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Activation of phospholipase D by calmodulin antagonists and mastoparan in carnation petals

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Abstract
An in vivo assay for phospholipase D (PLD; EC 3.1.4.4) activity, based on its transphosphatidylation property, is described in detail and was used to study putative post-translational regulation mechanisms of PLD activity in carnation (Dianthus caryophyllus L.) petals. A variety of agents was applied to petal discs. The calmodulin (CaM) antagonists propranolol, N-(6-aminohexyl)-5-chloro-1-naphthalenesulphonamide (W7) and N-(6-aminohexyl)-1-naphthalenesulphonamide (W5), stimulated PLD activity in a dose-dependent manner. EGTA partially inhibited the stimulation by the CaM antagonists. Erythrosin B, an inhibitor of CaM-dependent P-type Ca²⁺-ATPases, slightly stimulated PLD activity. The results suggest that part of the stimulation of PLD activity by CaM antagonists is due to an increased intracellular Ca²⁺-concentration. PLD activity was stimulated by mastoparan in a dose- and time-dependent manner. The signal-like activation kinetics suggests that mastoparan activates PLD (in)directly via a G protein.

Key words: Phospholipase D, CaM antagonists, mastoparan, Dianthus, calcium.

Introduction
Phospholipase D hydrolyses the phosphate ester between the phosphatidate moiety and the head group of glycerophospholipids. The enzyme was first discovered in carrot roots and spinach leaves (Hanahan and Chaikov, 1947) and has since been shown to be widely distributed throughout the plant kingdom (Quarles and Dawson, 1969; Heller, 1978). Despite its high activity in plants, the physiological function of PLD in growth and development is still not clear. PLD could be involved in phospholipid turnover that maintains cell viability and homeostasis (reviewed by Dawidowicz, 1987). Additionally, the observation that the phospholipid content of petals, fruits and leaves decreased during senescence (McArthur et al., 1964; Ferguson and Simon, 1973; Beutelmann and Kende, 1977; Thompson et al., 1982) led to the hypothesis that PLD initiated this lipid breakdown (Brown et al., 1990; Paliyath and Droillard, 1992). The idea was partly based on changes in PLD activity that were measured using in vitro assays (Herman and Chrispeels, 1980; Borochov et al., 1982; Salama and Pearce, 1993), but recently, it was shown that regulation of PLD activity is more complicated. During castor bean germination and leaf senescence, not only the total PLD content was changed, but also three isoforms were differentially expressed (Dyer et al., 1994; Ryu and Wang, 1995). Furthermore, the intracellular distribution of PLD changed, with increasing activity becoming associated with its substrate in membranes (Xu et al., 1996). This suggests that post-translational mechanisms contribute to the regulation of PLD activity.

Post-translational regulation mechanisms apply to PLD involved in signalling. A recent study of the green alga Chlamydomonas illustrated that G-protein activation triggered immediate increases in PLD activity (Munnik et al., 1995), generating PtdOH which is a second messenger in animal cells (Liscovitch and Cantley, 1994, 1995), and which had dramatic effects on Chlamydomonas, causing deflagellation (Munnik et al., 1995).

In order to investigate the different functions of PLD in higher plants, activity must be measured in vivo, and in a preliminary study it was shown that this is possible with live carnation petals by using the transphosphatidyl-
ation activity of PLD (Munnik et al., 1995). Here, the assay conditions are described in full detail and data presented to show that PLD is activated by CaM antagonists and mastoparan in vivo. The results are discussed in relation to possible post-translational activation mechanisms.

Materials and methods

Plant material

Carnation (Dianthus caryophyllus L.) flowers cv. White Sim were obtained from a commercial grower and transported dry to the laboratory. Stems were re-cut to 20 cm and placed in deionized water under controlled environmental conditions (Woltering et al., 1993a). Just before use, discs (0.8 cm diameter, approximately 15 mg fresh weight) were cut from the middle part of the petals. Only the petals of the outer whorl were used.

