Formation of a tumor-supportive microenvironment in chronic lymphocytic leukemia
Addressing the reciprocal interactions in the CLL—T cell—macrophage triad
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MACROPHAGES CONFER SURVIVAL SIGNALS VIA CCR1-DEPENDENT TRANSLATIONAL MCL-1 INDUCTION IN CHRONIC LYMPHOCYTIC LEUKEMIA

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ABSTRACT

Protective interactions with bystander cells in micro-environmental niches such as lymph nodes (LNs) contribute to survival and therapy resistance of chronic lymphocytic leukemia (CLL) cells. This is caused by a shift in expression of BCL-2 family members. Pro-survival proteins BCL-X\textsubscript{L}, BFL-1, and MCL-1 are upregulated by LN-residing T cells through CD40L interaction, presumably via NF-κB signaling. Macrophages also reside in the LN, and are assumed to provide important supportive functions for CLL cells. However, if and how macrophages are able to induce survival is incompletely known.

We first established that macrophages induced survival due to an exclusive upregulation of MCL-1. Next, we investigated the mechanism underlying MCL-1 induction by macrophages in comparison with CD40L. Genome-wide expression profiling of in vitro macrophage- and CD40L-stimulated CLL cells indicated activation of the PI3K-AKT-mTOR pathway, which was confirmed in ex vivo CLL LN material. Inhibition of PI3K-AKT-mTOR signaling abrogated MCL-1 upregulation and survival by macrophages as well as CD40 stimulation. MCL-1 can be regulated at multiple levels, and we established that AKT leads to increased MCL-1 translation, but does not affect MCL-1 transcription or protein stabilization. Furthermore, among macrophage-secreted factors that could activate AKT, we found that induction of MCL-1 and survival critically depended on C-C Motif Chemokine Receptor-1 (CCR1).

In conclusion, this study indicates that two distinct micro-environmental factors, CD40L and macrophages, signal via CCR1 to induce AKT activation resulting in translational stabilization of MCL-1, and hence can contribute to CLL cell survival.
INTRODUCTION

Chronic lymphocytic leukemia (CLL) is characterized by accumulation of monoclonal B cells in peripheral blood, lymph nodes (LNs) and the bone marrow. Interactions with bystander cells such as stromal cells, T cells or macrophages (MΦs) in the LN provide CLL cells with a survival benefit and resistance to chemotherapy, due to changes in the apoptotic balance in CLL cells\(^1\). The important role of MΦs was very recently shown in MΦ depletion experiments in the TCL1 CLL mouse model, in which a better overall survival was observed\(^2\).

With respect to relevant survival factors, we have previously shown that the effects of LN-residing T cells on CLL cells are largely governed by CD40L interaction, as CLL cells stimulated by CD40L and T cells have similar gene expression and apoptotic profiles\(^3\). Factors from monocyte-derived nurse-like cells that have been described to induce survival include CXCL12\(^4\), APRIL, and BAFF. These latter two factors are reported to induce NF-κB activation\(^5\). Using several complementary approaches, we however found negligible effects of APRIL in MΦ-mediated survival\(^6\), implying that other MΦ-factors must be involved.

Concerning the change in apoptotic balance, our group and others have previously shown increased expression of pro-survival B-cell lymphoma 2 (BCL-2) family members in CLL cells isolated from LNs\(^7\) as well as in CLL cells stimulated with T cell factor CD40L\(^3, 8-10\). Clinically, such changes in apoptosis regulation correlate with worse prognosis and resistance to chemotherapy, as several groups have shown for pro-survival proteins BCL2-related protein A1 (BFL-1) and B-cell lymphoma-extra large (BCL-X\(_L\))\(^11, 12\), as well as Induced myeloid leukemia cell differentiation protein (MCL-1) levels\(^13-16\). The effects of monocyte-derived cells such as MΦs on the apoptotic balance are less well studied.

The negative prognostic impact of MΦs in CLL\(^2\) and the fact that their extracellular and intracellular signaling events towards CLL cells are unknown, suggest that unraveling these pathways can contribute to development of new therapies. We therefore studied the effects of both MΦs and CD40L on CLL cell survival and identified chemokine receptor CCR1 as an important mediator of MΦ-induced CLL cell survival. Secondly, we found that within the CLL cell, both MΦs and CD40L increase AKT-mTOR dependent translation of MCL-1 protein.

RESULTS

T cells and MΦs induce CLL survival by changing the apoptotic balance

As we have shown previously that stimulation of CLL cells via CD40 almost fully mimics the effects of activated T cells on CLL\(^3\), we used NIH-3T3 cells transfected with CD40L (3T40 cells) as a model for the interaction with T cells. We also generated M1 and M2 differentiated macrophages (MΦs) from monocytes isolated from healthy donors by differentiation with IFN-γ (M1) or IL-4 (M2). Both types of MΦs and 3T40 cells increased survival of CLL cells after 72h co-culture (Figure 1a).

