Typing of Lymphogranuloma Venereum Chlamydia trachomatis Strains


Published in:
Emerging Infectious Diseases

DOI:
10.3201/eid1611.100379

Citation for published version (APA):
Typing of Lymphogranuloma Venereum Chlamydia trachomatis Strains

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We analyzed by multilocus sequence typing 77 lymphogranuloma venereum Chlamydia trachomatis strains from men who have sex with men in Europe and the United States. Specimens from an outbreak in 2003 in Europe were monoclonal. In contrast, several strains were in the United States in the 1980s, including a variant from Europe.

Lymphogranuloma venereum (LGV) is a sexually transmitted disease caused by Chlamydia trachomatis serovars L1, L2, and L3. This disease is endemic to parts of Africa, Latin America, and Asia but only occurred sporadically in Europe before 2003, at which time an outbreak of LGV occurred in the Netherlands among men who have sex with men (MSM) (1). Since then, a large number of LGV cases have been reported among MSM in Europe, North America, and Australia. Although the inguinal form (formation of buboes) is more common in heterosexual LGV patients, in the current epidemic among MSM, anal infections have been diagnosed in most LGV cases.

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DOI: 10.3201/eid1611.100379

Sequencing the highly variable outer membrane protein A (ompA) gene identified a new genetic variant designated L2b, which subsequently has been identified in nearly all recent LGV cases in MSM that have been investigated (2). This variant was also found in isolates obtained from MSM in San Francisco, California, USA, in the early 1980s (3). However, differences in other regions of the C. trachomatis genome are possible and these regions should be investigated.

Multilocus sequence typing (MLST) is a genotyping method based on amplification and sequencing of several genetic regions. Recently, 3 MLST systems for genotyping of Chlamydiaceae bacteria have been reported. Two are based on housekeeping genes, and their resolution is comparable to that of ompA (4,5), which gives these 2 systems limited usefulness in C. trachomatis strain discrimination and outbreak investigations. A third MLST system was reported by Klint et al. (6) for short-term epidemiologic analysis of C. trachomatis strains. This system showed a 3-fold higher resolution than conventional ompA genotyping when applied to serovars A–K (6,7), which cause ocular trachoma and genital chlamydia infections. Recent evaluation of typing schemes confirms the considerable discriminatory potential of our system and recommends it for typing of closely related clinical strains (8). This system includes 5 highly variable gene regions; 2 of them (penicillin-binding protein and histone H1–like protein [hctB]), are subjected to selection pressure, which facilitates analysis of epidemiologic changes over limited periods.

Our objective was to deduce the nature and origin of the LGV outbreak among MSM in Europe. We used the MLST system of Klint et al. (6) to investigate genetic variation in LGV strains. Strains were obtained from MSM from contemporary Europe and the United States and from MSM in San Francisco during the 1980s.

The Study

All LGV specimens were obtained from MSM attending outpatient clinics. Twenty-two specimens were obtained in San Francisco during 1979–1985, and 5 specimens were obtained in the Baltimore, MD–Washington, DC, USA, area during 2007–2009. Fifty specimens were obtained from patients in Europe (Denmark [n = 7], France [n = 15], Germany [n = 1], the Netherlands [n = 9], Norway [n = 2], Spain [n = 7], and Sweden [n = 9]) during 2004–2008. DNA purification, PCR amplification of the ompA gene and the 5 specific highly variable gene regions (by using high-fidelity polymerase), DNA sequencing, and analysis of data (mean 5,972 nt/specimen, primers excluded) were performed as described (6,9). All novel mutations were reamplified and resequenced to confirm their authenticity.

All except 1 of the 50 specimens from Europe had an ompA genotype identical to the L2b reference strain L2b/
UCH-1/proctitis (AM884177.1). The nonidentical specimen came from Spain and had a previously unpublished single C→T point mutation in variable segment 2 at position 517 when compared with L2b/UCH-1/proctitis. This sequence of this specimen has been deposited in GenBank (accession no. GQ413955). All 50 specimens from Europe shared an identical MLST genotype (Table). The 5 specimens from the contemporary United States had the same ompA and MLST genotype as specimens from Europe (Table).

The 22 specimens from San Francisco were separated into 3 ompA genotypes and 5 MLST genotypes (Table). Six specimens had an ompA genotype identical to the L2 reference strain L2/434/Bu, and 7 had a novel genotype that differed from the L1 reference strain L1/440 by 9 point mutations. The remaining 9 specimens had an ompA genotype identical those of specimens from Europe and L2 reference strain L2b/UCH-1/proctitis. Three of these 9 specimens had MLST genotypes of strains from Europe, and the other 6 had a novel 108-nt deletion in the hctB gene region. This hctB variant has not been found in other specimens of C. trachomatis serovars A–L3 and is unique in the MLST database (http://mlstdb.bmc.uu.se/), which contains genetic profiles for 496 specimens (February 2010).