Materials

[9,10 (n)-3H]Palmitic acid (40–60 Ci mmol-1) was obtained from Amersham (Buckinghamshire, UK). The analytical grade organic solvents and TLC silica 60 on plastic sheets were from Merck (Darmstadt, Germany). Phospholipid standards, phospholipase D (cabbage), erythrosin B, DL-propranolol, W5, W7, and mastoparan (Vespula lewisii) were purchased from Sigma (St Louis, MO, USA). Synthetic mastoparan and mastoparan (stock solutions of DL-propranolol, W5, W7, mastoparan, and W7) were obtained from Bachem (Bubendorf, Switzerland). A standard of PtdBut was prepared by conversion of egg-yolk PtdCho into PtdBut by phospholipase D in the presence of 5% (v/v) w-butanol (Comfurius and Zwaal, 1977).

Lipid labelling, extraction and analyses

Petals discs (a maximum of 10) were labelled with [3H]palmitic acid (10 μCi) in 2 ml labelling buffer (25 mM MES, 0.05% Tween 20, pH 5.5) by vacuum-infiltration (Munnik et al., 1994). After 5 min, the vacuum was released and the discs were incubated at room temperature for the times indicated. Incubations were stopped by immersing each petal disc in 600 μl of CHCl3:MeOH:HCI (100:100:1, by vol.) and freezing the samples in liquid nitrogen. Lipids were directly extracted after thawing and the petal discs were re-extracted with 450 μl of the same solution. Lipid fractions were pooled and a two-phase separation induced by the addition of 500 μl CHCl3 and 375 μl 0.9% (w/v) NaCl, I mM EGTA. The organic lower phase was washed once with 375 μl MeOH: H2O: HCI (50:50:1, by vol.). Lipid extracts were dried under a stream of nitrogen at 30°C and stored at -20°C until further use.

For TLC analyses, lipids were dissolved in CHCl3:MeOH (9:1, v/v) and samples applied to silica TLC plates which were then developed in the organic upper phase of ethyl acetate:isooctane:HAc:H2O (13:3:2:3:10, by vol.). Lipid spots were revealed with iodine vapour and identified by co-migration with unlabelled lipid standards. The RF values for the lipids were: PtdCho and PtdIns, 0.01; PtdEtn and PtdGro, 0.06; PtdOH, 0.24; monogalactosyl diacylglycerol, 0.44. Spots of interest were excised and their radioactivity was determined by liquid scintillation counting.

In vivo Phospholipase D assay

Separate petal discs were incubated at room temperature in 250 μl labelling buffer containing 3.5 μCi [3H]palmitic acid by vacuum-infiltration. After release of the vacuum, incubations were started by the simultaneous or successive addition of n-butanol (1 vol% in 250 μl of the labelling buffer) and the test reagents (stock solutions of DL-propranolol, W5, W7, mastoparan, and Mas17 in deionized water) as specified in the figure legends. Incubations were continued for the times indicated, the reactions quenched and the lipids extracted as described above.

A mixture of unlabelled PtdBut, PtdOH, and PtdCho in CHCl3:MeOH (9:1, v/v) was added to the dried samples. The total samples were chromatographed using three solvents consecutively, with intermediate drying. First, CHCl3 was used to reduce 3H-counts in the background due to the presence of unincorporated palmitic acid in the samples. Second, acetone was used to remove monogalactosyl diglyceride which otherwise co-migrated with PtdBut (RF value PtdBut, 0.43). Third, the TLC plate was developed using the ethyl acetate-isooctane solvent described above to separate PtdBut and PtdOH from the rest of the phospholipids.

PLD activity is presented as the formation of PtdBut which is expressed as the level of 3H-counts in PtdBut as a percentage of the label recovered from the structural phospholipids (SPL), PtdCho, PtdIns, PtdEtn, and PtdGro. The 3H-counts in the PtdBut fractions were corrected for those migrating at the position of PtdBut in control samples incubated without n-butanol. Lipid fractions isolated from petal discs incubated for 1 h with [3H]palmitate in the presence or absence of n-butanol (0.5%) contained on average (n=6–12, ± SE): SPL, 216 ± 24 000 dpm; PtdBut (not corrected), 1180 ± 120 dpm; background PtdBut, 470 ± 45 dpm.

