Spectral tuning, fluorescence and photoactivity in hybrids of photoactive Yellow Protein, Reconstituted with Native or Modified Chromophores
Kroon, A.; Hoff, W.D.; Fennema, H.P.M.; Gijzen, J.; Koomen, G.J.; Verhoeven, J.W.; Crielaard, W.; Hellingwerf, K.J.

Published in:
The Journal of Biological Chemistry

DOI:
10.1074/jbc.271.50.31949

Citation for published version (APA):
Spectral Tuning, Fluorescence, and Photoactivity in Hybrids of Photoactive Yellow Protein, Reconstituted with Native or Modified Chromophores*

(Received for publication, July 9, 1996, and in revised form, September 3, 1996)

Arthur R. Kroon‡, Wouter D. Hoff‡§, Herman P. M. Fennema¶, Jan W. Verhoeven¶, Wim Crielaard‡, and Klaas J. Hellingwerf‡

From the ‡Laboratory for Microbiology, E. C. Slater Institute, BioCentrum Amsterdam, Nieuwe Achtergracht 127, 1018 WS, Amsterdam and the ¶Department of Organic Chemistry, University of Amsterdam, Nieuwe Achtergracht 129, 1018 WS, Amsterdam, The Netherlands

Photoactive yellow proteins (PYPs) constitute a new class of eubacterial photoreceptors, containing a deprotonated thiol ester-linked 4-hydroxycinnamic acid chromophore. Interactions with the protein dramatically change the (photo)chemical properties of this cofactor. Here we describe the reconstitution of apoPYP with anhydrides of various chromophore analogues. The resulting hybrid PYPs, their acid-denatured states, and corresponding model compounds were characterized with respect to their absorption spectrum, pH for chromophore deprotonation, fluorescence quantum yield, and Stokes shift. Three factors contributing to the tuning of the absorption of the hybrid PYPs were quantified: (i) thiol ester bond formation, (ii) chromophore deprotonation, and (iii) specific chromophore-protein interactions. Analogues lacking the 4-hydroxy substituent lack both contributions (chromophore deprotonation and specific chromophore-protein interactions), confirming the importance of this substituent in optical tuning of PYP. Hydroxy and methoxy substituents in the 3- and/or 5-position do not disrupt strong interactions with the protein but increase their pH for protonation and the fluorescence quantum yield. Both deprotonation and binding to apoPYP strongly decrease the Stokes shift of chromophore fluorescence. Therefore, coupling of the chromophore to the apoprotein not only reduces the energy gap between its ground and excited state but also the extent of reorganization between these two states. Two of the PYP hybrids show photoactivity comparable with native PYP, although with retarded recovery of the initial state.

Many proteins bind cofactors to extend the physicochemical range of photoactivity, offered by their amino acid side chains. In photobiology these cofactors play a central role since they affect the flagellar rotation and thus leading to phototaxis. PYP is a water-soluble protein, which was first isolated from the halophilic purple phototrophic eubacterium *Ectothiorhodospira halophila* (1). Upon excitation, the protein enters a cyclic chain of dark reactions, i.e. a photocycle, resembling the one observed in the sensory rhodopsins from archaeabacteria (2). The first intermediate in this photocycle (red-shifted intermediate of PYP) is red-shifted to 465 nm. Subsequently, a blue-shifted intermediate (pBdark) develops on a sub-millisecond time scale. The photocycle is completed by the reformation of the initial state of the protein (pG), in about 1 s (3).

*E. halophila* displays negative phototaxis to blue light. PYP has been implicated to function as the photosensor in this response, since its absorption spectrum matches its wavelength dependency (4). Although PYP resembles the sensory rhodopsins both functionally and photochemically, its chromophore is not retinal but a novel type of chromophore, 4-hydroxycinnamic acid (5, 6), linked to Cys-69 via a thiol ester bond (7). Therefore, PYP represents a unique type of photoreceptor. PYP homologues have been detected in several eubacteria (8–11). Thus, it has been proposed to refer to this novel family of blue-light photoreceptors as Xanthopsins (11, 12).

The photochemical basis of the photocycle of PYP has recently been shown to reside in the photoisomerization of the vinyl double bond in the chromophore, from *trans* to *cis* (13). The quantum yield of this process was initially reported to be 0.64 (14); however, when measurements were extended to a large range of laser pulse energies, a value of 0.35 was obtained (15). Besides photoactivity, PYP shows weak fluorescence too. The quantum yield of pG is ~2 × 10⁻³, with an emission maximum at 495 nm (16, 17).

During progression through its photocycle PYP undergoes a large conformational change, which exposes hydrophobic sites to the solvent (14), due to a partial unfolding of the protein upon the formation of pB (18). The altered protein conformation of pB is thought to initiate signal transduction, ultimately affecting the flagellar rotation and thus leading to phototaxis.

* The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked “advertisement” in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

§ Present address: Dept. of Microbiology and Molecular Genetics, Health Science Center, University of Texas, Freeman Bldg., 6431 Fannin, Houston, TX 77030.

To whom correspondence should be addressed: Laboratory for Microbiology, E.C. Slater Institute, Nieuwe Achtergracht 127, NL-1018 WS Amsterdam, The Netherlands. Tel.: 31-20-5257055; Fax: 31-20-5257056; E-mail: a417hell@chem.uva.nl.