We analyzed 72h stimulated CLL cells for anti-apoptotic proteins MCL-1, BCL-X\(_L\), BFL-1 and BCL-2 using western blot. MCL-1 was upregulated by both MΦ types and 3T40 stimulation, while BCL-X\(_L\) and BFL-1 were only upregulated by 3T40 stimulation, consistent with activation of NF-κB by 3T40 cells\(^10\) (Figure 1b, Figure S1 for size markers).
Figure 1: Mφs and CD40L induce CLL survival by changing the apoptotic balance. Confluent feeder layers of macrophages (Mφ) and non-dividing CD40L-overexpressing fibroblasts (3T40) were generated as described under methods. a. CLL cells were co-cultured on the indicated feeder layers or without feeder layer (ctr) for 72h after which survival was measured by Dioc6-PI staining. The percentage of viable cells was defined as Dioc6 positive cells. Each point represents one stimulation of a CLL sample. Shown are mean ± s.e.m. b. After 72h co-culture as in Figure 1a, protein lysates of CLL cells were probed by western blot for the levels of pro-survival BCL-2 family members MCL-1, BCL-X<sub>L</sub>, BFL-1 and BCL-2. β-actin was used as a loading control. Data shown is representative of at least N=6. To improve clarity, western blot images were cropped in this and following figures (upper panel). The levels of each
To verify the relevance of MCL-1 upregulation in the observed survival effect of Mφ, siRNA interference using Amaza nucleofection was applied, prior to co-culture on M1 Mφs (Figure 1c). The Mφ-mediated survival effect was largely reverted after MCL-1 knockdown. Thus, in the Mφ setting, CLL cells depend on the upregulation of MCL-1 for their survival.

The expression of MCL-1 in CLL LNs and the presence of Mφs were verified by immunohistochemical staining. These stainings indicated that MCL-1 is present at significant levels in LN-residing CLL cells and that the LN is interspersed with Mφs. (Figure 1d, and Figure S2 for single channel images).

MCL-1 induction depends on PI3K-AKT-mTOR signaling after both CD40L and Mφ stimulation

To identify which intracellular pathway was responsible for the observed MCL-1 upregulation, genome-wide expression profiling (using microarrays) of stimulated CLL cells was performed. A comparison of the log-fold changes of the expression levels of each gene in stimulated cells versus the control condition showed a large overlap between regulated genes in M1 and M2 stimulated samples (Figure 2a; R²=0.72). When comparing either M1 or M2 to 3T40 stimulated samples, there was more discordance and only limited overlap (R²=0.03 and 0.00 respectively).

As both CD40L, M1, and M2 stimulation induced MCL-1 (Figure 1b), we hypothesized that the same upstream regulator was responsible for its induction by all stimuli. Using Ingenuity Pathway Analysis, potential upstream regulators for each condition were determined and these upstream regulators were marked in a scatterplot (Figure 2b). Interferon (IFN)α and phosphoinositide 3-kinase (PI3K), the upstream regulator of the PI3K/V-Akt Murine Thymoma Viral Oncogene Homolog (AKT)/mammalian target of rapamycin (mTOR) pathway, were predicted to be activated in all three conditions. These data were confirmed in a CAMERA pathway analysis using the Molecular Signatures Database v5.1 (MSigDB, Broad institute) (data not shown). Of note, neither M1 nor M2 Mφs expressed CD40L as determined by flow cytometry (data not shown), indicating that another factor is responsible for the observed Mφ effect. As M1 and M2-stimulated CLL cells had highly similar gene expression and functional profiles (Figure 1a-b and Figure 2a), we used the more tightly adherent M1 Mφs for further experiments (hereafter called Mφ).

Protein lysates of CLL cells were probed for MCL-1 levels by western blot to verify the knockdown. β-actin was used as a loading control. Data shown is representative of N=5. (upper panel). Protein were quantified for 6 patients using densitometry and calculated as the protein signal relative to the Actin loading control. (lower panel). A.U. denotes arbitrary units. c. CLL cells were transfected with one of two siRNAs directed against MCL-1 (siMCL-1) or a control (sictr) or not transfected (-) before co-culture on Mφs or without feeder layer (ctr). After 72h, the viability of CLL cells was determined as in Figure 1a. Each line represents one patient sample. Transfection efficiency using a GFP control plasmid has previously been determined to be approximately 50% (data not shown). (upper panel). 

β-actin was used as a loading control. Data shown is representative of N=5. (lower panel). d. Paraffin embedded lymph node slides from CLL patients were immunohistochemically stained for Mφ marker CD68 and nuclear counterstain Methyl Green (left panel), or MCL-1, B cell marker CD20, and nuclear counterstain DAPI (right panel). Data shown is representative of N=6.
Figure 2: MCL-1 induction depends on PI3K-AKT-mTOR signaling after both CD40L and Mφ stimulation. 

a. CLL cells were co-cultured as in Figure 1 (N=3 paired samples for each condition) for 16h and RNA from CD5/CD19 FACS-sorted CLL cells (>99% purity) was subjected to microarray analysis. Next, for each mRNA, the log fold change (LogFC) relative to control condition was calculated and plotted. The goodness of fit for the linear regression models (R²) is indicated.

b. A core analysis in the Ingenuity Pathway Analysis (IPA) software was performed with signatures created from the microarray data based on cut-off values logFC > 0.8 and P value < 0.01 to identify putative upstream regulators common to all three stimuli. The Z-score of candidate upstream regulators was visualized in scatterplots, in which upstream regulators with a Z-score over 2.0 were defined as activated, in line with IPA specifications. Several potentially relevant activated regulators are indicated by arrows.

c. CLL cells were serum starved for 3h and subsequently co-cultured as in Figure 1 on indicated feeder layers for indicated time points and the phosphorylation of AKT was determined by western blot.
Furthermore, considering that IFNα is not secreted by 3T40 cells, we decided to focus on the AKT pathway activation.

In accordance with the microarray data, we found that both 3T40 and Mφs induced AKT phosphorylation (Figure 2c). To test for sustained functional activation of AKT, phosphorylation of its downstream target GSK3β was probed after 6h, showing phosphorylation after both stimuli (Figure 2d). The phosphorylation of GSK3β was furthermore found in FACS sorted CLL cells from LN material, indicating active AKT signaling in vivo (Figure S3).