Results

[3H]Palmitic acid labelling of lipids

Carnation petal discs were incubated with [3H]palmitic acid, their lipids extracted at regular intervals, and the lipids separated by TLC. After 24 h, about half of the 3H-label in the lipid extract was present as phospholipids. Most was incorporated into PtdCho plus PtdIns (56%) and PtdEtn plus PtdGro (42%). Approximately 1.3% of the label was recovered as PtdOH. The rest of the 3H-label was recovered as glycolipids (3%) or migrated nearer the solvent front, probably representing diacylglycerols, triacylglycerols and unincorporated palmitic acid.

Incorporation of 3H into phospholipids increased linearly during the first 8 h, usually reached a maximum within 24 h (Fig. 1A) and then remained at the same level up to 48 h, suggesting that isotopic equilibrium had been reached. The 3H-label was incorporated into PtdOH and the structural phospholipids in the same relative proportions during the whole labelling period (Fig. 1B), indicat-
In vivo PLD activity

Fig. 1. Incorporation of [3H]palmitate into the phospholipids of carnation petals. Petal discs were labelled with [3H]palmitate (5 μCi/ml) for increasing times up to 24 h. Lipids were extracted, separated by TLC and analysed as described in Materials and methods. (A) Data are expressed as the sum of the radioactivity in PtdCho, PtdIns, PtdEtn, and PtdGro. The experiment was repeated once with essentially similar results. (B) Data are expressed as the relative [3H]incorporation per phospholipid species, (●) PtdOH; (○) PtdCho plus PtdIns; (▲) PtdEtn plus PtdGro. The results are means±SE of 2–17 values from different experiments. PL, phospholipids.

Fig. 2. Formation of PtdBut in control (●) and propranolol-treated (○) petal discs as a function of the incubation time (A) and concentration (B) of n-butanol. Petal discs were incubated with [3H]palmitate (3.5 μCi/250 μl) and propranolol (5 mM) for 2 h (A) and 1 h (B). (A) n-Butanol (0.5%) was present during the last period of the incubation for the indicated times. (B) n-Butanol was present at the indicated concentrations for 1 h. The results are means±SE, n≥2 from different experiments. SPL, structural phospholipids.

The local anaesthetic propranolol, a CaM antagonist, (Volpi et al., 1981), was tested for its effect on in vivo PLD activity. When 0.5% n-butanol was added to petal discs pretreated with 5 mM propranolol, PtdBut formation was rapidly elevated above the control and increased for at least 60 min (Fig. 2A, open circles). The increase was linear with increasing n-butanol concentrations up to 2% (Fig. 2B). Membrane permeability was not adversely affected in as much as treatment with 4% n-butanol for 24 h did not increase ion leakage (conductivity: control petals, 0.42% h⁻¹, SE±0.02; n-butanol-treated petals, 0.48% h⁻¹, SE±0.07).

PtdBut formation was measured after 1 or 17 h of prelabelling with [3H]palmitate (see Fig. 1A for relative incorporation levels) using 0.5% n-butanol and 5 mM propranolol. After 1 h under both labelling conditions,
approximately 2.4% of the phospholipid label was converted into PtdBut. Therefore, in subsequent experiments, PLD activity was routinely measured using petal discs incubated with 0.5% n-butanol and $[^3]$H]palmitate during 1 h.

Addition of 5 mM propranolol to petal discs rapidly stimulated the formation of $[^3]$H]PtdBut and after 2 h of treatment $[^3]$H]PtdBut accumulated up to 10-fold (Fig. 3). The effect was dose-dependent and increased up to 10 mM (Fig. 4A). Higher concentrations of propranolol inhibited the incorporation of $[^3]$H]palmitate into the structural phospholipids (data not shown).

The naphthalenesulphonamides W7 and W5, two other CaM antagonists, also stimulated PLD activity in a dose-dependent manner (Fig. 4B). W7 (closed circles) was found to be more active than W5 (open circles), which correlated well with their order of potency (Hidaka and Tanaka, 1983). Higher concentrations of these CaM antagonists again inhibited the incorporation of label into the structural phospholipids.