This paper is available on line at http://www-jbc.stanford.edu/jbc/

---

1 The abbreviations used are: PYP, photoactive yellow protein; HAP, histidine-tagged apoPYP overproduced in *E. coli*; pCA, 4-hydroxycinnamic acid (4-hydroxycinnamic acid); GFP, green fluorescent protein; GdnHCl, guanidinium hydrochloride; pG, Initial or groundstate of PYP and its hybrids; pB, blue-shifted intermediate of PYP; pBdark, Low pH-induced blue-shifted intermediate of PYP.
chromophore analogues. We refer to these proteins as hybrid produced HAP with its natural chromophore and a number of the chromophore (21), and (iii) specific protein-chromophore interaction to this shift have been identified (6, 7, 19): (i) formation of the coupling reaction between 4-methoxycinnamic acid and 4-nitrothiophenol. This latter anhydride has been obtained exclusively in crystalline form. 4-Aminocinnamic acid, 4-dimethylaminocinnamic acid, and 4-methoxycinnamic acid (3-methoxy, 4-hydroxycinnamic acid), and caffeic acid (3, 4-dihydroxycinnamic acid) were obtained from Sigma. 4-Aminocinnamic acid, 4-dimethylaminocinnamic acid, and 4-methoxycinnamic acid were obtained from Aldrich. All were obtained as the trans-isomer.

4-Methoxycinnamic acid anhydride was obtained as a by-product of the coupling reaction between 4-methoxycinnamic acid and 4-nitrophenol. This latter anhydride has been obtained exclusively in crystalline form.

Synthesis of Butyl Thiol Ester Model Compounds—We used the butyl thiol esters of I–IV as model compounds for the characterization of the absorption and fluorescence of hybrids I–IV. These model compounds were synthesized according to Ref. 25. Each carboxylic acid (1 mmol) was dissolved in dry N,N-dimethylformamide and 20 mg of dimethylaminopyridine, and 4 equivalents of isouethyl thiol was added. This mixture was used for the synthesis of the 4-hydroxycinnamic acid thiol ester, for which tertiary butyl thiol was used. Subsequently, 1.2 equivalents of dicyclohexylcarbodiimide was added. The reaction was carried out overnight, while slowly stirring at room temperature. The precipitated dicyclohexyl urea was filtered off repeatedly, and following concentration in vacuo, the filtrate was taken up in CH2Cl2. It was subsequently extracted two times with 0.5 M HCl, two times with a saturated NaHCO3 solution, and concentrated in vacuo. The product was purified by flash chromatography on silica gel (0.035–0.07 mm), using a petroleum ether (60–80 °C), ethyl acetate mixture (2:1, v/v). The purity of each product was checked by IR, NMR, and mass spectrometry.

Production of HAP—The polyhistidine-tagged apoPYP (HAP) was expressed in E. coli M15, containing pMph to express the synthetic pPYP (11). HAP production was induced during exponential growth in Luria Bertani medium in the presence of ampicillin (100 mg/liter), via addition of 0.2 mM isopropylthiogalactoside.

Purification and Reconstitution of HAP into PYP Hybrids—Two protocols were developed to obtain reconstituted PYP and its hybrids from HAP. Both protocols were carried out at room temperature. To investigate the kinetics of the reaction between HAP and the anhydride of pC, HAP was purified before its use as a substrate for the reconstitution reaction. E. coli M15/pPYP cells were harvested through centrifugation after 5 h of induction. The cell pellets were resuspended in 50 mM phosphate buffer (pH = 7.5), and cells were disrupted by sonication. The membrane and cytosolic fraction were separated by centrifugation (1 h at 250,000 × g) in a Centrifuge T-105 ultracentrifuge. The cytoplasmatic fraction, containing HAP, was dialyzed 16 h against 50 mM phosphate buffer (pH = 7.5) with two changes. Next, the extract was mixed with Ni2+-nitrilotriacetic acid resin, incubated for 1 h under gentle agitation at room temperature, loaded in a column, and washed with 0.2 M phosphate buffer (pH = 7.2) until the absorption at 280 nm (A280) was constant. The absorbed proteins were eluted with the same buffer, using a linear pH gradient from pH = 7.2 to pH = 3. Fractions with the highest A280 values were brought to neutral pH with sodium hydroxide, washed in an Amicon pressure filter concentrator, and stored at −20 °C or used immediately for reconstitution experiments. We observed that HAP has a tendency to precipitate more readily than holopPYP, especially at lower pH values.

