Tumor cells can't stand the heat

Oei, A.L.

Citation for published version (APA):
Oei, A. L. (2017). Tumor cells can't stand the heat: Boosting the effectiveness of hyperthermia in cervical carcinoma
Chapter 7

Synthetic lethality *in vitro* and *in vivo* of hyperthermia, chemotherapy and PARP1-inhibition in BRCA-proficient tumor types


Under revision
Abstract

Background: Poly-(ADP-ribose)-polymerase1 (PARP1) is involved in repair of DNA single strand breaks. PARP1-inhibitors (PARP1-i) cause an accumulation of DNA double strand breaks, which are generally repaired by homologous recombination (HR). Therefore, cancer cells harboring HR deficiencies are exceptionally sensitive to PARP1-i. For patients with HR-proficient tumors, HR can be temporarily inhibited by hyperthermia, thereby inducing synthetic lethal conditions in every tumor type. Since cisplatin is successfully used combined with hyperthermia (thermochemotherapy), we investigated the effectiveness of combining PARP1-i with thermochemotherapy.

Methods: Survival of three HR-proficient cell lines after cisplatin, hyperthermia and/or PARP1-i was studied. Cell cycle analyses, quantification of γ-H2AX foci and apoptotic assays were performed to understand these survival data. The effects of treatments were further evaluated by monitoring tumor responses in an in vivo rat model.

Results: The in vitro data demonstrate a decrease in cell survival after addition of PARP1-i to thermochemotherapy, which can be explained by increased DNA damage induction. These in vitro findings are in line with in vivo model, in which a decreased tumor growth is observed upon addition of PARP1-i.

Conclusions: Our results in HR-proficient cell lines suggest that PARP1-i combined with thermochemotherapy can be a promising clinical approach for all tumors independent of HR status.
Introduction

Many cancer treatment modalities either interfere with DNA metabolism or with the repair of DNA damage. Combination of different, carefully selected modalities can enhance clinical outcomes. Here, we investigate the effectiveness of different combinations of three modalities which interfere with DNA integrity or DNA repair: cisplatin (cDDP), mild hyperthermia (heating the tumor to 40-42.5°C for 1 h) and Poly-(ADP-ribose)-polymerase1 inhibitors (PARP1-i).

A high percentage of breast and ovarian cancers (~50-85% and ~30-66%, respectively) are caused by mutations in BRCA1 or BRCA2 genes (Antoniou et al, 2000; Easton et al, 1995; Easton et al, 1997; Ford et al, 1994). Tumors with mutations in either of these genes require homologous recombination (HR) for repair (Bryant et al, 2005). Inactive HR can be due to mutations in BRCA1 or BRCA2, which may result in potentially lethal accumulation of DNA DSBs. HR-deficient (c.q. BRCA-deficient) cells are thus exquisitely sensitive to PARP1-inhibition (Hellday, 2011). Importantly, this also implies that healthy, HR-proficient cells are not targeted by PARP1-i, which makes this therapy particularly desirable for patients with HR-deficient tumors (Bryant et al, 2005; Girolimetti et al, 2014). Clinical trials have indeed confirmed the effectiveness of PARP1-i as a single treatment against BRCA-deficient tumors (Audeh et al, 2010; Huehls et al, 2012).