To test whether MCL-1 induction depended on AKT-mTOR signaling, we used two pharmacological inhibitors of mTOR; AZD8055, which competes for the ATP binding pocket and the allosteric inhibitor rapamycin. Both inhibitors were able to completely revert the CLL survival effect conferred by Mφs. However, they did not revert CD40-induced survival, likely due to the concurrent upregulation of anti-apoptotic proteins BFL-1 and BCL-XL. AZD8055 reduced MCL-1 levels in both Mφ and 3T40 stimulated CLL cells. Rapamycin on the other hand showed only an effect in CD40L-stimulated CLL cells (Figure 2e). Rapamycin, as opposed to AZD8055, is unable to inhibit the mTORC2 mediated positive feedback loop and mTOR mediated 4E-BP phosphorylation has been shown to be resistant to rapamycin treatment. Next, CAL101 (idelalisib), which inhibits PI3Kδ, was used to decrease AKT activation. Idelalisib treatment showed a reduction in MCL-1 protein for both conditions, but only reduced Mφ- (and not 3T40-) mediated CLL survival, (Figure 2f).

Collectively, these data suggest that the observed induction of MCL-1 after CD40L or Mφ stimulation depends on PI3K-AKT-mTOR signaling.

MCL-1 is induced on the translational level

Protein stabilization via AKT-dependent GSK3β phosphorylation has been attributed an important role in MCL-1 induction. We therefore measured MCL-1 turnover after addition of translation inhibitor cycloheximide. However, these analyses indicated similar stability of MCL-1 protein before and after co-culture with Mφ or 3T40 cells (Figure 3a). Furthermore, treatment with CHIR99021, a selective GSK3 inhibitor, induced no changes (shown blot is representative of N=4). β-actin was used as a loading control. After resting for 3h, CLL samples were co-cultured as in Figure 1 on indicated feeder layers for 6h and the activation of AKT signaling was determined by western blotting for phosphorylated GSK3β (blot representative of N=4). CLL samples were co-cultured as in Figure 1 in the presence or absence of mTOR inhibitors AZD8055 (500nM) or rapamycin (1μM) and survival was determined as in Figure 1a. Shown are mean ± s.e.m. for N=9 (untreated and RAPA) or N=3 (AZD) samples. Similarly treated CLL samples were subjected to western blot and probed for MCL-1. To exclude caspase-mediated MCL-1 breakdown, 5μM Q-VD-OPh was added to all samples during culture when analyzed for MCL-1 levels by western blotting. β-actin was used as a loading control. Western blot representative for N=6. CLL samples were co-cultured as in Figure 1 in the presence or absence of 1μM PI3K inhibitor CAL101 (idelalisib) and survival was determined as in Figure 1a. Shown are mean ± s.e.m. for N=7 (upper panel). Similarly treated CLL samples were subjected to western blot and probed for MCL-1, as in Fig2e. Western blot representative for N=4.
Figure 3: MCL-1 is regulated on translational level. a. After co-culture for 72h, turnover of MCL-1 in CLL cells was determined by quantifying levels at different time points after addition of 25μg/mL translation inhibitor cycloheximide, and compared to unstimulated samples (N=6). Each point represents mean ± s.e.m. and regression lines were calculated using a best-fit exponential decay model fitted with the formula $Y = Y_0 * e^{-k*X}$, in which $Y_0$ was set to 1. The best fit k value was not significantly different between lines (upper panel). A representative turnover western blot for 1 CLL sample is shown (lower panel). Note the higher MCL-1 starting levels in the stimulated samples. A.U. denotes arbitrary units. 

b. Unstimulated CLL samples were pre-treated or not treated for 1h with 3μM GSK3 inhibitor CHIR99021 before MCL-1 turnover was determined as in Figure 3a. To exclude caspase-mediated MCL-1 breakdown, 5μM Q-VD-OPh was added to all samples during culture when analyzed for MCL-1 levels by western blotting. A representative CLL sample of N=3 is shown.

c. The translational efficiency of MCL-1 mRNA was determined after 48h co-culture by calculating the percentage of MCL-1 mRNA bound in polysomes, by performing qPCR on sucrose gradient-separated non-polysomal and polysomal fractions (see methods and Figure S4a). An MCL-1 and 18S qPCR were performed on pooled samples (polysomal/non-polysomal) and the relative amount of polysomal bound mRNA ± s.e.m. (N=4 independent experiments) compared to the control condition (fold induction) was calculated for MCL-1 or 18S RNA.

d. CLL samples used in Figure 1b were again subjected to western blot and probed for phosphorylation of translation initiation factors and phosphorylation of ribosomal protein S6, the latter is indicative of active translation. β-actin was used as a loading control. Note that while the anti-p-4E-BP antibody was expected to detect only the slowest migrating form of 4E-BP, the non- and partially phosphorylated forms are also detected, most likely due to cross-reactivity to the unphosphorylated site.

e. The western blot presented in Figure 2c was probed for ERK phosphorylation (shown blot is representative of N=4).

f. Protein lysates from 3 CLL LN samples were analyzed by western blot for the phosphorylation of ribosomal protein S6 as a measure for active translation. An unmatched unstimulated and 3T40-stimulated CLL sample were included for comparison.
in MCL-1 half-life while the upregulation of β-catenin, another GSK3β substrate, indicated effective GSK3β inhibition (Figure 3b).

As AKT is furthermore involved in the formation of the translation initiation complex, we evaluated changes in translation of MCL-1 mRNA, by measuring the amount of mRNA bound in actively translating ribosome chains, or polysomes. Sucrose gradient centrifugation of cell lysates was used to separate polysomal and non-polysomal mRNA and determine the fraction of polysomal MCL-1 mRNA. These analyses showed an increase in polysomal bound MCL-1 mRNA of approximately 2-fold in 3T40 and 3.5-fold in Mφ-stimulated samples, indicating an increase in MCL-1 mRNA translation. The distribution of 18S rRNA, in contrast, was not altered upon exposure to 3T40 cells or Mφs, implying that the number of actively translating ribosomes did not change (Figure 3c and S4a for a representative BioAnalyzer profile).