Hybrids used for spectral characterization were isolated after reconstitution with the chromophore analogues listed in Fig. 2, because this considerably improved the yield in the purification procedure. Cell-free extract was prepared according to the following procedure. E. coli cells (from 20 liters of culture broth) were harvested by centrifugation and resuspended in 5.5 liters of 50 mM potassium phosphate buffer (pH = 7.2). The cells were prepared for lysis by 60 min incubation with 0.5 g of lysozyme (Sigma). Subsequently, the pH was increased to pH = 10 with 8 N NaOH, and cells were incubated for 5 min at this pH to reduce the viscosity of the lysate, after which the pH was re-adjusted to 7 with 37% (w/w) HCl. After removal of the debris by low speed centrifugation (30 min, 10,000 × g, room temperature). After binding, the resin was washed with 50 mM phosphate buffer (pH = 7.2) until the absorbance at 280 nm (A280) was constant. The supernatant was brought to 50% ammonium sulfate saturation and subjected to high speed centrifugation (30 min, 40,000 × g) to clarify the extract. The clear supernatant was distributed over 7 fractions (∼200 ml per fraction, each containing approximately 140 mg of HAP). These fractions were incubated for 60 min, each with an excess of one of the anhydrides of the chromophore analogues, under gentle mixing (2 mmol in total, i.e. a 200-fold molar excess over the two successive additions). The high concentration of proteins other than HAP, and of ammonium sulfate, in these samples did not significantly interfere with the rate or the final level of reconstitution with the anhydrides tested (data not shown). After reconstitution, the mixture was dialyzed overnight against 50 mM phosphate buffer (pH = 7.2) to remove excess anhydride and free acid and concentrated to a volume of 25 ml.

To purify the seven hybrid PYPs in parallel, we modified a rapid isolation procedure using Ni2+-nitrilotriacetic acid resin (27). Resin (0.3 ml bed volume) was incubated for 1 h with 1 ml of reconstituted extract, containing approximately 3 mg of hybrid PYP, under gentle agitation at room temperature. After binding, the resin was washed with 50 bed volumes 50 mM phosphate buffer (pH = 7.2). This was carried out by centrifuging each sample 15 times for 10 s in an Eppendorf centrifuge, followed by resuspension of the resin. The adsorbed proteins were eluted using 10 bed volumes 100 mM Na2-EDTA in 50 mM phosphate buffer (pH = 7.2). The eluates were washed with 25 mM phosphate buffer (pH = 7), concentrated to a volume of 1 ml (Centricon 10), and stored at −20 °C until further analysis. The purity of the samples was estimated by their purity index (pI) which has been defined as the ratio

of its absorption at 280 and at 446 nm (1). The pI of the sample reconstituted with the native chromophore was 0.9 (see Fig. 3). Thus this sample is approximately 60% pure.

Absorption and Fluorescence Spectroscopy—To monitor the reconstitution reaction between HAP and the anhydride of pCA, quartz cuvettes with two compartments, separated by a transparent barrier, were used to measure sum spectra of the apoprotein and the anhydrides in solution, before and after mixing (Hellma Benelux V.V., Rijswijk, The Netherlands). Electronic absorption spectra were recorded in an SLM-Instruments Aminco DW2000 spectrophotometer with a resolution of approximately 1 nm. Fluorescence spectra were recorded in a SPEX-Fluorolog 2 spectrometer. Quantum yields were measured relative to quinine sulfate (dissolved in 1 mM H$_2$SO$_4$, $d = 0.55$ (27)).

Photoactivity was analyzed by measuring the electronic absorption spectrum of the PYP hybrids in 50 mM sodium phosphate buffer, pH = 7 (or 50 mM glycine buffer, pH = 10, for hybrid IV), on a Hewlett-Packard 8453A diode array spectrometer, modified to allow illumination of the sample with a 200-watt high pressure mercury arc lamp, at right angles to the measuring beam, with a time resolution of approximately 0.1 s. All spectroscopy was carried out at room temperature.

Miscellaneous—HAP in complex samples was quantified with rocket immunoelectrophoresis (11). pK$_v$ values of model compounds and hybrids were determined through manual fits of the data of spectrophotometric titrations, recorded with a Cary-3 UV/Vis spectrophotometer (Varian).

RESULTS AND DISCUSSION

Purification of Polyhistidine-tagged ApoPYP and Its Reconstitution with Various Chromophores—HAP was isolated from the cytoplasmic fraction of recombinant E. coli by Ni$^{2+}$-affinity chromatography and reconstituted into holoPYP with the anhydride of pCA. This process was monitored by UV/Vis absorption spectroscopy after mixing 4 mM pCA anhydride with 10 $\mu$M HAP (Fig. 1). At neutral pH values the rate of the trans-esterification reaction is beyond the time resolution of a conventional spectrophotometer, as was found for the reconstitution of apoPYP from E. halophila with this anhydride (24). However, lowering the pH to 4.8 slows down the rate of the reconstitution reaction (see Fig. 1), allowing its kinetic analysis. The increase in absorption at 446 nm indicates the formation of holoPYP, while the decrease in absorption at 365 nm is caused by the decomposition of pCA anhydride. The reconstitution reaction is complicated by the fact that pCA anhydride not only reacts with HAP, to form holoPYP, but also with water, yielding pCA. This can account for the absence of an isosbestic point in the difference spectra, recorded during the reconstitution reaction (see Fig. 1). The rate of both reactions increases with increasing pH. For the reaction between the anhydride and HAP this can be explained by assuming that Cys-69, binding the chromophore, reacts with the anhydride in its ionized form, which is expected to be formed with a $pK$ of approximately 10. The spontaneous hydrolysis of the anhydride in water presumably is base-catalyzed.

The progress of the reaction as monitored by the absorption changes at 446 nm can best be described as a second-order reaction, with a specific rate constant of the reaction between HAP and 4-hydroxycinnamic acid anhydride of 0.16 M$^{-1}$s$^{-1}$ at pH = 4.8 (see Fig. 1, inset). More detailed investigations of this reaction requires stopped-flow analysis.