However, in HR-proficient tumors, synthetic lethality can also be induced by combining PARP1-i with a local treatment of mild hyperthermia (Bryant et al, 2005; Dedes et al, 2011; Eppink et al, 2012; Farmer et al, 2005; Hellday, 2011; Krawczyk et al, 2011; Schaaf et al, 2016; van den Tempel et al, 2016), which causes degradation of BRCA2 for several hours (Krawczyk et al, 2011) and HR deficiency at the heated tumor site. Combination of hyperthermia (HT) with PARP1-i thus creates a possibility to induce synthetic lethality in every tumor type that can be heated locally. Cisplatin (cDDP) is a widely used chemotherapeutic agent that is combined with HT (hence called thermochemotherapy) as standard treatment for previously irradiated patients with recurrent cervical cancer in the Netherlands (de Wit et al, 1999; Franckena et al, 2007; Rietbroek et al, 1997). cDDP induces DSBs that are usually repaired by HR and additionally interferes with non-homologous end joining (NHEJ), the other major DSB repair pathway (Poklar et al, 1996; Rudd et al, 1995). In absence of HR and NHEJ, a PARP1-dependent back-up NHEJ (b-NHEJ) pathway can take over the repair of DSBs (Audebert et al, 2004). As a consequence, a combination of HT, cDDP and PARP1-i could potentially cause an overload of DSBs while simultaneously interfering with all major DSB repair pathways (Metzger et al, 2013). The accumulation of unrepairred DSBs can result in cell death.
In this study, HR-proficient cell lines (R1, SiHa, HeLa) and a HR-proficient rhabdomyosarcoma allograft model were used to investigate the effectiveness of treatments combining PARP1-i, mild HT and cDDP. Cell survival as well as cell cycle analysis, quantification of γ-H2AX foci and apoptotic assays were performed. Finally, the effects of the different treatments on tumor outgrowth were measured *in vivo*.

**Results**

**Addition of PARP1-i to cDDP-based thermochemotherapy diminishes cell survival**

The response of cells to HT was investigated for BRCA2 levels (Figure 1A). After HT, BRCA2 levels are downregulated. To directly evaluate the effect of combinations of treatments with HT, cDDP and PARP1-i, clonogenic assays were conducted using the R1, SiHa and HeLa cells (Figure 1B). For all cell lines, therapy with the PARP1-i alone killed 30-40% of the cells. As such, treatment with PARP1-i was only slightly more effective than HT as a single treatment. cDDP was the most effective monotherapy. The combination treatment of PARP1-i with HT was equally effective as cDDP alone, and more effective than PARP1-i or HT alone. PARP1-i combined with cDDP was more effective than cDDP alone in the R1 cell line. In SiHa and HeLa cells, PARP1-i plus cDDP demonstrated a small decrease in cell survival, compared to cDDP alone. Combinational treatment of cDDP and HT was highly toxic and around 80-90% of the cells did not survive this treatment. The addition of PARP1-i to cDDP-based thermochemotherapy caused a higher than 2-fold reduction in cell survival in R1 cells, an almost 2-fold reduction in SiHa cells and a ~1.5-fold reduction in HeLa cells.

**Triple modality treatment leads to accumulation of DNA damage**

Formation of γ-H2AX, which represents unrepaired DSBs, was analyzed by flow cytometry, in order to identify a possible mechanism for differences in cell survival analyses after the triple modality treatment (Figure 2A). Cells grown on cover slips, treated with different combinations of cDDP, hyperthermia and PARP1-i were used for immunocytochemistry. For each condition one representative cell is depicted in Figure 2B. An up to 1.5-fold increase in γ-H2AX intensity was found after any of the single- and double-treatments. The load of DNA damage after addition of PARP1-i to cDDP-based thermochemotherapy was significantly higher than after cDDP-based thermochemotherapy alone.
Figure 1. Cell survival showing the effects of PARP1-i, HT and cDDP. (A) Western blots demonstrating R1, SiHa and HeLa cells are BRCA2-proficient. After HT, BRCA2 is downregulated. (B) Clonogenic assays were performed to study the effect of the different treatment combinations, 10-12 days after treatments. The addition of PARP1-i to cDDP-based thermochemotherapy resulted in a significantly lower cell survival compared to cDDP-based thermochemotherapy alone. R1: p=0.0008, SiHa: p=0.034, HeLa: p=0.021. The bar graph shows the mean of at least five independent experiments. From left to right: R1, SiHa, HeLa cells. * p<0.05, ** p<0.01, *** p<0.001.
Triple modality treatment increases the fraction of cells in S-phase

