [13]. Here, we investigate the spontaneous aggregation and phase separation of active-polymerlike living worms. At first sight, the phase separation of active polymers may seem similar to that of solutions [14,15] and crystallizing solids [16], for which detailed theories are available: in both cases, the aggregating particles move randomly and tend to stick together when they are in close proximity. As such, one might expect the phase separation of active particles to involve a mechanism similar to Ostwald ripening [17], where the aggregation is driven by the combined effect of diffusion of the aggregating particles and surface tension of the aggregates. However, recent work has shown that the phase separation of active particles can involve mechanisms that rely on the activity [2,3,6,13,18–20], and our results indicate that this also holds for the phase separation of active polymers.

The T. tubifex worms that we investigate are active swimmers and approximately 300 μm thick and 10–40 mm long (see Supplemental Material [21]). The thermal random motion of the worms (estimated from the Stokes-Einstein equation) is negligible compared to their active motion, so they constitute a good model system for active polymers [25–30]. When randomly distributed over a volume of water, the worms aggregate spontaneously (Fig. 1) into highly entangled “blobs.” In the remainder of this Letter, we use the term “phase separation” for the final state in which there are macroscopic domains (i.e., containing large numbers of worms) of high worm density in a space with

FIG. 1. Aggregation and phase separation of T. tubifex. (a)–(c) Snapshots of active-worm aggregation in a 25 × 25 × 2.5 cm volume at (a) t = 0, (b) 9.5, and (c) 60 min. (d) Snapshots from a 1D experiment in a square tube of 51 × 1 × 1 cm. (e)–(h) Close-ups from another experiment at (e) t = 25, (f) 28.5, (g) 29, and (h) 31.5 min, showing the coalescence of two blobs. See Supplemental Material [21] for the videos.
We use "aggregation" to refer to the growth of these domains from individual worms. The biological function of this aggregation is to minimize exposure to dissolved oxygen, high levels of which are poisonous to *tubifex* [31]. The worms cling together by entanglement, which is aided by small bristles on their bodies [32]. The aggregation is irreversible on the timescale of the experiment and the worms are submerged in water, which makes the aggregation different from motility-induced phase separation [20] and from the recently studied aggregation of *C. elegans*, which forms dynamical bundle-shaped networks kept together mainly by surface tension of the water around the worms [9]. We can analyze the aggregation of *T. tubifex* with relatively simple methods, which makes these living worms an excellent system to investigate the phase separation of active polymers.

In the experiments, we disperse a specific number of worms in a thermostated water volume and observe their aggregation in real time by recording videos (see Supplemental Material [21]). In other experiments, the water volume is a 51 × 1 × 1 cm channel in which the worms are confined in an effectively one-dimensional space (1D). Figures 1(a)–1(c) show snapshots from a typical 2D experiment, and Fig. 1(d) is from a typical 1D experiment [21]. In both geometries, as time progresses the worms form ever larger aggregates until, after about one hour, all worms are concentrated into a few large aggregates, which have the shape of a slightly flattened sphere [21], a compromise between minimum exposed surface and minimum gravitational energy. In the following, we refer to these worm aggregates as blobs.

The aggregation of the active worms seems similar to that generally observed for polymer phase separation. Hence, one might expect that the aggregation occurs through Ostwald ripening, in which larger aggregates grow at the cost of smaller ones [33]. This mechanism is driven by the reduction of the total surface tension with increasing average blob size, and we do in fact measure a finite surface tension for the worm blobs [21]. However, closer...
inspection [Figs. 1(e)–1(h)] shows that blobs of all sizes are growing, and that the growth does not occur by Ostwald ripening, but rather by the merging of smaller aggregates into larger ones. Such growth by coalescence of diffusing droplets has been investigated previously in the context of the growth of vapor-deposited thin films and of droplets on a surface [34–36].

To quantify our observations, we characterize the average blob size $\langle R \rangle$ as a function of time by taking the 2D Fourier transform of the images and determining the wave vector magnitude $q_{\text{max}}$ at which the spectrum reaches its maximum intensity [Fig. 2(a)]. Since the worm density in a blob is independent of its size [21], the size of a blob gives direct information on the number of constituent worms. With increasing time, $q_{\text{max}}^{-1}$ shifts to lower values, corresponding to an increasing average blob size $\langle R \rangle$.