The formation of polysomes is initiated after the phosphorylation of several translation initiation factors that are part of the Eukaryotic translation initiation factor 4 (eIF4) complex. Association of this complex with mRNA depends on the cap-binding factor eIF4E, which is inhibited by Eukaryotic translation initiation factor 4E-binding protein (4E-BP). The mTOR dependent phosphorylation of 4E-BP releases eIF4E to increase translation initiation, while ERK dependent phosphorylation of eIF4E enhances formation of the eIF4 complex. Furthermore, the phosphorylation of ribosomal protein S6, that is part of the 40S ribosome, has been implicated in translation initiation. Thus, the phosphorylation of eIF4E, 4E-BP, and S6 correlate with an increase in translation.

We therefore analyzed the phosphorylation status of these proteins and found that stimulation with either Mφs or 3T40 cells consistently resulted in upregulation of phosphorylation of 4E-BP and S6. Moreover, 3T40 stimulation also increased eIF4E phosphorylation (Figure 3d). In accordance, ERK was phosphorylated exclusively by 3T40 and not by Mφs (Figure 3e). As PIM1 and PIM2 kinases have also been described to correlate with 4E-BP phosphorylation, we analyzed their expression levels, but found no difference in either PIM1 or PIM2 expression upon stimulation (Figure S4b).

Lastly, we used protein lysates from FACS-sorted CLL cells isolated from LNs to investigate whether translation was activated in vivo and found phosphorylated S6 in all investigated LNs (Figure 3f). Due to a high background signal in the CLL LN lysates, we were unable to detect p-4E-BP (not shown). The upregulation of MCL-1 in CLL on the same LN samples was previously shown by our group.

In line with our findings of translational stabilization, we found no transcriptional induction of MCL-1 when analyzing the microarray data sets generated for Figure 2 by any stimulation, in contrast to BCL-XL and BFL-1, known targets of NF-κB (Figure S4c). In agreement, after Mφ co-culture, no NF-κB subunit translocation occurred (Figure S4d), and no NF-κB DNA binding activity could be detected in CLL cells using ELISA (Figure S4e).

In summary, 3T40 cells nor Mφs led to transcriptional upregulation or post-translational stabilization of MCL-1, but both stimuli induced translation of MCL-1 mRNA by activation of the initiation complex. This translational activation signature was also present in ex vivo LN samples.
MCL-1 upregulation and CLL cell survival are CCR1-dependent

We then aimed to identify the Mφ-produced factor responsible for the observed survival increase and MCL-1 upregulation via AKT. Various Mφ-secreted factors have been described to induce AKT signaling, among which are growth factors, integrin signals, and chemokines. As trans-well culturing experiments showed Mφ-mediated survival in the non-contact setting, we focused on soluble factors for their ability to induce CLL cell survival, while not excluding contact-dependent factors. Using several recombinant growth factors, we could not recapitulate the Mφ survival effect. To test the effect of integrin signaling, CLL cells were cultured on plates pre-coated with fibronectin or VCAM, that stimulate several integrin receptors. As integrin signaling has been described to affect growth factor signaling, these stimulations were performed with or without a combination of the growth factors used in Figure 4a. Again, none of these stimuli induced survival in CLL cells. Furthermore, inhibition of integrin signaling in Mφ-stimulated samples did not affect survival (data not shown). Lastly, highly specific chemokine inhibitors for CCR1, CCR2, and CCR5 were tested in the context of Mφ-stimulated CLL cells. Of these, inhibition of only CCR1 lead to a complete abrogation of the Mφ-induced survival effect. This survival reduction was not the result of non-specific cytotoxic effects, as unstimulated cell survival was not affected by CCR1 inhibition. We verified the effect of CCR1 inhibition in 9 CLL samples, and moreover included 3T40-stimulated CLL cells. Again, CCR1 inhibition completely negated the Mφ-mediated CLL cell survival, whereas it had no effect on 3T40-mediated survival. The absence of an effect in the context of 3T40 stimulation likely results from the concurrent upregulation of BCL-XL and BFL-1 after 3T40 stimulation.

We then investigated whether the reduction in survival was mediated via MCL-1 and found that in both Mφ and 3T40-stimulated CLL cells, CCR1 inhibition led to a strong reduction in MCL-1 protein levels. In addition, the phosphorylation of 4E-BP was reduced after CCR1 inhibition, indicating involvement of AKT-mTOR signaling in CCR1-mediated MCL-1 induction.

As several chemokine ligands have been described to signal via CCR1, we tested if a combination of rhCCL3, CCL5 and CCL23 could mimic the Mφ survival effect. As chemokine signaling can be dependent on presentation via heparin sulphate proteoglycans (HSPGs), these experiments were also performed using feeder layers of VCAM or NIH-3T3 cells, that express HSPGs. However, no survival benefit of the recombinant proteins was observed, suggesting that the relevant chemokine acts in conjunction with a second Mφ-secreted factor.

Altogether, these data indicate a dependence on CCR1 in both Mφ- and CD40L-mediated survival.

DISCUSSION

In this article, we investigated how two important micro-environmental signals, CD40L and macrophages (Mφs), lead to upregulation of key anti-apoptotic protein MCL-1 in primary leukemic cells, studying both extracellular and intracellular factors. In contrast to our starting assumption that MCL-1 in CLL cells is mostly post-translationally regulated,
**Figure 4:** MCL-1 upregulation and CLL cell survival are CCR1 dependent. 