Mastoparan stimulates PLD activity

The G protein activator mastoparan was used to get insight into the possible involvement of G proteins in the regulation of PLD activity in carnation petals. Mastoparan stimulated PLD activity in a concentration- and time-dependent manner (Fig. 5A, B). At 25 μM, activity was stimulated more than 5-fold. Mas17, a mastoparan analogue which is a far less potent activator of G proteins (Higashijima et al., 1990), did not significantly stimulate PLD activity (Fig. 5A). The effect of mastoparan was very rapid, half-maximal activity being reached within 5 min, producing kinetics of stimulation that were very different to those found for propranolol (Fig. 3).

Besides an increase in $[^3]$H]PtdBut formation, treatment of petal discs with either mastoparan or propranolol also resulted in a stimulation of the percentage of $[^3]$H-label in PtdOH. Interestingly though, mastoparan and propranolol differed in the ratio of PtdBut/PtdOH formation. To understand this point one must appreciate that the primary alcohol competes with water to accept the phosphatidyl group. Thus the PtdBut to PtdOH ratio is determined by the affinity of PLD for the acceptor and its local concentration. Even when stimulated, the PtdBut to PtdOH ratio is expected to be constant. Indeed, that was found for both stimulation by mastoparan and propranolol (Fig. 6). However, the PtdBut : PtdOH ratio for mastoparan was 1:5.5 (SE ± 0.1, n = 2, data were derived from two series of concentrations of mastoparan), while that for propranolol was 1:3.2 (SE ± 0.3, n = 8, data were derived from a series of concentrations of propranolol and kinetic experiments).
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Fig. 5. Mastoparan stimulates PLD activity dose- (A) and time-
dependent (B). Petal discs were incubated with [3H]palmitate and n-
butanol (0.5%) for 1 h. (A) Mastoparan (•) and Masl7 (○) were
present at the indicated concentrations for 1 h. The results are
means±SE, n≥2 from different experiments. (B) Mastoparan was
added during the last period of the incubation for the indicated times
Data are from one experiment except those from 0, 5 and 60 min which
are expressed as means±SE (n=9, 2 and 3, respectively). SPL,
structural phospholipids.

Discussion

It has been shown that phosphatidylation of primary
alcohols can be used as a measure of in vivo PLD in plant
tissues. The advantages of this assay are first that phos-
phatidylalcohols are stable products while PtdOH and,
for example, choline from PtdCho, are not. Second, PLD
activity can be determined in vivo because the formation
of phosphatidylalcohols is specific for PLD while that of PtdOH is not. That transphosphatidylation is
indeed a property of PLD was demonstrated by introdu-
cing a PLD gene from Ricinus communis into E. coli and
showing that only when the gene was expressed were
extracts of the bacterium able to hydrolyse phosphatidyl-
choline and transphosphatidylate ethanol (Wang et al.,
1994). Similar results have recently been obtained using
a human PLD gene expressed in insect cells (Hammond
et al., 1995).

In order to use the transphosphatidylation assay for
PLD activity, the phosphatidyl moiety of the PLD sub-
strate must be labelled. If the substrate has not been
determined, this means that all structural phospholip-
ids must be effectively labelled. Incubating discs in
[3H]palmitate achieves this goal, and in our system it was
incorporated into all major phospholipids at equal rates.
When 0.5% n-butanol was then added, the formation of
PtdBut increased linearly with time. While it is tempting
to use higher concentrations of n-butanol to facilitate
the detection of PtdBut, one should realize that alcohols can
themselves activate signalling enzymes such as PLC and
PLD (Hoek et al., 1992; Musgrave et al., 1992; Munnik
et al., 1995). Higher concentrations of n-butanol added
to the carnation petals certainly increased PtdBut forma-
tion, but whether this reflects PLD activation or just an
increase in substrate concentration was not tested.

The ability to measure PLD activity in vivo allowed the
study of some regulatory aspects of PLD. The CaM
antagonists W7, W5, and propranolol stimulated PLD
activity to a varying extent. PLD itself is not a CaM-
regulated enzyme, but CaM-dependent enzymes in plants
have been described. CaM antagonists have been shown
to raise cytosolic calcium levels in plant protoplasts
(Gilroy et al., 1987), perhaps by inhibiting CaM-depend-
dent P-type Ca2+-ATPases in the plasma- and ER mem-
branes (Dieter and Marme, 1980, 1981; Briskin, 1990;
Evans et al., 1991; Asklerlund and Evans, 1992). Since in
vitro PLD activity has been shown to be calcium-
dependent (Heller, 1978) and since the recently cloned
PLD genes from plants contain a potential calcium-