Cell cycle distribution was studied by incorporation of BrdU. In the untreated samples, ~50% of R1, SiHa and HeLa cells were in G₁-phase, ~40% in S-phase and ~10% in G₂-phase of the cell cycle (Figure 3). Treatment with PARP1-i caused modest changes in cell cycle distribution, while after HT a slight decrease in G₁ cells was observed, combined with a minor increase of cells in the G₂-phase. Of all monotherapies, cDDP had the strongest effects on cell cycle distribution in R1 cells, causing increased fraction of S-phase cells, as compared to the control. Moreover, in all cell lines, an increased proportion of cells in S-phase was found after a single treatment with cDDP and hyperthermia and after the triple modality.
Addition of PARP1-i to cDDP and hyperthermia increases apoptosis

To examine whether cell death detected in the clonogenic assays is due to induction of apoptosis, a Nicoletti assay was performed. The results of this assay demonstrate a 2 to 5-fold induction in apoptotic levels in R1 cells after any combination of treatment, except after the triple modality, for which there was an almost 9-fold induction of apoptosis (Figure 4). The degree of induction differed per cell line, but the trend was similar for all tested cell lines.

**Figure 3.** Cell cycle distribution using BrdU incorporation. While the monotherapies hardly ever changed the cell cycle analyses, any treatment containing cDDP resulted in a larger proportion of cells in S-phase. The differences found after adding PARP1-i to thermochemotherapy were not significant. From left to right: R1, SiHa, Hela cells.

**Figure 4.** Apoptotic levels measured using the Nicoletti assay. An ~1.5-fold induction of apoptosis is found after treatment with PARP1-i, cDDP and HT compare to cDDP with HT in R1 and SiHa cells (R1: p=0.0007, SiHa: p=0.0054). A smaller, though significant effect was found in HeLa cells after addition of a PARP1-i (HeLa: p=0.026). From left to right: R1, SiHa, Hela cells. * p<0.05, ** p<0.01, *** p<0.001.
The triple modality treatment delays tumor outgrowth in vivo

In vivo, tumors were treated with any combination of PARP1-i, HT and cDDP to test whether the therapy induces similar effects in vivo as in vitro. In untreated rats the tumor volume increased approximately exponentially, without a significant change in the slope of the growth curve. A treatment with cDDP alone, cDDP combined with either PARP1-i or HT, or cDDP combined with both induced a change in the steepness of the curve (Figure 5A), indicating that these treatments contributed to tumor control. Effects on tumor growth delay range from modest in the groups treated with PARP1-i, HT and HT+PARP1-i to substantial after cDDP alone, PARP1-i + cDDP, PARP1-i combined with HT and the triple modality. The differences in tumor growth were also calculated as the period of time needed to reach 10 times the tumor starting volume (T10 x SV) (Figure 5B). While the mono-therapies had only marginal influence on reaching T10 x SV, compared to the untreated animals, the double treatments did cause a delay of a few days. The highest level of tumor control was seen after the triple modality treatment, where the tumors took, on average, almost twice as long to reach the T10 x SV, as compared to the single therapies and approximately 1.5 times longer compared to cDDP-based thermochemotherapy. Importantly, no (acute) toxicities were detected in any of the rats.

Figure 5. In vivo experiments performed on rats. (A) Relative tumor volume demonstrated that PARP1-i enhanced the effectiveness of cDDP-based thermochemotherapy and reduced the in vivo tumor outgrowth. Arrows present the time of treatment. (B) Time needed to achieve ten times the start volume (T10 x SV). Animals treated with the triple modality showed a delay in reaching T10 x SV compared to thermochemotherapy or any single treatment. All animal experiments included at least five rats per group.
Discussion and conclusions

Our data demonstrate that addition of PARP1-i to cDDP-based thermochemotherapy results in an almost 2-fold reduction in cell survival, as compared to cDDP-based thermochemotherapy alone. This can be explained in part by an increased induction of apoptosis, which might be due to the higher DNA damage load, as indicated by γ-H2AX analysis. This, in turn, could result in increased activation of the $S/G_2$ checkpoint, as we observed a higher fraction of cells in S-phase after cDDP-based thermochemotherapy and an even more profound increase after the triple modality treatment. Prolonged cell cycle block could eventually lead to apoptosis. It is important to stress that all these effects were caused by a single treatment session and we expect that a repeated treatment regimen, more akin to clinical settings, would lead to stronger responses.