**a.** CLL samples were cultured in the presence of 25ng/mL Epidermal growth factor (EGF), 20ng/mL Basic fibroblast growth factor (FGF2), 10ng/mL Platelet-derived growth factor (PDGF), 50ng/mL Stem cell factor (SCF), 25ng/mL Fms-related tyrosine kinase 3 ligand (FLT3L), 10ng/mL Vascular endothelial growth factor (VEGF), or nothing or on Mφ for 72h and survival was measured. Shown are mean ± s.e.m. for N=6 CLL samples. 

**b.** CLL cells were cultured without feeder layer, or on fibronectin coated plates (FN), on VCAM coated plates (VCAM), or on Mφ for 72h in the presence of absence of a combination of the growth factors (rhGFs) used in Figure 5a. Survival is shown as mean ± s.e.m. for N=3 CLL samples. 

**c.** CLL cells were co-cultured on Mφ or without feeder layer (ctr) for 72h in the presence of absence of specific CCR inhibitors against CCR1 (1µg/mL BX471), CCR2 (100ng/mL INCB3284), or CCR5 (1µM Maraviroc). Next, survival was measured as in Figure 1a. Shown are mean ± s.e.m. for N=3 CLL samples. 

**d.** CLL cells were co-cultured on indicated feeder layers or without feeder layer (ctr) for 72h in the presence of absence of CCR1 inhibitor BX471. Next, survival was measured as in Figure 1a. Shown are mean ± s.e.m. for N=9 CLL samples. 

**e.** After 72h co-culture as in Figure 3d, protein lysates of CLL cells were probed by western blot for MCL-1 and 4E-BP. Data shown for 1 patient is representative of N=3. 

**f.** CLL cells were co-cultured on indicated feeder layers for 72h or cultured with a combination of CCR1-binding chemokines CCL3, 5, and 23 on non-coated or VCAM-coated plates or on 3T3 cells. Next, survival was measured as in Figure 1a. Shown are mean ± s.e.m. for N=3 CLL samples comparing each feeder layer with to without chemokines or Mφ or 3T40 to control.
we determined that both Mφ and CD40L stimulation induced MCL-1 via AKT-dependent activation of the translation initiation complex (Figure 5). Several observations support the notion that activation of the initiation complex results in a specific translational increase of MCL-1. First, mRNAs with long GC-rich highly structured 5' UTRs, such as the mRNA from MCL-1, are particularly sensitive to translational regulation. Second, in the Tsc2(+/−) Eμ-Myc mouse model, which has constitutive Akt activation, translationally induced Mcl-1 appeared to be the main determinant of mTOR dependent survival. Third, the survival effect that resulted from expression of constitutively active AKT (myr-AKT) in CLL cells, could be reverted by downregulation of MCL-1 by siRNAs. Fourth, overexpression of a phosphomimetic S209D eIF4E variant in cancer cell lines selectively increases the translation of a limited number of proteins, among which is MCL-1. Fifth, inhibition of the translation complex following glucose deprivation sensitizes cells to death receptor-mediated apoptosis as a result of translational MCL-1 downregulation. Modification(s) of translation factors could thus be a micro-environmental regulatory mechanism inducing specifically pro-survival proteins such as MCL-1.

In addition to translational control, several reports describe post-translational stabilization of MCL-1 that is mediated via GSK3β. Other reports show an increase in MCL-1 transcription in CLL after co-culture with mesenchymal stromal cells, or after STAT activation with cytokines. Our observation of translational regulation of MCL-1 independent of GSK3β adds to the wide spectrum of MCL-1 control. These distinct mechanisms of MCL-1 control are not mutually exclusive and their relevance will probably depend on the cellular context, and apparently post-translational stabilization is not the dominant mechanism in primary CLL cells.

Furthermore, MCL-1 upregulation after CD40L or Mφ stimulation was independent of NF-κB activation (Figure S4d-e). In agreement, no consensus NF-κB binding sites can be found in the MCL-1 promotor (Figure S4f). Although several publications report a correlation between NF-κB activation and MCL-1 levels in CLL cells in response to other stimuli, this may very well be due to autocrine interleukin-mediated MCL-1 upregulating signals that respond to NF-κB activation, or alternatively, MCL-1 and NF-κB are both regulated by an upstream activator, as is the case for CD40 signals. In summary, it appears that under certain conditions NF-κB activation can indirectly induce MCL-1 transcription.

We have found that the responsible Mφ factor for CLL survival is a chemokine that signals via CCR1, while excluding growth factors, and integrins. In this light, we have moreover excluded the involvement of A proliferation inducing ligand (APRIL) and CD40L (data not shown). Chemokines that can signal via CCR1 include CCL3, 4, 5, 7, 14, 15, 16, and 23. Interestingly, the reduction in MCL-1 levels after CCR1 inhibition of 3T40-stimulated CLL samples suggests that this upregulation of MCL-1 depends on (indirect) chemokine-mediated signals. Indeed, we recently observed production of several chemokines such as CCL3, 5 and 7 by CLL cells after 3T40 stimulation in support of a model in which MCL-1 upregulation after 3T40 stimulation depends on autocrine chemokine stimulation of CLL cells (Figure 5).

In conclusion, our data indicate that two model systems of important micro-environmental stimuli (CD40L and Mφs) are able to induce survival in primary CLL cells by
Figure 5: Schematic model for the chemokine-mediated micro-environmental translational regulation of MCL-1. Chemokines secreted directly by macrophages or in an autocrine fashion by CLL cells after CD40 stimulation lead to triggering of the CCR1 receptor. This triggering induces AKT signaling and subsequent induction of mTOR dependent 4E-BP phosphorylation and release of its inhibited binding partner eIF4E. Concurrently, the induction of ERK signaling by CD40 stimulation results in the phosphorylation of eIF4E. These processes facilitate the recruitment of eIF4E to the 5’ mRNA cap and formation of the ribosome, which results in the active translation of mRNA of pro-survival proteins like MCL-1.

upregulating MCL-1 translation via AKT signaling. This MCL-1 upregulation resulted from CCR1-mediated signals after both stimuli. These insights may be applicable in designing new treatment strategies for CLL.