Figure 2(b) shows the average blob size $q_{\text{max}}^{-1}$ determined in this way as a function of time, at several temperatures. The growth exhibits power-law behavior with $\langle R_{2D} \rangle \sim t^{1/3}$ in the 2D experiments and $\langle R_{1D} \rangle \sim t^{1/4}$ in the 1D experiments, independent of the initial worm concentration (Supplemental Material Fig. S6 [21]). In addition, we observe in the 2D experiments that the power-law growth behavior is independent of temperature.

We also investigate the motion of individual worms by recording image sequences of single, isolated worms at different temperatures (see Supplemental Material [21]). Figure 2(c) shows an example of the center-of-mass trajectory of an individual worm at various temperatures. The motion is a random walk with an effective diffusion constant that increases with temperature, as is confirmed by extracting the mean square displacement from which we retrieve the diffusion coefficient [Fig. 2(e)] [21]. By rescaling the time axis $\tau \propto D_{\text{worm}} t$ all the growth curves collapse onto a single master curve $\propto t^{1/3}$ [Fig. 2(f)], confirming that the aggregation kinetics is determined by the random motion of the worms. Unlike C. elegans which forms parallel bundles [9], the worms are not aligned but highly entangled in the blobs [Supplemental Material Fig. S4(c)], and the probability of two worms coalescing is independent of the angle between their velocities during collision [21], probably because their motion is wriggling rather than slithering.

To shed more light on the observed power-law growth kinetics, we perform computer simulations of 2D growth by coalescence of diffusing droplets, using an approach similar to that of Ref. [35] (see Supplemental Material [21] for details). We assume that the blobs are spherical and move randomly in a 2D space of size 2000 × 2000, with radius 1, which represent the individual worms, in a 2D space of size 200 × 200. At every time step, each droplet is moved in a random direction by a distance equal to $\sqrt{D_{\text{blob}}}$ where $D_{\text{blob}}$ is the diffusion constant.

Figure 3(a) shows snapshots from two simulations with two different $D_{\text{blob}}(R)$ functional dependencies, and Fig. 3(b) shows the time-dependent mass-weighted average size $\langle R \rangle = \sum_i R_i^3 / \sum_i R_i^1$, where both summations run over all the spheres present in the volume (similar results are obtained when using the number-weighted average, see Fig. S4 in the Supplemental Material [21]). If we assume that the effective diffusion constant $D_{\text{blob}}$ of a randomly moving blob depends on its radius $R$ as $D_{\text{blob}} = R^{-1}$, as is the case for blobs undergoing Brownian random motion (Stokes-Einstein equation) [33], we obtain power-law growth of the average blob size with an exponent of $\sim 0.2$ [purple circles in Fig. 3(b)]. This exponent is much smaller than the experimentally observed value of $\sim 0.3$ [Fig. 2(f)]. It may be noted that the exponent obtained from the simulation is different from that in Ref. [35] because, in our case, we have conservation of total mass, and so depletion in the space between the blobs.

We believe that the discrepancy between the simulated and experimentally observed power-law exponents is due...
exerted by $N$ worms is $F(t) = \sum_i f_i(t)$, and assuming zero cross-correlation between the individual worm forces, we have $\langle F(t) \rangle^2 = \sum_i (f_i(t))^2 = N f_0^2$, so the rms magnitude of the total force is $\sqrt{N} f_0$, and $\langle F(t) \cdot F(0) \rangle = N f_0^2 e^{-t/\tau}$. Since the worms inside the entangled blob are effectively immobilized, only the worms at the outer surface of a blob contribute force. The number of worms at the surface of a blob of radius $R_{blob}$ is $N \sim R_{blob}^2$, so the total random force exerted by these $N$ worms has a rms magnitude $\sqrt{N} f_0 \sim R_{blob} f_0$ and a correlation time $\tau$. The drag force on the blob as a function of speed $v$ is given by Stokes’s law, $F_{\text{drag}} \sim \eta R_{blob} v$. To obtain the steady-state speed $v$, we equate the driving and drag forces and obtain a velocity $v$ that is independent of the blob radius $R_{blob}$. The random walk of a blob occurs by random steps (of duration $\sim \tau$) in which it has approximately the steady-state velocity $v$ (independent of size), and so the blob-diffusion constant is independent of blob size. It might be that worms at the bottom of a blob have slightly less activity than the ones on the sides and on the top, but this would not change the essentials of the scaling argument: the blob shape is to a good approximation independent of size [Supplemental Material Fig. S5(a)], so the fraction of surface worms located at the bottom surface is approximately constant, and taking their potentially reduced activity into account would only give rise to a size-independent prefactor in the scaling expression for the total force exerted by the worms.