The experiments in rats demonstrate that the tumor growth got influenced after every additional treatment session of HT+PARP1-i+cDDP. After the last treatment the tumor growth increases. Although the double treatment with HT+cDDP and PARP1-i+cDDP are almost as effective in controlling tumor growth (Fig 5A), in terms of reaching ten times the start volume, the triple modality considerably delays tumor growth.

Clinical trials that have been performed on patients with BRCA-deficient tumors received daily Olaparib, sometimes even twice a day (Audeh et al., 2010). However, in our study, we mimic the BRCA-deficiency by applying HT. This results in a temporary downregulation of BRCA and within this timeframe, the PARP1-i should be administered. Therefore, the animals in our study were treated twice a week with two times Olaparib and one dose of cisplatin on the day of HT treatment. Thus, although it is not an identical schedule to the previous clinical trials with Olaparib, it is the most optimal scheme for treating BRCA-proficient tumors with HT and PARP1-i. As a consequence, since we have not treated daily, and only for a period of three weeks, it may be a valid consideration to extend the treatments with a couple of weeks, as HT sessions in patients are normally given once or twice a week for a period of five weeks in our center. Therefore, the outcomes of this study are promising, because we have treated these tumors with just 60% of the normal HT scheme and already the tumor shrank to its starting volume. However, an additional agent to standard thermochemotherapy may increase toxicity levels. Whether this occurs should be investigated.

There is growing evidence that platinum-based agents can also contribute in interfering with HR, resulting in HR-deficiency (McCabe et al., 2006). Subsequently, it might be interesting to combine platinum-based agents with PARP1-i. However, some people state that the use of PARP1-i may be limited in the clinic due to an increase in observed toxicities in combination with chemotherapeutic agents (Kummar et al., 2011; Rajan et al., 2012).
The triple modality that we propose, PARP1-i combined with cDDP-based thermochemotherapy might have more potential, because HT can make chemotherapeutic agents penetrate more deeply into the tumor and HT targets hypoxic areas, which are generally least sensitive to other anti-cancer therapies. Thus, lower chemotherapeutic dose may be as effective as normal concentrations without HT, but at a reduced toxicity (Oei et al, 2016).

In summary, there have been many studies that use the HR-deficiency in BRCA-mutated tumors to induce potentially lethal DNA damages by inhibition of PARP1. The advantage of our study is the possibility to treat BRCA-proficient cells with an already successful clinically applied therapy that can temporarily inhibit the HR. Therefore, we may induce synthetic lethality for any tumor type.

Acknowledgements
This work was supported by the Dutch Cancer Society (UVA 2012-5540).

Competing interests
Authors declare that they have no competing interests.
Material and Methods

Cell lines and cell culture. Rat rhabdomyosarcoma cells (R1, a cell line developed in our own institute for in vitro cultures as well as for in vivo growing tumors (Reinhold, 1965) were grown in MEM. The cervical carcinoma cells (SiHa and HeLa) were obtained from the American Type Culture Collection (ATCC) and grown in EMEM. All cell lines are BRCAwt. All media contained 25 mM Hepes (Gibco-BRL life technologies, Breda, The Netherlands) supplemented with 10% heat-inactivated foetal bovine serum (FBS) and 2 mM glutamine. Cells were maintained in a 37°C incubator with humidified air supplemented with 2% and 5% CO₂, respectively. The cell division time of R1 cells was approximately 16 h and of SiHa and HeLa cells 24 h.

In vitro treatments. Cells were treated for 1 h with 5 µM cisplatin (cDDP; Platosin®, Pharmachemie B.V., Haarlem, The Netherlands) and/or continuously with 100 µM of PARP1-i (dissolved in DMSO; NU1025, Tocris Bioscience, Bristol, UK). Cisplatin was added 5 minutes before HT, while PARP1-i was added 30 min prior to the HT treatment. Immediately after HT, medium was refreshed and PARP1-i was re-added until the end of the experiment. HT was performed by partially submerging the culture dishes in a thermostatically controlled water bath (Lauda aqualine AL12, Beun de Ronde, Abcoude, The Netherlands) for 1 h at 42°C. In order to check the temperature, thermocouples were placed in parallel culture dishes; the desired temperature (±0.1°C) was reached in approximately 5 min. HT was performed in an atmosphere containing 2% (R1 cells) or 5% (SiHa and HeLa cells) CO₂ with gas mixture inflow rate of 2 L/min.