MATERIAL AND METHODS

CLL and healthy donor material and isolation

Patient material was obtained from CLL patients, after written informed consent, during routine follow-up or diagnostic procedures at the Academic Medical Center, Amsterdam, the Netherlands. The studies were approved by our Ethical Review Board and conducted...
in agreement with the Helsinki Declaration of 1975, revised in 1983. Peripheral Blood mononuclear cells (PBMCs) of CLL patients were isolated using ficoll (Pharmacia Biotech, Roosendaal, The Netherlands) and stored in liquid nitrogen. Expression of CD5 and CD19 (both Beckton Dickinson Biosciences [BD], San Jose, CA) on leukemic cells was assessed by flow cytometry (FACS Canto, BD) and analyzed with FACSDiva software (BD). All samples contained at least 90% CD5+/CD19+. More information on the characteristics of the CLL patients that provided material can be found in supplemental table 1.

Monocyte derived Mφs were obtained by differentiating monocytes isolated from healthy donor buffy coats after obtaining written informed consent. To this end, PBMCs were isolated using ficoll gradient purification according to manufacturer’s instructions (Lucron, Dieren, The Netherlands), after which monocytes were separated from peripheral blood lymphocytes using percoll gradient purification (GE healthcare, Milwaukee, USA). Next, monocytes were incubated to adhere at 37°C in 5% CO₂ for 40min at a concentration of 0.75*10⁶ cells/mL in 6-well plates (3mL) in IMDM/1% Fetal Bovine Serum (FBS, Invitrogen, Carlsbad, CA, USA) and washed to remove non-adherent cells. The monocytes were then differentiated to either M1 using 10ng/mL IFN-γ or M2 using 10ng/mL IL-4 (both R&D systems, Minneapolis, MN, USA) in IMDM supplemented with 10% FBS, 100u/mL Penicillin-100μg/mL Streptomycin (Life Technologies, Austin, TX, USA), 2mM L-glutamine (Life Technologies) and 0.00036% β-mercaptoethanol (Sigma, St. Louis, MO, USA) (IMDM+/-) for 72h.

Cell culture and co-culture experiments
NIH-3T3 mouse embryofibroblasts (3T3 cells) were supplied and characterized (for identity control, cytogenetics, and immunophenotype) by the Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ, Braunschweig, Germany). To mimic T cell induced CD40 signaling, these 3T3 cells were stably transfected with human CD40L (3T40 cells) as described previously8. When used as adherent feeder layer, fibroblasts were irradiated (30Gy) to stop proliferation before being seeded. Mφs were created as described under “CLL and healthy donor material and isolation”. After differentiation, Mφs were washed twice with IMDM+/-: CLL samples were thawed and diluted to a concentration of 1.5x10⁶ cells/mL before being plated on the respective feeder cells and co-cultured for indicated times. Trans-well experiments were performed according to manufacturer’s instructions using 0.4μm pore size 24-well culture plates (Corning, Corning, NY, USA).

Microarrays and bioinformatic analyses
CLL samples were stimulated with macrophages or not stimulated as described under “Cell culture and co-culture experiments”. Total RNA from these stimulated samples was prepared and microarray experiments were performed as they were for 3T40/non-stimulated samples as described before3. In short, RNA was isolated using TriReagent (Sigma) according to manufacturer’s instruction and RNA was further purified using the RNeasy micro kit (Qiagen, Valencia, CA, USA) according to manufacturer’s instruction including a DNAse (Qiagen) treatment. RNA was then hybridized on a U133plus2 microarray chip (Affymetrix, Santa Clara, CA, USA). The macrophage and 3T40 experiments were analyzed separately using Bioconductor packages in the statistical software package R
Measurement of levels of polysomal bound MCL-1 mRNA

40x10^6 CLL cells were co-cultured for 48h on Mφs or 3T40 cells or left unstimulated as described under “Cell culture and co-culture experiments”. Cells were then lysed in 1mL NP-40 buffer (0.5% NP40, 10mM TrisHCl pH 8.0, 140mM NaCl, 1.5mM MgCl2) with freshly added DTT (Sigma D0632 20mM), cycloheximide (Sigma C7698 150ng/mL), RNAsin (Promega N2515, 12μl/mL), and protease inhibitors (Calbiochem 539131 1:100). Lysates were loaded on linear 40%-15% sucrose gradients and centrifuged for 120min at 38000RPM. Gradients were subsequently separated in 18 fractions and each fraction was incubated for 30min at 37°C with 1% SDS, 10mM EDTA and 150μg/mL proteinase K (Roche), after which RNA was isolated from each fraction using phenol-chloroform extraction. To identify the presence of polysomes, 1μl of each fraction was run on a 2100 Bioanalyzer (Agilent technologies, Santa Clara, CA, USA). Non-polysomal and polysomal fractions were pooled and equal amounts of RNA were used for cDNA synthesis by reverse transcriptase reaction according to manufacturer’s instructions (Promega, Madison, USA). The cDNA was subsequently used as input for a real-time PCR using SYBR green (Life Technologies) reaction (40 cycles of 3sec at 95°C followed by 30sec at 60°C), using the following primers: MCL-1 5’-TCGTAAGGACAAACGGGAC-3’ and 5’-CATTCCTGATGCCACCTTCT-3’, 18S 5’-CGGCTACCACATCCAAGGAA-3’ and 5’-GCTGGAATTACCGCGGCT-3’. The abundance of each RNA was then calculated using the formula