Again simulating the blob growth, but now using a size-independent blob-diffusion constant ($D_{blob} = 1$), we obtain a growth exponent of $\sim 0.3$ [red points and line in Fig. 3(b)], in good agreement with the experimentally observed value. Thus, a size-independent diffusion constant for the random motion of the blobs explains the observed power-law growth of the worm blobs (Figs. 2 and 3), at least in the 2D experiments. In the 1D experiments, the situation is complicated by the fact that blob motion slows down when the blob size becomes comparable to the channel width, possibly due to friction at the glass-worm surface. This makes quantitative analysis of the blob diffusion difficult. However, the fact that, in this case, the diffusion constant does decrease with $R_{blob}$ and that at the same time the growth exponent is closer to that predicted by the conventional droplet-coalescence model, does provide a qualitative confirmation of the above ideas.

The coalescence of blobs can be modeled by the Smoluchowski aggregation equation, but here we give a scaling argument for the dynamics. We assume stochastic coalescence of the blobs and a size-independent blob-diffusion constant. We characterize the blob-size distribution by a single blob radius $R(t)$ that increases with time and we start the system with a mass per unit area $\sigma$. For worms of unit mass density, the mass of a single blob is $M_{blob} \sim R^3$. The number density of blobs in the plane is $n(t)$ such that $n(t) R^3 = \sigma$, so the separation $a(t)$ between

FIG. 4. Blob diffusion. Effective diffusion constant as a function of average blob size ($R_{blob}$) at $T = 20^\circ$C as determined from the slopes of experimental MSDs (see Supplemental Material [21]). The error bars are mostly due to sample-to-sample variability. The purple line shows the expected scaling for particles undergoing Brownian random motion ($D_{blob} \sim (R_{blob})^{-1}$). The experimental data (blue symbols) indicate a diffusion constant independent of blob size (dotted line). (Lower insets) Photographs of blobs of different sizes from which we measured the diffusion coefficient.
aggregates varies as $a(t) \sim 1/\sqrt{n(t)} = \sqrt{R^3(t)/\sigma}$. The blobs diffuse and the time for a blob to find a neighbor and coalesce with it is given by $a^2 \sim Dt$ (with $D$ the diffusion constant). Thus, in time $t = cR^3$ (with $c$ a proportionality constant) the mass of a blob doubles since we have just coalesced two neighbors. Hence, the evolution of the characteristic blob mass obeys the scaling relation $[t,M_{\text{blob}}] \rightarrow [t + cR^3, 2M_{\text{blob}}]$; thus, we have $t \sim cR^3 \approx k t$, with $k = O(1)$, for all $t$, and hence $t \sim cR^3$ and so $R \sim t^{1/3}$, in excellent agreement with the observed value [Figs. 2(f) and 3(b)]. Repeating the argument for a diffusion constant varying as $D \sim 1/R$ implies that $R(t) \sim t^{1/4}$.

To conclude, the active motion of tubifex worms leads to a phase-separation mechanism that is different from that of normal polymers, and that seems to be due only to the active nature of the living worms and to the immobilization of the worms at the inside of the entangled blobs; it may therefore be a generic phenomenon, also occurring in other types of polymeric active matter with entanglement interactions. We hope that the results presented here will stimulate further experimental and theoretical work in this direction.

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