From the pI of the reconstituted protein, it can be concluded that the extent of reconstitution is more than 95% of the amount of HAP available (data not shown). The absorption spectra of native PYP, isolated from E. halophila, and of reconstituted HAP are indistinguishable, including the absorption maximum at 446 nm and a characteristic fine-structure at 318 nm. From this, it can be concluded that HAP is an excellent substrate for reconstitution. HAP is overproduced at a level of approximately 50 mg per liter of culture per OD unit at 660 nm (see Ref. 11) and therefore now available for biophysical and structural studies in large amounts. This also renders PYP accessible to studies by site-directed mutagenesis, which are in progress in our group, and it opens the way to reconstitute apoPYP with pCA analogues.

To assess the feasibility of this latter option, we synthesized the anhydrides of six PYP analogues (Fig. 2). In the first three analogues the aromatic ring of cinnamic acid carries one (II, III) or two (IV) additional substituents at the 3- and/or 5-position. In the remaining three analogues (V–VII) the phenolic hydroxy group is lacking, being substituted by an amino, a methoxy, and a dimethyl amino group, respectively. These chromophore analogues were chosen (i) to test the effect of perturbation of the chromophore binding pocket, by the added...
substituents, on the absorption and fluorescence characteristics of PYP, and (ii) to investigate the proposed importance of the 4-OH group for spectral tuning of the chromophore. Like the reconstitution with the anhydride of pCA, the reaction of the other chromophore analogues with HAP at neutral pH also proceeds beyond the time resolution of the spectrophotometer used (data not shown). This indicates that even in IV, which carries two bulky methoxy substituents flanking the 4-hydroxy group, no compelling steric impediments exist to fitting the (trans)-cinnamyl chromophore unit into its binding site in the apoprotein. This result shows that the reaction between apoPYP and anhydrides can be used as a general method to obtain hybrid PYPs.

In the study of photoactive proteins, the use of modified chromophores has proven to be a very powerful strategy. Besides the ones used in this study, several additional chromophore analogues may further help elucidating the mechanism of functioning of PYP. Some examples are as follows: (i) replacement of the vinyl bond by either a single or a triple bond, (ii) isotopically labeled derivatives, and (iii) chromophore analogues in which the vinyl bond is locked through a covalent bridge (compare Ref. 28).

**UV/Vis Absorption Characteristics of Hybrid PYPs**—To investigate the mechanism of spectral tuning in PYP, we aimed at the separation of the three factors involved in this process (see Introduction) in the seven hybrids and their quantification (in cm$^{-1}$). In this report we will refer to the spectral shift induced by the formation of the thiol ester bond for the uncharged chromophore as $\Delta l_{\text{thiol est}}$, to the shift caused by the deprotonation of the thiol ester-bound chromophore as $\Delta l_{\text{deprot}}$, and to the shift caused by specific interactions between the chromophore and the protein as $\Delta l_{\text{protein}}$. The sum of these three factors is referred to as $\Delta l_{\text{tot}}$.

The $\Delta l_{\text{tot}}$ of the hybrids was determined by measuring the absorption spectra of these proteins, after their purification, and of the corresponding chromophore-derived model compounds (Fig. 3 and Table I). The values for $\Delta l_{\text{tot}}$ were calculated, using the absorption maxima observed for the hybrid PYPs and the free acids at pH = 7, with the exception of the PYP hybrid containing IV, which displayed maximal spectral tuning at pH values above 9 (see below).

The two classes of chromophores clearly lead to very different magnitudes of spectral tuning. For hybrids II to IV, $\Delta l_{\text{tot}}$ was approximately 13,000 cm$^{-1}$, essentially identical to the value observed in PYP, containing its native chromophore (Table II). However, for V to VII, $\Delta l_{\text{tot}}$ is reduced to approximately 7,100 cm$^{-1}$, confirming the importance of the 4-OH group in the process of the tuning of the chromophore absorption. In line with this, hybrids reconstituted from analogues lacking the 4-hydroxy group show pH-independent spectral characteristics in the pH range investigated (Table I). Nevertheless, also these latter hybrids (i.e. containing a class B chromophore), all display an absorption band of the aromatic amino acids of the apoprotein and a clearly discernible absorption band caused by the chromophore. The absorption maxima of this latter band range from 355 (V) to 436 nm (VI) in class B hybrids.

To quantify the contribution of $\Delta l_{\text{thiol est}}$, the absorption spectra of the hybrids, denatured by 4 M guanidinium HCl (GdnHCl) at pH = 7, where the chromophore is present in its neutral form, were compared with those recorded at pH = 11. At the latter pH, the chromophores containing an OH group are present in their anionic form (see below; note that denaturation of PYP at pH = 13 yielded almost identical absorption maxima) (Fig. 3). Comparison of the absorption maxima of the free acids and the GdnHCl-denatured hybrid PYPs at pH = 7 (and 3) allowed the calculation of $\Delta l_{\text{thiol est}}$. For PYP and all hybrids, the $\Delta l_{\text{thiol est}}$ was approximately 6,000 cm$^{-1}$, whereas $\Delta l_{\text{deprot}}$ was around 4700 cm$^{-1}$ for I–IV and absent for V–VII (see Table I).