\[
\% \text{ polysomal RNA} = \frac{\left(2^\Delta - (CtHF - CtLF)/(input HF/input LF)\right)}{\left(2^\Delta - (CtHF - CtLF)/(input HF/input LF)\right) + 1} \times 100\%
\]

where Ct = Ct value of HF = heavy (polysomal) fraction and LF = light (non-polysomal) fraction and input = the volume of RNA solution used for cDNA synthesis. The abundance of polysomal RNA was then plotted relative to the unstimulated condition.
Reagents
Recombinant human proteins were obtained from the following manufacturers: Basic fibroblast growth factor, Platelet-derived growth factor, Fms-related tyrosine kinase 3 ligand, and Vascular endothelial growth factor (all R&D systems), Epidermal growth factor (Sigma), Stem cell factor (Peprotech, Rocky Hill, NJ, USA), CCL3 (R&D systems), CCL5 (R&D systems), CCL23 (BioLegend, San Diego, CA, USA;)
The following inhibitors were used: mTOR inhibitors AZD8055 (Selleckchem, Houston, TX, USA) and rapamycin (Cell Signaling, Boston, MA, USA), PI3K inhibitor CAL101 (Selleckchem), caspase inhibitor Q-VD-OPh (Apexbio, Houston, TX, USA), GSK3 inhibitor CHIR99021 (Sigma), CCR1 inhibitor BX471 (Sigma), CCR2 inhibitor INCB3284 (Tocris Bioscience, Bristol, UK), CCR5 inhibitor Maraviroc (Apexbio).

Cell viability assays
To assess cell viability, 100μl of CLL cell suspension was incubated with 0.01μM Dihexyloxacarbocyanine Iodide (Dioc6, Molecular Probes) for 30-40min at 37°C. Prior to analysis, propidium iodide (PI, Sigma) was added (final concentration 2 μg/mL). Signals were measured on a FACS Calibur (BD) and analyzed using FlowJo software (TreeStar, San Carlos, CA, USA). Viable cells were defined as being Dioc6+/PI-.

Histology and immunofluorescence
Paraffin embedded tissue was obtained from our institute’s pathology department. Four-micron sections were de-waxed by immersion in xylene and hydrated by serial immersion in ethanol and PBS. Antigen retrieval was performed by heating sections for 20min in sodium citrate buffer (10mM sodium citrate, 0.05% Tween20, pH 6.0). Sections were washed with PBS (2 x 10min) and blocking buffer (TBS containing 10% BSA and 0.3% Triton X-100) was added for 1 hour. Sections were incubated with primary antibody, anti-CD20 (1:500, eBioscience, San Diego, CA, USA) and anti-MCL-1 (1:150, Abcam, Cambridge, MA, USA), or CD68 (Biolegend Y1-82A) in blocking buffer overnight at 4°C. Subsequently, the slides were washed with PBS (2 x 10min) and incubated with Alexa Fluor 488 labeled goat anti-mouse, Alexa Fluor 594 labeled goat anti-rabbit antibodies (1:400, Invitrogen) for 1 hour, after which the slides were stained for 10min with DAPI (0.1μg/mL in PBS). Alternatively, CD68 was developed using the VECTOR Blue Alkaline Phosphatase (AP) Substrate Kit (VectorLabs, Burlingame, CA, USA) and counterstained with Methyl Green. Sections were mounted with Fluoromount-G (eBioscience) and immunofluorescent imaging was performed using a Leica DMRA fluorescence microscope equipped with a cooled camera. Images were acquired using Image Pro Plus and composed in Adobe Photoshop CS5 (Adobe systems, San Jose, CA, USA).

NF-κB binding Enzyme-Linked Immuno Sorbent Assay (ELISA)
After washing CLL cells in ice-cold PBS, nuclear lysates were prepared using the NucBuster™ protein extraction kit (Millipore, Billerica, USA) according to manufacturer’s instructions. Protein concentration was measured with the BCA protein assay kit (Pierce, Thermo Fisher Scientific, Rockford, IL, USA) and 5μg of protein was subsequently used as input for the TransAM® NF-κB Family Transcription Factor Assay Kit, using provided antibodies for
p65 and p52 according to manufacturer's instructions. Signal intensity at 450nm was then determined by spectrophotometer (Biorad, Hercules, CA, USA).

**MCL-1 Knockdown**

CLL cells were transfected using the AMAXA nucleofection technology (Amaxa, Cologne, Germany) with one of 2 different MCL-1 siRNAs or control siRNA according to the manufacturer's instructions and as described in previous studies. In short, CLL cells were left to recover after thawing for at least 3h at 37°C in 5% CO₂. Cells (5.5x10⁶) were resuspended in 100 μl human B-Cell nucleofector kit solution (Lonza, Basel, Switzerland) and nucleofected with Silencer® Select siRNAs (final concentration 3μM) directed against MCL-1 or a negative control (Ambion, Paisley, UK; catalog numbers s8585 (2 CLL samples) and s8583 (3 CLL samples) for MCL-1 and AM4635 as negative control) using program X-001. After transfection, cells were directly resuspended in pre-warmed IMDM+/- and plated in 6-well plates to recover for at least 1h before commencing stimulation.