**Conclusions**

We aimed to deduce the nature and origin of the LGV outbreak among MSM in Europe. Because of the conserved nature of Chlamydia spp. genomes, high-resolution genotyping, rather than following evolutionary changes over long periods as with conventional MLST systems, was essential for investigating this outbreak. We found that the high-resolution MLST system described by Klint et al. (6) was most suitable for the current study.

**Table. Genetic profiles of lymphogranuloma venereum specimens, Europe and United States**

<table>
<thead>
<tr>
<th>Location</th>
<th>Sample years</th>
<th>No. specimens</th>
<th>hctB</th>
<th>CT058</th>
<th>CT144</th>
<th>CT172</th>
<th>pbpB</th>
<th>ompA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Europe†</td>
<td>2004–2009</td>
<td>49</td>
<td>27</td>
<td>13</td>
<td>17</td>
<td>13</td>
<td>29</td>
<td>28†</td>
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<tr>
<td>Europe§</td>
<td>2004–2009</td>
<td>1</td>
<td>27</td>
<td>13</td>
<td>17</td>
<td>13</td>
<td>29</td>
<td>39¶</td>
</tr>
<tr>
<td>USA#</td>
<td>2007–2009</td>
<td>5</td>
<td>27</td>
<td>13</td>
<td>17</td>
<td>13</td>
<td>29</td>
<td>28¶</td>
</tr>
<tr>
<td>USA**</td>
<td>1979–1985</td>
<td>3</td>
<td>27</td>
<td>13</td>
<td>17</td>
<td>13</td>
<td>29</td>
<td>28‡</td>
</tr>
<tr>
<td>USA**</td>
<td>1979–1985</td>
<td>6</td>
<td>44†</td>
<td>13</td>
<td>17</td>
<td>13</td>
<td>29</td>
<td>28‡</td>
</tr>
<tr>
<td>USA**</td>
<td>1979–1985</td>
<td>7</td>
<td>18</td>
<td>13</td>
<td>23</td>
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</tr>
<tr>
<td>USA**</td>
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<td>5</td>
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<td>13</td>
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<td>6</td>
<td>28</td>
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<tr>
<td>USA**</td>
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<td>37</td>
<td>19</td>
<td>6</td>
<td>28</td>
<td>22‡‡</td>
</tr>
</tbody>
</table>

**Note:**

- MLST, multilocus sequence typing; hctB, histone H1–like protein; pbpB, penicillin-binding protein; ompA, outer membrane protein A. Values are arbitrary designations referring to allele variants in our Chlamydia trachomatis MLST database (http://mlstdb.bmc.uu.se/). All MLST variants differ within regions with <5 point mutations unless otherwise indicated.
- †Denmark (n = 7), France (n = 15), Germany (n = 1), the Netherlands (n = 9), Norway (n = 2), Spain (n = 6), and Sweden (n = 9).
- *ompA variant 28 is identical to the reference strain L2b/UCH-1/proctitis (AM884177.1).§Spain.
- ‡ompA variant 39 contains a single point mutation compared with L2b/UCH-1/proctitis and has been deposited in GenBank under accession no. GQ413956.
- ††hctB variant 44 contains a novel 108-nt deletion and is unique in the MLST database.
- ¶ompA variant 40 contains 9 point mutations compared with reference strain L1/440 (DQ064294.1) and has been deposited in GenBank under accession no. GQ413956.
- §§ompA variant 22 is identical to reference strain L2/434/Bu (AM884176.1).
limitation of our study is that it does not include old LGV strains from Europe. However, after widespread investigations, we believe that no such strains are available.

Our study shows that the MLST system of Klint et al. (6) is suitable for epidemiologic analysis of C. trachomatis transmission, as indicated by our ability to differentiate the L2b ompA variant found in San Francisco into 2 strains. Investigation of additional LGV specimens from the United States and regions to which LGV is endemic with the MLST system could help determine a more detailed epidemiologic picture of the LGV outbreak in 2003 among MSM.

Acknowledgments

We thank Maria Blomqvist, Estrella Caballero, Vroni Girbinger, Thomas Morin, and Jolein Pleijster for assistance with laboratory work.

This study was supported by local funds from Uppsala University Hospital and was part of a collaboration with the European EpiGenChlamydia Consortium (www.EpiGenChlamydia.eu) (14), which is supported by the European Commission within the Sixth Framework Programme through contract LSHG-CT-2007-037637.

Mr Christerson is a PhD candidate in the Department of Clinical Microbiology at Uppsala University Hospital, Uppsala, Sweden. His primary research interests are development of genotyping methods and epidemiology of C. trachomatis.

References


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