Fig. 6. Linear relationship between PtdBut- and PtdOH-formation.
PLD activity was stimulated by different concentrations of mastoparan
(0 to 50 µM) (•) for 1 h or by 5 mM propranolol (○) for different
times (0, 0.5, 1, 2 h). PtdOH formation was expressed as the level of
labelled PtdOH as a percentage of the sum of the radioactivity of the
structural phospholipids (SPL).
binding domain (Wang et al., 1994; Ueki et al., 1995), calcium could be a general trigger that activates PLD. Propranolol, W7 and W5 were effective at relatively high concentrations ranging from 0.5 to 5 mM, but their order of potency (W7 > propranolol ≈ W5) correlated with their IC50s for CaM-dependent phosphodiesterase activity (Volpi et al., 1981; Hidaka and Tanaka, 1983), suggesting that all three worked via CaM. The fact that EGTa (5 mM) partially inhibited propranolol-stimulated PLD activity (to 70% of the control) suggests that part of the effect of propranolol is due to an influx of extracellular Ca2+. Additionally, the significant stimulation of PLD by erythrosin B (50 μM, 1.6-fold increase), a specific inhibitor of P-type Ca2+-ATPases (Rasi-Caldogno et al., 1989; Askerlund and Evans, 1992), implicates calcium as the common denominator in all these responses. This is in agreement with a recent study where it is proposed that activation of PLD upon wounding of castor bean leaves is mediated by an increase in cytoplasmic Ca2+ (Ryu and Wang, 1996). Intracellular Ca2+-measurements will be needed to provide conclusive evidence about a possible role of Ca2+ in regulation of PLD activity. Besides specific effects of the CaM antagonists, aspecific effects such as perturbation of the membranes by these lipophilic molecules might contribute to a stimulation of PLD activity.

Mastoparan also stimulated PLD activity in carnation petals. It is thought to mimic the peptide domain of agonist-bound receptors that activates G proteins (reviewed by Ross and Higashijima, 1994). As such it could increase the intracellular Ca2+-concentration via a G protein activated Ca2+-channel or via PLC. There are several reports that mastoparan stimulates PLC activity in plants (Quarmby et al., 1992; Legendre et al., 1993; Drøbak and Watkins, 1994; Quarmby and Hartzell, 1994; Cho et al., 1995). By increasing the production of inositol-(1,4,5)trisphosphate, calcium can be released from intracellular stores and this could again stimulate PLD activity. Another possibility is that PLD is directly coupled to a G protein. This could account for the signal-like activation kinetics and is in agreement with the recent study of PLD activation in the alga Chlamydomonas (Munnik et al., 1995), where it was shown that treatments that are known to raise the calcium concentration and cause deflagellation, did not activate PLD, whereas mastoparan and other G protein activators did activate PLD. Lastly, mastoparan could directly activate PLD as was indicated recently for animal cells (Mizuno et al., 1995).

Both the kinetics of stimulation and the ratio of PtdBut:PtdOH formation for mastoparan and propranolol were different. The differences in kinetics could reflect different mechanisms of PLD stimulation or could be determined by different uptake rates or physical properties such as membrane affinity. The different ratios of PtdBut to PtdOH indicate that mastoparan and propranolol could stimulate or inhibit enzymes that affect PtdOH levels such as PLC in combination with diacylglycerol kinase, PtdOH kinase, and PtdOH phosphatase (Munnik et al., 1996). An alternative explanation is that mastoparan and propranolol activate two different isoforms. The idea of different PLD isoforms in higher plants has already been established in castor bean based on immunoblotting and activity assays (Dyer et al., 1994). In addition, three cDNAs have recently been identified in Arabidopsis (Dyer et al., 1995). The possibility that PLD isoforms can be differentially activated by CaM antagonists and mastoparan is new, exciting and deserves more attention.

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References


Drøbak BK, Watkins PAC. 1994. Inositol(1,4,5)trisphosphate production in plant cells: stimulation of venom peptides,
melittin and mastoparan. Biochemical and Biophysical Research Communications 205, 739–45.