Western blotting. To identify the BRCA2 status of these cell lines, western blots were conducted. Also the response to HT was tested. Cells were harvested 30 min post treatment. Next, pellets were lysed on ice in ice-cold RIPA buffer (20 mM Tris-HCl, 150 mM NaCl, 1 mM Na₂EDTA, 1% NP-40, 1% sodium deoxycholate, 2.5 mM sodium pyrophosphate, 1 mM beta-glycerophosphate, 1 mM Na₃VO₄, 1 µg/ml leupeptin) with protein inhibitors for 30 min (van Bree et al., 2004). Laemmli buffer containing fresh added 2-mercaptoethanol (355 mM) was mixed to the supernatant (1:1), before heated in boiling water for 2-5 min. Finally, samples were sonificated (Sonics & Materials Inc). One µg of protein was loaded on 4-15% SDS-PAGE gradient precast gels (BioRad) and transferred to PVDF membranes. Loading of protein were checked by Ponceau S staining. Immunodetection was performed for anti-BRCA2 (antibodies-online) and anti-ERK2 (Bethyl Laboratories), and a secondary anti-rabbit (Invitrogen Life Technologies). All samples were enhanced using chemoluminescence (Amersham Pharmacia Biotech). Eventually, blots were analyzed using LAS4000 (GE, Healthcare life sciences).

Cell survival assay. Cell survival was studied for R1, SiHa and HeLa cells using different combinations of HT, cDDP and PARP1-i. Clonogenic assays were conducted as described by Franken et al (Franken et al, 2006). Cells were plated 4 h prior to treatment into 6-well culture plates (Costar, USA). Dishes were placed in a 37°C incubator with the desired percentages of CO₂ at 37°C until sufficiently large clones were formed, which took approximately 10-12 days. Afterwards, the medium was removed and cells were washed with PBS before a 30-min incubation in a 2-3 ml of a mixture containing 6.0% glutaraldehyde and 0.5% crystal violet, at room temperature. Finally, plates were washed with tap water
and dried at room temperature. Colonies were counted under a light microscope (Bergs et al., 2006). Surviving fractions were calculated by dividing the plating efficiency of treated cells by that of control cells (van Bree et al., 2001).

**Detection of DNA DSBs quantifying γ-H2AX foci and flow cytometric detection of γ-H2AX.** To identify a possible mechanism for the differences in cell survival after the triple modality treatment, we quantified the formation of γ-H2AX, representing unrepaired DSBs, by flow cytometry. For immunocytochemistry, to present a representative image, cells were plated 24 h before any combination of cDDP, PARP1-i and HT treatment on sterile coverslips. On the next day, cells were fixed for 10 min in PBS containing 2% paraformaldehyde, three times washed with PBS. Then, cells were permeabilised during a 30-min incubation in TNBS (PBS containing 0.1% Triton X-100 and 1% FCS). Afterwards, cells were stained for 90 min at room temperature with 50 μl primary antibody of a mouse monoclonal anti-γ-H2AX (Millipore, dilution 1:100 in TNBS). Cells were washed once in PBS and two times in TNBS before staining for 30 min at room temperature with a secondary antibody anti-mouse Cy3 (Jackson, diluted 1:100 in TNBS). After three times washing with TNBS, vectashield-containing DAPI (Life technologies, USA) was dropped onto glass slides, before placing the coverslips upside-down on the slides. γ-H2AX foci were scored under a fluorescence microscope. To quantify γ-H2AX using flow cytometry, the staining was performed at 24 h after treatment. Cells were centrifuged (1200 rpm, 10 min), prior to fixation in a mixture containing 2 ml PBS and 6 ml absolute ethanol. Next day, cells were washed and incubated on ice for 10 min with PBS + 0.1% triton and 4% BSA. After centrifugation, cells were stained with antibodies against γ-H2AX-FITC (4 μg/ml, Merck Millipore, USA). Samples were then analyzed using FACS Canto (BD Biosciences, USA).