**Western blot and MCL-1 turnover analysis**

Sample preparation and western blot were performed as described before, using antibodies directed against MCL-1 (Cell Signaling 4572), BCL-X (BD 610211), BCL-2 (Alexis ALX-210-701-C1), BFL-1 (kind gift from Jannie Borst, Immunology Department, Dutch Cancer Institute, Amsterdam), NF-κB p65 (Santa Cruz 372), NF-κB p100/p52 (Cell Signaling 4882), phospho-4E-BP T37/46 (Cell Signaling 2855), phospho-elF4E S209 (Cell Signaling 9741), phospho-S6 S240/244 (Cell Signaling 5364), phospho-AKT S473 (Cell Signaling 4060), phospho-ERK (Cell Signaling 4370), phospho-GSK3B S9 (Santa Cruz 11757-R), and Actin (Santa Cruz 1616) and Histon H3 (Cell Signaling 9715). MCL-1 turnover time was determined by densitometric quantification of the MCL-1 western blot bands that were normalized to Actin levels at different time points. After calculating relative expression compared to T=0, exponential decay regression models were fitted using Prism 5.0 software (Graphpad, La Jolla, CA, USA). Presented images were composed in Adobe Photoshop CS5.

**Statistical analysis**

When comparing multiple groups, an analysis of variance (ANOVA) test with Dunnett's (comparing to control) or Newman-Keuls (comparing all groups) post-hoc tests were performed to test for significant differences between multiple groups. When comparing 2 groups, a Student's t-test was used. P values <.05 (*), <.01 (**) and <.001 (***) were considered statistically significant.

**ACKNOWLEDGEMENTS**

We would like to thank Richard Volckmann for his help with the analysis of the microarray data, Jannie Borst for providing the BFL-1 antibody, and Steven Pals for providing us with the CLL LN slides. A.P.K. is a Dutch Cancer Foundation fellow.
REFERENCES


6. Chapter 3 of this thesis.


46. Chapter 2 of this thesis.

Figure S1: Size marker indication for presented western blots. Molecular size markers are indicated for all western blots presented in the main figures.
**Figure S2:** Single channel images of figure 1d, right. Single channel images from the merged color image in figure 1d are presented for DAPI, CD20 and MCL-1.

**Figure S3:** AKT signaling is active in CLL LNs. Protein lysates from 3 CLL LN samples were analyzed by western blot for the phosphorylation of GSK3β. An unmatched unstimulated and 3T40-stimulated CLL sample were included for comparison. β-actin was used as a loading control.
**Figure S4:** No indications for transcriptional MCL-1 induction or post-translational stabilization. **a.** A representative Bioanalyzer profile for the polysome analysis presented in Figure 3c is shown for 18 sucrose gradient fractions, where left is the top fraction in the gradient. **b.** Expression data generated in Figure 2 was combined with the dataset from Pascutti et al. (3) and analyzed for differential expression of PIM1 and PIM2. Shown are mean ± s.e.m. for N=11 for ctr, N=13 for 3T40, N=3 for Mφ, and N=5 for Tact. **c.** Using the same dataset as in Figure S4b, the differential expression of anti-apoptotic BCL-2 family members was analyzed. Shown are mean ± s.e.m. for N=11 for ctr, N=13 for 3T40, N=3 for Mφ, and N=5 for Tact. **d.** Nuclei from the same co-cultured CLL samples used in Figure 1b were separated by centrifugation. Nuclear lysates were run on western blot and probed for nuclear translocation of canonical (p65) and non-canonical (p52) NF-κB subunits. Histone H3 was used as a nuclear loading control. **e.** Similarly as in panel d, nuclear lysates were prepared after 24h co-culture as in Figure 1 to detect the binding of activated NF-κB subunits p65 and p52 to consensus sequence oligonucleotides by using ELISA. Bars show mean ± s.e.m. for N=4 in 2 independent experiments for co-cultured samples (as indicated) or samples directly after thawing (T=0) (lower panel). **f.** Promoter sites of MCL-1 (Bcl2L3, NG_029146.1), BCL-XL (Bcl2L1, NG_029002.1), and BFL-1 (Bcl2A1, NG_029487.1) were retrieved from the PubMed RefSeq database by obtaining 5000 bases upstream of the transcription start site (TSS) of every gene. Each gene was then analyzed for the presence of the general NF-κB consensus sequence, (GGGRNNYYCC) using a Python 3 script. Each horizontal line represents 5000 bases upstream of the TSS and NF-κB binding sites are marked to scale.
Figure S5: Mφs induce CLL cell survival via a soluble factor. Mφs were seeded as in Figure 1a in the lower chambers of a trans-well culture system and CLL cells were cultured either in direct contact with the feeder cells (TW-), or in the upper chambers of the trans-well inserts (TW+) for 72h after which survival was measured as in Figure 1a. Shown are mean ± s.e.m. for N=4 CLL samples.
Supplementary script

```python
import os  # to scan all files in current directory
cons="GGGRNYYYCC"  # consensus sequence

def consscan(cons, promo):
    """scans text (promo) for presence of consensus and returns tuple of positions""
    lc,lp=len(cons),len(promo)
    return(i-lp for i in range(lp-lc,-1,-1) if patmatch(cons,promo[i:i+lc]))

def patmatch(seq1,seq2):
    """returns boolean whether seq1 matches to seq2. Seq1 also accepts wildcards as defined in md (match dictionary)"
    assert len(seq1)==len(seq2)
    md={"A":"A","C":"C","G":"G","T":"T","R":"AG","Y":"CT","W":"AT","S":"CG","M":"AC","K":"GT","B":"CGT","D":"AGT","H":"ACT","V":"ACG","N":"ATCG"}
    return all(j in md[i] for i,j in zip(seq1,seq2))

def scandir(cons):
    """prints all consensus sites (cons) in all .txt files in the current directory"
    for file in os.listdir("./"):
        if file.endswith(".txt"):
            with open(file) as fo:
                assert len(promo)!=0
                filename=fo.name
                sites=",".join(map(str,consscan(cons,promo)))
                print("Binding positions of {} in {}: {} sites if sites else "no match found")

if __name__ == '__main__':
    scandir(cons)
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