This analysis indicates (see Table II) that approximately 80% of $\Delta l_{\text{tot}}$ can be explained on the basis of two chemical modifications of the chromophore upon binding to the apoprotein, i.e. formation of the thiol ester bond and the deprotonation. This leaves approximately 20% of the shift to be caused by $\Delta l_{\text{protein}}$. This effect is largest for the native chromophore (i.e. 3,000 cm$^{-1}$; see Table I) but still significantly present in II–IV (approximately 2,000 cm$^{-1}$). Apparently, the modifications in these latter three class A chromophores leave the specific protein-chromophore interactions, which give rise to $\Delta l_{\text{protein}}$, largely intact. However, in the chromophores that lack the 4-OH group (V–VII), the effect of $\Delta l_{\text{protein}}$ is absent. Apparently, this phenolic hydroxy group does not only contribute to the spectral tuning by its deprotonation but is also essential for the interactions with the protein, leading to $\Delta l_{\text{protein}}$.

Below pH = 3, PYP is converted to a blue-shifted state (1), which can be regarded as the acid-denatured state of PYP. This state, called pB$_{\text{dark}}$, has an absorption spectrum, similar to the blue-shifted intermediate pB from the photocycle of PYP, but is slightly red-shifted with respect to the absorption expected (and observed) for thiol ester-linked pCA (19). We have also examined the presence of a red-shift in this latter intermediate (pB$_{\text{dark}}$) in the hybrids studied here (Tables I and II) and found that for II–IV this parameter is approximately 720 cm$^{-1}$, slightly lower but similar to the value observed for native
Photoactive Yellow Protein Containing Modified Chromophores

Summary of spectroscopic data of cinnamic acid derivatives, their respective anhydrides, and the hybrid PYPs

The \( \lambda_{max} \) of the free acid, the anhydride, and the PYP-linked chromophore, at various pH values and in the presence and absence of guanidinium hydrochloride, has been indicated. The asterisk indicates that this sample was measured at pH 11 rather than at pH 7. ND, not determined; GdnHCl, guanidinium hydrochloride.

<table>
<thead>
<tr>
<th>Hybrid PYP in buffer</th>
<th>Hybrid PYP in 4 M GdnHCl</th>
<th>Free acid</th>
<th>Anhydridea</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \lambda_{max} )</td>
<td>( \epsilon )</td>
<td>fwhm(^a)</td>
<td>( \Delta \lambda )</td>
</tr>
<tr>
<td>( \text{nm} )</td>
<td>( \text{m}^{-1}\text{cm}^{-1} )</td>
<td>( \Delta \text{cm}^{-1} )</td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>350</td>
<td>446</td>
<td>394</td>
</tr>
<tr>
<td>II</td>
<td>361</td>
<td>457</td>
<td>417</td>
</tr>
<tr>
<td>III</td>
<td>359</td>
<td>460</td>
<td>417</td>
</tr>
<tr>
<td>IV</td>
<td>357</td>
<td>488*</td>
<td>444</td>
</tr>
<tr>
<td>V</td>
<td>353</td>
<td>353</td>
<td>353</td>
</tr>
<tr>
<td>VI</td>
<td>436</td>
<td>436</td>
<td>436</td>
</tr>
<tr>
<td>VII</td>
<td>ND</td>
<td>355</td>
<td>ND</td>
</tr>
</tbody>
</table>

\(^{a}\) Determined from absorbance difference spectra.

<table>
<thead>
<tr>
<th>Hybrid</th>
<th>Absorption characteristics</th>
<th>Tuning parameters</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \lambda_{max} )</td>
<td>( \epsilon )</td>
<td>( fwhm(^a) )</td>
</tr>
<tr>
<td>( \text{nm} )</td>
<td>( \text{m}^{-1}\text{cm}^{-1} )</td>
<td>( \Delta \text{cm}^{-1} )</td>
</tr>
<tr>
<td>I</td>
<td>22,421</td>
<td>4.55 * 10(^4)</td>
</tr>
<tr>
<td>II</td>
<td>21,882</td>
<td>2 * 10(^4)</td>
</tr>
<tr>
<td>III</td>
<td>21,739</td>
<td>3 * 10(^4)</td>
</tr>
<tr>
<td>IV</td>
<td>20,491</td>
<td>2 * 10(^4)</td>
</tr>
</tbody>
</table>

\(^{a}\) fwhm, full-width at half-maximum.

PYP (927 cm\(^{-1}\)), whereas it is absent in V and VI. This indicates that (i) also in pB\(_{dark}\) a protein-induced bathochromic shift occurs in I–IV and that (ii) like \( \Delta \lambda_{protein} \), this shift depends on the presence of the 4-OH group. Apparently, also in pB\(_{dark}\), the chromophore absorption is red-shifted by specific interactions with the protein. It is interesting to note that in hybrids II–IV, both \( \Delta \lambda_{protein} \) and \( \Delta \lambda_{pB_{dark}} \) are reduced to approximately 75% of the value observed in native PYP.