**Cell cycle analysis.** The thymidine analogue 5-Bromo-2'-deoxy-uridine (BrdU, Sigma Aldrich, USA) was used to analyze cell cycle distribution. At 16 h after treatments, BrdU (diluted 1:100) was added for 60 min at 37°C, before fixing cells in a mixture containing 2 ml PBS and 6 ml of absolute ethanol. Cells were stored at 4°C. The next day, after spinning down samples at 1200 rpm for 2 min, pellets were resuspended in pepsin-HCl (0.4 mg/ml, 0.1 N HCl) and cells were incubated for 30 min at room temperature. Cells were then washed with PBT (PBS containing 0.5% Tween-20 (Sigma Aldrich USA)). After centrifugation, pellets were resuspended in HCl (2 N, Merck) and incubated for 30 min at 37°C. Next, cells were stained with rat-anti-BrdU primary antibody (Abcam, UK) diluted 1:100 in PBTb (1% bovine serum albumin, Sigma, in PBT) for 60 min at 37°C. After washing cells two times with PBT and once with PBTg (PBT containing 1% normal goat serum (Dako, USA)), cells were stained with a goat-anti-rat secondary antibody labelled with FITC (Abcam, UK) diluted 1:100 in PBTg for 60 min at 37°C. Cells were washed three times with PBS before resuspending the pellets in PBS. Finally, propidium iodide (Sigma-Aldrich, USA) was added and cell suspensions were vortexed. Analysis was performed using the FACS Canto flow cytometer (BD Biosciences, US).

**Apoptosis assay.** To study apoptosis after any combination of treatments, the Nicoletti assay (Riccardi & Nicoletti, 2006) was performed. Cells were collected 48 h after the different treatments and pellets were resuspended in Nicoletti buffer (0.1% w/v Sodium citrate, 0.1% v/v Triton X-100 in demi water, pH 7.4), containing propidium iodide (Sigma-Aldrich, USA). Analyses were performed using the FACS Canto flow cytometer (BD Biosciences, USA).
Animal experiments. Female WAG-Rij rats were implanted with 1 mm³ piece of rhabdomyosarcoma tumor tissue in the right hind leg. After approximately three weeks, when the tumors had grown to 50-100 mm³, the animals were treated twice a week for a period of 3 weeks. The treatment involved HT for 1.5 h at 42°C in a thermostatically controlled water bath. In order to monitor the temperature thermocouples were placed next to the tumors. Earlier experiments (Krawczyk *et al.*, 2011; van Bree *et al.*, 1996) demonstrated that when the water bath was set to 42.7°C, the intra-tumor temperature was 42°C. Animals were cooled to prevent them from receiving whole body hyperthermia. The PARP1-i Olaparib was dissolved in a mixture of 10% 2-hydroxy-propyl-β-cyclodextrin and 10% DMSO in PBS and administrated at a dosage of 50 mg/kg p.o. 16 h and 2 h prior to HT. cDDP (Platosin®, Pharmachemie B.V., Haarlem, The Netherlands) was given i.p. at the dosage of 2.0 mg/kg 2 h prior to hyperthermia. Animal weight was monitored every two days and tumor sizes were measured using a Vernier caliper every two days, until the end of the experiment. Animals were sacrificed when the tumor volume exceeded 3500 mm³. The treated hind leg of rats was checked every two days for skin irritation. All animal experiments included at least five rats per group.

Statistical Analyses. All in vitro data represent means (of at least three experiments) with standard error of the mean (SEM) of at least three independent experiments. SPSS (Chicago, IL, USA) statistical software using a non-parametric Mann-Whitney test was used to analyze cell survival, γ-H2AX foci and apoptosis. P-values lower than 0.05 were considered statistically significant.


repair defect in BRCA mutant cells as a therapeutic strategy. Nature 434(7035): 917-21


Helleday T (2011) The underlying mechanism for the PARP and BRCA synthetic lethality: clearing up the misunderstandings. Molecular oncology 5(4): 387-93