In addition to \( \Delta \lambda_{tot} \), we also examined the bandwidth and maximal extinction coefficient \( \epsilon \) of hybrids II–IV and compared it to native PYP (Table II). Assuming identical levels of reconstitution and purity for HAP reconstituted with I–IV, \( \epsilon \) is reduced from 45.5 to approximately 20–30 m\(^{-1}\) cm\(^{-1}\) in II–IV (Table II). Thus, compared with PYP, the other hybrids have a lower \( \epsilon \). However, since they also have a larger bandwidth at half-height (see Table II), this probably only has a minor effect on the optical transition probability in hybrids II and III.

Protein-induced Shifts in the pK for Chromophore Deprotonation—As described above, deprotonation of the phenolic hydroxy group of I–IV leads to a significant bathochromatic shift in the absorption spectrum. The pK for deprotonation of the thiol ester model compounds of I–IV is approximately 8.7 (Table III). Therefore, chromophore deprotonation at neutral pH can only occur if the protein environment lowers the pK for this process. For native PYP it has been reported that a \( \Delta pK \) occurs from 8.8 to 2.8.\(^6\) It has been proposed that Glu-46 and Arg-52 are important factors in causing this \( \Delta pK \) (see below).

To further examine the effect of the chromophore modifications in II–IV, we measured the pK for chromophore deprotonation in these hybrids (Table III). It is clear that the pK in hybrids II and III is also strongly affected by the protein, be it somewhat less than in native PYP (shifting it to 3.8 and 3.5, respectively). However, in hybrid IV the pK of the chromophore is not at all affected by binding to the protein. This suggests that in this hybrid the phenolate anion at neutral pH is proto-nated, possibly because steric hindrance by the two methoxy groups prevents Arg-52 from approaching the chromophore homologue sufficiently to allow it to charge-compensate a phenolate anion. It is interesting to note that hybrid IV is the only one in which the absorption maxima observed at pH = 13 and = 11 in GdnHCl are not indistinguishable. This supports the notion that the electrostatic interactions in this hybrid are significantly different from those in native PYP, apparently leading to an increased resistance of the protein toward alkaline denaturation. Since it has been proposed that this interaction is important for the stability of PYP, it will be of interest to investigate how the temperature stability of hybrids I–IV correlates with these pK values.

From Table II it can be seen that the \( \Delta \lambda_{tot} \) in hybrid IV (at high pH) is still quite significant and almost indistinguishable from the corresponding values of II and III, whereas its \( \Delta pK \) has been reduced to 0. Apparently, the \( \Delta pK \) and \( \Delta \lambda_{tot} \) are not directly correlated.

Fluorescence of Hybrids I–IV—To complement the data obtained by absorption spectroscopy described above, we further probed the transition between pG and its excited state pG* by studying the fluorescence characteristics of hybrids I–IV. We focused on two parameters, the fluorescence quantum yield (\( \Phi_{f} \)) and the Stokes shift, as calculated from the difference

<table>
<thead>
<tr>
<th>Hybrid PYPs</th>
<th>Model thiol esters</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>2.8</td>
</tr>
<tr>
<td>II</td>
<td>3.8</td>
</tr>
<tr>
<td>III</td>
<td>3.5</td>
</tr>
<tr>
<td>IV</td>
<td>8.7</td>
</tr>
</tbody>
</table>

For further details: see “Materials and Methods.”
Photoactive Yellow Protein Containing Modified Chromophores

Fluorescence characteristics of PYP, three of its hybrids, and their corresponding model thiol esters

| Hybrid PYPs | pH = 2 | | | pH = 7 | | | Butyl thioesters | pH = 7 | | | pH = 11 |
|-------------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|--------|
| λ<sub>ex</sub> | λ<sub>em</sub> | St. | Φ | λ<sub>ex</sub> | λ<sub>em</sub> | St. | Φ | λ<sub>ex</sub> | λ<sub>em</sub> | St. | Φ |
| nm | nm | % | cm<sup>-1</sup> | nm | nm | % | cm<sup>-1</sup> | nm | nm | % | cm<sup>-1</sup> |
| I | 360 | 439 | 4999 | 0.3 | 446 | 494 | 2173 | 0.2 | 340 | 460 | 7673 | 0.1 |
| II | 360 | 458 | 5944 | 0.4 | 450 | 527 | 2761 | 1.9 | 349 | 489 | 8203 | 0.03 |
| III | 358 | 471 | 6702 | 0.9 | 461 | 527 | 2717 | 5.4 | 344 | 490 | 8662 | 0.3 |
| IV | 372 | 482 | 6135 | 0.7 | 485 | 548 | 2371 | 2.7 | 345 | 505 | 9184 | 0.1 |

Readings were taken at pH = 11 for hybrid IV.

between the maxima of chromophore fluorescence emission and excitation (Table IV). Comparison with Table I shows that a strict correlation exists between the position of the absorption maxima and the maxima of fluorescence excitation. The only exception is hybrid IV at low pH. This is related to the difficulty of accurate determination of the absorption maximum at this pH, which may be affected by light scattering. Also, increased absorption of the apoprotein, due to tyrosine deprotonation, may complicate this measurement. The quantum yield determined for I, i.e. 2 × 10<sup>-3</sup>, agrees reasonably well with previously determined values (16, 17).

Examination of fluorescence emission and excitation spectra (as an example, the spectra of IV are displayed in Fig. 4, for both the pG and pB<sub>dark</sub> state) shows that the Stokes shift in hybrids II–IV is approximately 2600 cm<sup>-1</sup>, similar to the value of 2179 cm<sup>-1</sup> observed in I (Table IV). However, their Φ<sub>fl</sub> is significantly increased, up to 5 × 10<sup>-2</sup> for II. To examine whether the origin of this increased Φ<sub>fl</sub> is intrinsic to the chromophore analogues used, or to protein-chromophore interactions, we also determined the fluorescence spectra and Φ<sub>fl</sub> for the thiol ester model compounds I–IV. At pH 11, the Stokes shift of these compounds (in their deprotonated form) is approximately 5900 cm<sup>-1</sup>, significantly larger than in the PYP hybrids. This may be caused by a significantly impaired flexibility of the chromophore in the apoprotein environment. However, the Φ<sub>fl</sub> of model compounds II–IV was approximately 2 × 10<sup>-3</sup>, comparable with the low value observed in native pG. Apparently, interaction of chromophores II–IV with the apoprotein increases their Φ<sub>fl</sub> 5–25-fold (Table IV). In parallel to the increased quantum yields, it was observed that also the fluorescence lifetime of II and III is strongly increased with respect to I. The multi-exponential nature of these lifetimes, however, complicates their straightforward analysis.

To further probe the protein-chromophore interactions in pB<sub>dark</sub>, alluded to above, we also determined the fluorescence characteristics of hybrids I–IV at pH 2 and compared these results with the corresponding protonated model compounds (at pH 7). This analysis shows that I–IV display fluorescence not only after excitation of their pG form but also of the pB<sub>dark</sub> state (Table IV). As for the pG state, it was found that the Stokes shifts of the chromophores in pB<sub>dark</sub> (approximately 5900 cm<sup>-1</sup>) are significantly smaller than those observed for the protonated model compounds, measured at pH 7 (approximately 8400 cm<sup>-1</sup>). For hybrid I the Φ<sub>fl</sub> is higher in pB<sub>dark</sub> than in pG, whereas for hybrids II–IV the opposite is observed; the fluorescence in II–IV is much less affected by the protein in its pB<sub>dark</sub> state than in its pG state. This suggests that in hybrids II–IV the protein-chromophore interactions in pB<sub>dark</sub> are less perturbed by the additional ring substituents than those in pG.

Spectral Tuning of PYP in Its pG State—To find possible mechanisms involved in Δλ<sub>protein</sub> it is useful to compare spectral tuning in PYP and in the rhodopsins, since its physical basis has been studied extensively in the latter proteins. Two chemical modifications occur upon binding of retinal to opsins, both of which are analogous to those occurring in PYP: (i) the formation of a Schiff base between a Lys residue and retinal, and (ii) the protonation of this Schiff base, aided by a strong protein-induced increase in its pK. These two processes shift λ<sub>max</sub> from 370 to 440 nm but are usually not considered to be part of the opsin shift. Specific protein-chromophore interactions lead to a further bathochromic shift from 440 to 568 nm (called opsin shift (29)) for the light-adapted state of bacteriorhodopsin. This shift of 5,100 cm<sup>-1</sup> corresponds to the Δλ<sub>protein</sub>
of approximately 3,000 cm\(^{-1}\) in hybrid I. Although the opsin shift in bacteriorhodopsin is still only partially understood, two contributing factors involved are generally accepted (30–32). First, the protonated Schiff base is weakly stabilized (the so-called external point charge model (29)) by a complex countereion, involving the charges of Asp-85, Asp-212, and Arg-82. Second, the protein forces the retinal ring into the 6-\(\text{trans}\)-conformation, thus leading to a co-planarization of the \(\beta\)-ionone ring and the polycene chain of retinal, thereby extending the conjugated system (30). Recently, a third factor has been identified, the stabilization of the excited state of retinal by polar or polarizable side chains in the retinal binding pocket (31).

Since the negative charge on the deprotonated PYP chromophore(s) is buried within the protein (22), the presence of a counterion for this charge at a relatively large distance can be proposed to explain the \(\Delta\lambda\)\(_{\text{protein}}\). Arg-52 is a likely candidate to contribute to this function. Our analysis indicates that the 4-OH group is essential for \(\Delta\lambda\)\(_{\text{protein}}\). Second, the counterpart of the 6-\(\text{trans}\)-conformation in retinal bound to native bacteriorhodopsin is formed by the conformation around the C–C single bond of the \(-\text{C}=\text{C}(\text{C}(-\text{S}(-\text{O})\text{-})\text{-O}\) fragment. Both the 1.4-Å x-ray data (22) and resonance Raman data (21) indicate that this single bond is in the \(s\)-cis conformation. Since it seems reasonable to expect that for model compounds in solution this bond is in the \(s\)-\(\text{trans}\) conformation, this factor may affect the absorption spectrum of the chromophore. However, at this point it is difficult to quantitate this effect.

A third possibility that we have considered is protein-induced torsional strain on the \(\text{trans}\) C–C bond in the chromophore. Such strain would destabilize the ground state and would stabilize the excited state, since this latter state is expected to have an energy minimum at a double bond angle of 135°. Initial results of essential dynamics calculations\(^3\) suggest that such strain indeed is present in the pG state. This proposal implies that the degree of torsional strain imposed on the chromophore is decreased by the additional ring substituents in II–IV. Four residues in the pCA binding site of PYP are of primary importance: (i) Arg-52, which has already been discussed, and (ii) the hydrogen bonding network between the phenolate anion and Glu-46, Tyr-42, and Thr-50. These interactions can be expected to reduce the mobility of the chromophore, which may be a prerequisite for the application of torsional stress on the C–C bond. In this way, this proposal can explain the absence of a \(\Delta\lambda\)\(_{\text{protein}}\) in V–VII.

Binding of a chromophore to apoPYP leads to a decrease in the magnitude of its Stokes shift by more than a factor of 2. It is interesting to note that PYP displays both the strongest decrease in Stokes shift and the largest \(\Delta\lambda\)\(_{\text{protein}}\). This decrease in Stokes shift is not easily compatible with the notion of torsional stress; however, torsional stress would tend to decrease the difference between the ground state and the first excited state.

**Photoactivity of the Hybrid PYPs**—The typical photobleaching and dark recovery of native PYP (and of I) is readily observable in hybrids II and III but not in IV. The rate of the dark recovery reaction, however, was slightly (in III) and even strongly (in II) decreased. For the other hybrids (in particular IV) transient kinetic analyses will have to be applied, to determine whether or not a short-lived intermediate exists.

**Comparison of PYP Hybrids with Green Fluorescent Protein (GFP)**—An interesting comparison can be made between the highly fluorescent hybrids (III and IV; see Table IV) and GFP, based on their mutual similarity and differences. At neutral pH, for instance, IV and GFP display two absorption bands (for the latter at approximately 395 and 475 nm). However, whereas in IV these two forms are due to the titration of the phenolate anion, in GFP they appear to be due to isomerization (33), although some interconversion of the absorption bands at 395 and 475 nm occurs, in the pH range in which GFP can be titrated, without interference by rapid denaturation (34). Surprisingly, fluorescence emission of GFP from both its absorption bands (data not shown) gives rise only to a single emission band (at 508 nm), with a only minor red shoulder. This is in striking contrast to PYP, in which both protonation states of the apoprotein-bound chromophore give rise to a fluorescent protein, with slightly higher fluorescence quantum yield at neutral pH but with clearly separated emission bands for the two states. This can be concluded from comparisons of the fluorescence of pG and pB\(_{\text{dark}}\). It is not known what the effect is of isomerization of the chromophore of PYP on the fluorescence emission. The transient character of pB complicates its fluorescence characterization. In view of the intriguing fluorescence characteristics of GFP, it will be of great interest to characterize the isomerization state of the chromophore of GFP with IR spectroscopy, for example, as a function of irradiation dose and pH. The fluorescence characteristics of GFP may be due to emission from a deprotonated chromophore.

The increase in \(\Phi\)\(_{\text{f}}\) of hybrids II and III and their photochemical activity provides new approaches for future work. First, time-resolved fluorescence spectroscopy can be employed as a new and powerful tool to investigate the primary photochemistry of PYP. Second, the photochemical properties of PYP can be manipulated by developing different chromophore analogues, strongly enhancing the scope for application of PYP hybrids in practical applications, for example, like in optical data storage. In addition, the biophysical basis for the distribution of the quantum yield of each of the three parallel reaction pathways, available to the excited state of pG (i.e. photochemistry, fluorescence, and radiationless decay (see Ref. 15), becomes accessible to experiments. This may lead to better insight in the way nature tunes chromophores, to function optimally as photosensory light absorber (PYP) or bioluminescent light emitter (GFP).

**Concluding Remarks**—In this study we present a general method for the reconstitution of PYP with various chromophores, based on the ability to reconstitute apoPYP with 4-hydroxyemminic acid anhydride (24) and the heterologous overexpression of HAP (11). The N-terminal histidine tag of the overproduced protein allows for its efficient purification and if necessary can be specifically removed via enterokinase digestion of the isolated protein. This makes PYP available in large amounts and amenable to site-directed mutagenesis and labeling with NMR-visible isotopes (e.g. with \(^{15}\)N and \(^{13}\)C).

Here we have characterized seven PYP hybrids and have found that the protein dramatically changes chemical and physical properties of the various chromophores. Hybrids II–IV, which have additional ring substituents, still show a strong chromophore-protein interaction, as is apparent from the change in (i) wavelength of maximal absorption of the chromophore, (ii) \(pK_a\) for deprotonation, (iii) fluorescence quantum yield, and (iv) Stokes shift. For all hybrids studied here, a quantitative description of the absorption changes between the free acid and the chromophore bound to the native protein was obtained, in which the formation of the thiol ester accounts for a red-shift of \(-6000\) cm\(^{-1}\) (\(\Delta\lambda_{\text{thiol ester}}\)), chromophore deprotonation accounts for \(-4700\) cm\(^{-1}\) (\(\Delta\lambda_{\text{deprot}}\)), and further protein-chromophore interactions for \(-2300\) cm\(^{-1}\) (\(\Delta\lambda_{\text{protein}}\)). Our data are consistent with the presence of torsional stress on the vinyl double bond of the chromophore, but also the conformation of

---

\(^3\) D. M. F. Van Aalten, W. D. Hoff, J. B. C. Findlay, and K. J. Hellingwerf, submitted for publication.
the single bond in the –C≡C–(–S–)=O fragment may be of importance for ΔG_protein.

REFERENCES